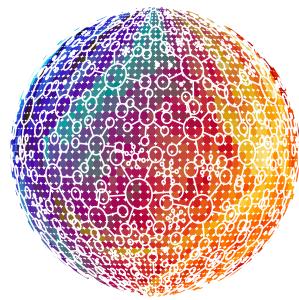

Graphein - Supplementary material

Graphein



Contents

A Featurisation Schemes for Protein Structure Graphs	4
B Featurisation Schemes for RNA Structure Graphs	5
C Featurisation Schemes for Molecular Graphs	6
D Parameters for protein-protein interaction graphs	7
D.1 General parameters	7
D.2 BioGRID	7
D.3 STRING	9
E Parameters for gene regulatory networks	9
E.1 General parameters	9
F Graph Construction Performance	9
F.1 C α Graphs	10
F.1.1 Peptide Bonds	10
F.1.2 Distance-based	10
F.1.3 Intramolecular	10
F.2 Atomic Graphs	10
G Societal Impact	11

A Featurisation Schemes for Protein Structure Graphs

Table 1: Geometric representation options for a protein structure

<i>Node Features</i>		
Node Type	Feature	Source
Residue		
	Molecular Weight	
	x, y, z Co-ordinates	
	Sidechain Vector	
	Neighbour Vectors	
	β Carbon Vectors	
	ϕ Torsion Angle	DSSP [1]
	ψ Torsion Angle	DSSP [1]
	Secondary Structure	DSSP [1]
	Solvent Accessibility	DDSP [1]
	Low-dimensional embeddings of physicochemical properties	Meiler et al. [2]
	ExPaSy Protein Scale	[3]
	AAIndex Descriptors (various)	[4]
	ESM Transformer Protein Language Model Embedding	ESM [5]
	BioVec Protein Language Model Embedding	ProtVec [6]
Atom		
	Atomic Weight	
	Covalent Radius	[7]
	H-bond Donor Status	
	H-bond Acceptor Status	
<i>Edge Types</i>		
Node Type	Edge Type	Source
Atom	Covalent Bonds	
Residue		
	Hydrophobic Interactions	
	Disulfide Interactions	
	Hydrogen Bonds	
	Ionic Interactions	
	Aromatic Interactions	
	Aromatic-Sulphur Interactions	
	Cation- π Interactions	
	Peptide Bonds	
	π Stacking Interactions	[8]
	Salt Bridge	[8]
	t Stacking	[8]
	Van der Waals	[8]
Any		
	K-Nearest Neighbours	
	Delaunay Triangulation	
	Distance Threshold	
	Distance Window	
	Sequence Distance	
<i>Graph-level Features</i>		
	Features	Source
Sequence		
	Molecular Weight	
	Transformer Positional Encoding	
	ESM Transformer Protein Language Model Embedding	ESM [5]
	BioVec Protein Language Model Embedding	ProtVec [6]
	Amino Acid Composition	ProPy [9]
	Dipeptide Composition	ProPy [9]
	Tripeptide Composition	ProPy [9]
	Moreau-Broto Autocorrelation	ProPy [9]
	Moran Autocorrelation	ProPy [9]
	Geary Autocorrelation	ProPy [9]
	Sequence-order-coupling Number	ProPy [9]
	Quasi-Sequence-Order Descriptors	ProPy [9]
	CTD Descriptors	ProPy [9]

B Featurisation Schemes for RNA Structure Graphs

Table 2: Geometric representation options for a RNA structure

<i>Node Features</i>		
Node Type	Feature	Source
Residue	Molecular Weight	
	x, y, z Co-ordinates	
Atom	Atomic Weight	
	Covalent Radius	[7]
<i>Edge Types</i>		
Node Type	Edge Type	Source
Atom	Covalent Bonds	
	K-Nearest Neighbours	
Base	Delaunay Triangulation	
	Distance Threshold	
	Distance Window	
	Sequence Distance	
	Phosphodiester Bonds	
	Base Pairing Interactions	
Sequence	Pseudoknots	
	Molecular Weight	
<i>Graph-level Features</i>		
Features	Source	
Sequence	Transformer Positional Encoding	

C Featurisation Schemes for Molecular Graphs

Table 3: Geometric representation options for a molecular graph

Node Type	<i>Node Features</i> Feature	Source
Atom/Junction Tree		
	Atomic Mass	RDKit
	Covalent Radius	
	Atom Type	
	Covalent Degree	
	Total Valence	RDKit
	Explicit Valence	RDKit
	Implicit Valence	RDKit
	Implicit Hydrogens	RDKit
	Explicit Hydrogens	RDKit
	Total Hydrogens	RDKit
	Radical Electrons	RDKit
	Formal Charge	RDKit
	Hybridization	RDKit
	Aromatic Status	RDKit
	Ring Status	RDKit
	Ring Size X Status	
	Isotope Status	RDKit
	Formal Charge	RDKit
	Chiral Status	RDKit
Edge Types		
Node Type	Edge Type	Source
Atom/Junction Tree		
	Covalent Bonds	
	K-Nearest Neighbours	
	Delaunay Triangulation	
	Distance Threshold	
	Distance Window	
	Fully Connected	
Edge Features		
Feature		Source
	Bond Order	RDKit
	Aromatic Status	RDKit
	Conjugation Status	RDKit
	Ring Status	RDKit
	Ring Size X Status	
	Stereo Configuration	RDKit
Graph-level Features		
Features		Source
	ChEMBL Metadata	[10]
	Molecular Weight	RDKit
	Geometric Center	RDKit
	Principal Moments of Inertia	
	Max Ring Size	
	Morgan Fingerprint	RDKit
	Fragment Counts	
	QED Score	RDKit

D Parameters for protein-protein interaction graphs

D.1 General parameters

The following table shows the generic Graphein parameters for protein-protein interaction graphs.

Parameter	Default	Valid values	Description
protein_list	—	protein IDs (list of string)	Proteins to include in the graph.
ncbi_taxon_id	9606 (human)	integer	NCBI taxon identifier.
sources	all sources	'biogrid', 'string'	List of sources (databases) to retrieve the data from.
paginate	True	True, False	Whether to paginate the API calls for the sources that require it.

D.2 BioGRID

The following table shows the Graphein parameters for BioGRID. See also the BioGRID API.

Parameter	Default	Valid values	Description
searchNames	True	True, False	If True, the interactor OFFICIAL_SYMBOL will be examined for a match with the protein list.
searchIds	True	True, False	If True, the interactor ENTREZ_GENE, ORDERED_LOCUS and SYSTEMATIC_NAME (orf) will be examined for a match with the protein list.
searchSynonyms	True	True, False	If True, the interactor SYNONYMS will be examined for a match with the protein list.
searchBiogridIds	True	True, False	If True, the entries in the protein list will be compared to BIOGRID internal IDS which are provided in all Tab2 formatted files.
additionalIdentifierTypes	empty	string	Identifier types on this list are examined for a match with the protein list.
max	10000	integer	Number of results to fetch. Used for pagination.

interSpeciesExcluded	True	True, False	If True, interactions with interactors from different species will be excluded.
selfInteractionsExcluded	False	True, False	If True, interactions with one interactor will be excluded.
evidenceList	empty	Pipe-separated list of evidence codes from here	Any interaction evidence with its Experimental System in the list will be excluded from the results unless includeEvidence is set to true.
includeEvidence	False	True, False	If set to true, any interaction evidence with its Experimental System in the evidenceList will be included in the result
excludeGenes	False	True, False	If true, interactions containing genes in the input list will be excluded from the results.
includeInteractors	True	True, False	If true, in addition to interactions between genes on the input list, interactions will also be fetched which have only one interactor on the input list.
includeInteractorInteractions	False	True, False	If true, interactions between the input list's first order interactors will be included.
pubmedList	empty	string	Interactions will be fetched whose Pubmed Id is/ is not in this list, depending on the value of excludePubmeds.
excludePubmeds	False	True, False	If False, interactions with Pubmed ID in pubmedList will be included in the results; if 'true' they will be excluded.
htpThreshold	20	integer	Interactions whose Pubmed ID has more than this number of interactions will be excluded from the results. Ignored if excludePubmeds is False.

throughputTag	'any'	'low', 'high', 'any'	If set to 'low' or 'high', only interactions with 'Low throughput' or 'High throughput' in the 'throughput' field will be returned.
---------------	-------	----------------------	---

D.3 STRING

The following table shows the Graphein parameters for STRING. See also the STRING API.

Parameter	Default	Valid values	Description
network_type	'functional'	'functional', 'physical'	Network type: functional (default), physical.
add_nodes	0	integer	Adds a number of proteins to the network based on their confidence score, e.g., extends the interaction neighborhood of selected proteins to desired value.
show_query_node_labels	False	True or False	When available use submitted names in the preferredName column.

E Parameters for gene regulatory networks

E.1 General parameters

We download gene regulatory networks from TRRUST and RegNetwork. We build a directed graph where nodes are genes and attributed edges represent regulatory effects (activation, repression, or unknown).

Parameter	Default	Valid values	Description
gene_list	—	gene symbols (list of string)	Genes to include in the graph.

F Graph Construction Performance

We provide performance assessment performed on an AMD EPYC 7742 64-Core Processor. We use 20 PDBs (number os residues in brackets) with a mean length of 1,089.1 residues: 1n9u (10), 1j51 (30), 1ip0 (50), 1eod (100), 1agy (200), 1wzu (300), 1inp (400), 1a8h (500), 2ywe (600), 6cgm (700), 7lst (800), 4a7k (900), 4nab (1,000), 6mfw (1,210), 7edd (1519), 4f93 (1724), 6lqa (2059), 6r9t (2547), 4rh7 (3005), 5w1r (4128).

We observe construction time is marginal compared to download time for coarsened structural representations (Figure 1).

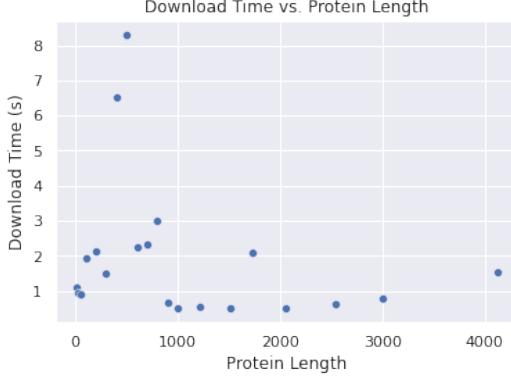


Figure 1: Download times for protein structures from the PDB.

F.1 C_α Graphs

F.1.1 Peptide Bonds

We compute peptide bond graphs for the above list of 20 structures 50 times (i.e. 1,000 evaluations) in parallel (16 workers) and average across 7 runs to obtain an average construction time of 0.0071s per protein.

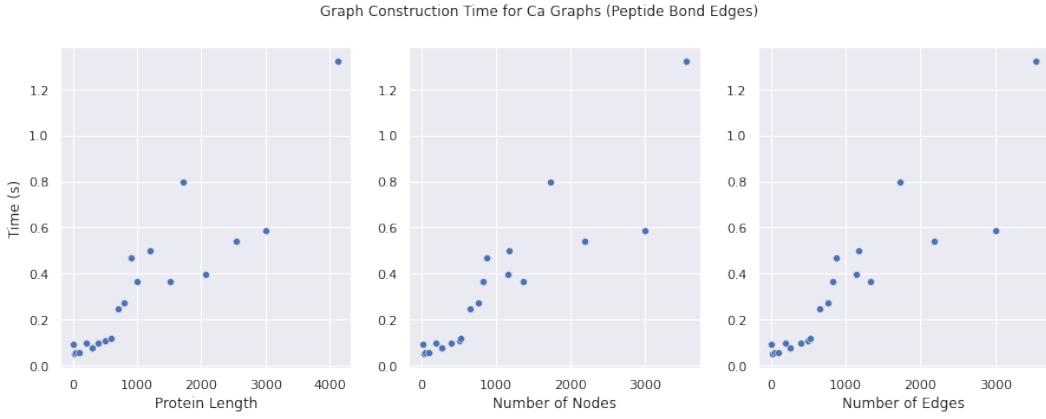


Figure 2: Graph construction times for peptide bond graphs.

F.1.2 Distance-based

We compute epsilon graphs ($\epsilon = 5$) for the above list of structures following the same procedure as F.1.1 to obtain an average construction time of 0.091s per protein.

F.1.3 Intramolecular

We compute protein structure graphs with Van der Waals interactions, Hydrogen bonds, salt bridges and disulfide interactions. We use the same procedure as in F.1.1 to obtain an average construction time of 0.117s per protein.

F.2 Atomic Graphs

We compute atomic covalent bond graphs for the above list of 20 structures 5 times (i.e. 100 evaluations) in parallel (16 workers) and average across 7 runs to obtain an average construction time of 2.5s per protein.

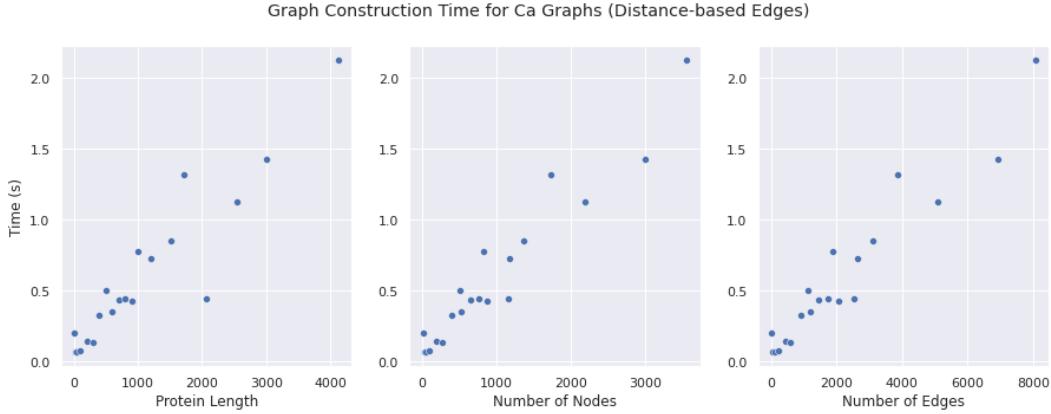


Figure 3: Graph construction times for distance-based edges.

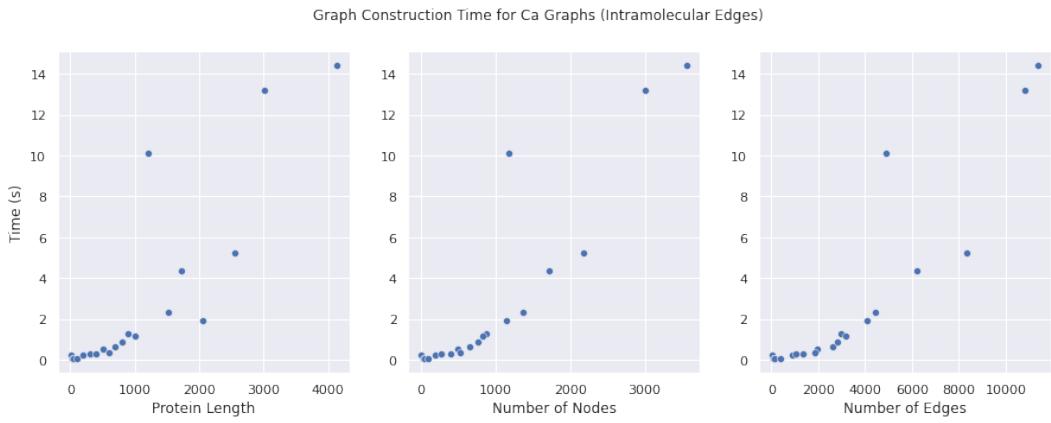


Figure 4: Graph construction times for intramolecular interaction graphs.

G Societal Impact

The potential risks of our library are minimal. A foreseeable hazardous application of our work is its use by nefarious actors to engineer harmful biomolecules, such as engineering toxins with greater efficacy. However, we believe these risks are minimal and shared across any developments in making computational design of biomolecules more accessible. We believe that utility of Graphein in the context of therapeutic development significantly outweighs these unlikely scenarios and anticipate our contribution to be a net force for social good and public health.

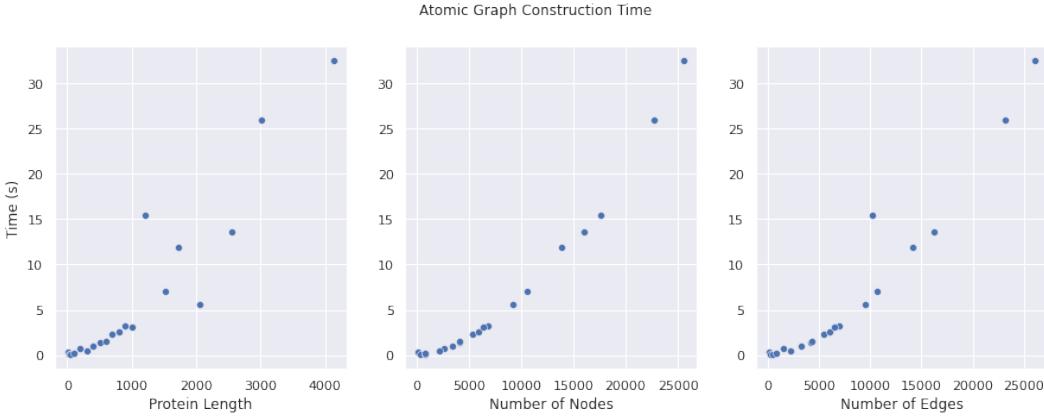


Figure 5: Graph construction times for atomic structure graphs.

References

- [1] Wolfgang Kabsch and Christian Sander. Dictionary of protein secondary structure: Pattern recognition of hydrogen-bonded and geometrical features. *Biopolymers*, 22(12):2577–2637, December 1983. doi: 10.1002/bip.360221211. URL <https://doi.org/10.1002/bip.360221211>.
- [2] Jens Meiler, Anita Zeidler, Felix Schmuschke, and Michael Muller. Generation and evaluation of dimension-reduced amino acid parameter representations by artificial neural networks. *Journal of Molecular Modeling*, 7(9):360–369, September 2001. doi: 10.1007/s008940100038. URL <https://doi.org/10.1007/s008940100038>.
- [3] Elisabeth Gasteiger, Christine Hoogland, Alexandre Gattiker, Severine Duvaud, Marc R. Wilkins, Ron D. Appel, and Amos Bairoch. Protein identification and analysis tools on the ExPASy server. In *The Proteomics Protocols Handbook*, pages 571–607. Humana Press, 2005. doi: 10.1385/1-59259-890-0:571. URL <https://doi.org/10.1385/1-59259-890-0:571>.
- [4] S. Kawashima, P. Pokarowski, M. Pokarowska, A. Kolinski, T. Katayama, and M. Kanehisa. AAindex: amino acid index database, progress report 2008. *Nucleic Acids Research*, 36 (Database):D202–D205, December 2007. doi: 10.1093/nar/gkm998. URL <https://doi.org/10.1093/nar/gkm998>.
- [5] Alexander Rives, Joshua Meier, Tom Sercu, Siddharth Goyal, Zeming Lin, Jason Liu, Demi Guo, Myle Ott, C. Lawrence Zitnick, Jerry Ma, and Rob Fergus. Biological structure and function emerge from scaling unsupervised learning to 250 million protein sequences. *Proceedings of the National Academy of Sciences*, 118(15), 2021. ISSN 0027-8424. doi: 10.1073/pnas.2016239118. URL <https://www.pnas.org/content/118/15/e2016239118>.
- [6] Ehsaneddin Asgari and Mohammad R. K. Mofrad. Continuous distributed representation of biological sequences for deep proteomics and genomics. *PLOS ONE*, 10(11):e0141287, November 2015. doi: 10.1371/journal.pone.0141287. URL <https://doi.org/10.1371/journal.pone.0141287>.
- [7] Raji Heyrovská. Atomic structures of all the twenty essential amino acids and a tripeptide, with bond lengths as sums of atomic covalent radii, 2008.
- [8] GetContacts. Getcontacts. URL <https://getcontacts.github.io/>.
- [9] Dong-Sheng Cao, Qing-Song Xu, and Yi-Zeng Liang. propy: a tool to generate various modes of Chou’s PseAAC. *Bioinformatics*, 29(7):960–962, 02 2013. ISSN 1367-4803. doi: 10.1093/bioinformatics/btt072. URL <https://doi.org/10.1093/bioinformatics/btt072>.
- [10] Anna Gaulton, Anne Hersey, Michał Nowotka, A. Patrícia Bento, Jon Chambers, David Mendez, Prudence Mutowo, Francis Atkinson, Louisa J. Bellis, Elena Cibrián-Uhalte, Mark

Davies, Nathan Dedman, Anneli Karlsson, María Paula Magariños, John P. Overington, George Papadatos, Ines Smit, and Andrew R. Leach. The ChEMBL database in 2017. *Nucleic Acids Research*, 45(D1):D945–D954, November 2016. doi: 10.1093/nar/gkw1074. URL <https://doi.org/10.1093/nar/gkw1074>.