

GENERATING SYNTHETIC GENOTYPES USING DIFFUSION MODELS

Anonymous authors

Paper under double-blind review

ABSTRACT

In this paper, we introduce the first diffusion model designed to generate *complete* synthetic human genotypes, which, by standard protocols, one can straightforwardly expand into full-length, DNA-level genomes. The synthetic genotypes mimic real human genotypes without just reproducing known genotypes, in terms of approved metrics. When training biomedically relevant classifiers with synthetic genotypes, accuracy is near-identical to the accuracy achieved when training classifiers with real data. We further demonstrate that augmenting small amounts of real with synthetically generated genotypes drastically improves performance rates. This addresses a significant challenge in translational human genetics: real human genotypes, although emerging in large volumes from genome wide association studies, are sensitive private data, which limits their public availability. Therefore, the integration of additional, insensitive data when striving for rapid sharing of biomedical knowledge of public interest appears imperative.

1 INTRODUCTION

Deep learning has enabled significant advancements in the field of computational biology (Jumper et al., 2021; Cheng et al., 2023; Wong et al., 2024). However, the vast majority of such approaches resort to processing smaller portions of human genomes, such as coding regions and their products (proteins), or small local segments of human genomes (Guo et al., 2023; Shen et al., 2022).

The reasons for this are threefold. First, the enormous length of human genomes prevents their straightforward usage in neural network architectures. Second, whole human genome data are expensive, because of the massive clinical and experimental efforts required in order to obtain them. Third, whole human genome data are usually subject to strict access regulations because of privacy concerns. The difficulty to process, gather and share sufficient (training) data impedes scientific progress through reliable extraction of knowledge, rapid dissemination of relevant data, and render easy and full reproducibility of already obtained results impossible.

While the first reason is a (challenging) technical concern, the second and the third reason establish our key motivation from the point of view of applications in biomedicine.

The solution that we suggest here is the generation of synthetically generated whole genome data that is cheap, easy to gather, and privacy-enhancing. We present a diffusion model based framework that can generate synthetic whole-genome human genotype data.

Despite the relatively small amounts of data used during training, we ensure that our diffusion models do, in fact, *generate novel whole genome human genotypes*. By means of approved reliable metrics, we ensure that the synthetically generated genotypes are of high quality, that is realistic in terms of stemming from the distribution governing real human genotypes, and also diverse, that is they do not exactly reproduce the individual genomes used for training the diffusion models, which translates into preservation of privacy in the setting at hand.

Note that, unlike previous work (Guo et al., 2023), whole-genome genotypes can be straightforwardly expanded into whole DNA-level genomes, by means of applicable genome reference systems. Furthermore, diffusion models enable us to generate disease-affected and non-disease-affected genotypes in a targeted manner.

Beyond demonstrating that the synthetically generated genomes are realistic by the measures that ensure that the diffusion model approximately captures the distribution of human genotypes, we further demonstrate that training classifiers (Luo et al., 2023) using synthetically generated data, by either integration or replacement, and evaluating them on the original data achieves performance rates that rival those of the original classifiers. This provides further evidence that the diffusion model has not only captured the general structure of human genomes, but also has picked up the mechanisms that distinguish diseased from non-diseased genotypes.

2 RELATED WORK

2.1 PREVIOUS MODELS

In Table 1, we systematically compare most previous work which has tried to produce genomes. To the very best of our knowledge, we are the first approach which succeeds in generating full-length human genotypes. After thoroughly and carefully revisiting the landscape of existing tools and approaches (see Table 1), we conclude that there are indeed no approaches that can generate synthetic full-length human genotypes (or even human genomes directly at the level of DNA). All approaches presented so far address generating smaller segments of human genomes, most not even spanning the length of one chromosome. However, classifiers as the one presented in Luo et al. (2023) require full-length human genome data to work well.

Table 1: An overview of related work on generating synthetic genomes and its differences / similarities in comparison with our work. Row headers are: Reference, the modeling approach used, the data type the model works on, length of generated genomes presented, whether or not the model can be conditioned to produce specific types of data. Our novelties are **highlighted**.

Reference	Model	Data Type	Genome Length	Cond.
DNAGPT Zhang et al. (2023)	Autoregressive	Base-Pairs	24k BPS	x
HyenaDNA Nguyen et al. (2023)	Autoregressive	Base-Pairs	10^6 BPS	x
HAPNEST Wharrie et al. (2023)	LD & Markov	SNPs	1 Chromosome	x
Perera et al. (2022)	GMMNs	SNPs	1 Chromosome	✓
Yelmen et al. (2021)	GAN,RBM	SNPs	10k SNPs	x
Yelmen et al. (2023)	WGAN	SNPs	10k SNPs	x
Szatkownik et al. (2024)	WGAN	PCA+SNPs	65k SNPs	x
Ahronoviz & Gronau (2024)	GAN	SNPs	10k SNPs	✓
Burnard et al. (2023)	VAE	SNPs	1 Chromosome	x
Dang et al. (2023)	HCLTs	SNPs	10k SNPs	x
GeneticDiffusion (Ours)	Diffusion	PCA+SNPs	Full Genome	✓

2.2 GENERATIVE MODELS

We prefer diffusion models over alternative generative deep learning models for the following reasons: a) Diffusion models have become the tool of choice in generative modeling (Dhariwal & Nichol, 2021), and b) previous successes in the use of diffusion models in regulatory human genomics (Sarkar et al., 2024; Avdeyev et al., 2023; Senan et al., 2024; Li et al., 2024) further support their usage. Unlike these works, we emphasize that in our work we make use of the full length of genotypes, and do not have to restrict ourselves to smaller portions of the human genome. We considered skipping steps for generation as done in DDIM (Song et al., 2022) as a way to speed up generation, but realized that the high quality generated by using the full step length for our generation was important for our use case. Furthermore, we considered classifier free guidance (Ho & Salimans, 2022) to increase the quality of the conditioning during generation, like in Azizi et al. (2023), but observed no positive effects for classification accuracy on the test data.

2.3 WORKING WITH LONG SEQUENCES

One of the driving problems when working with genetic data is the enormous length of the genomes. This is exacerbated by the long range interactions that affect parts of the genomes that are far apart in terms of the sequential order in which they appear.

It is therefore no surprise that recent genomics research is employing techniques that can accommodate the large length of human genomes e.g. HyenaDNA (Nguyen et al., 2023), which is a powerful architecture that is able to process up to a million tokens simultaneously via an adapted form of attention, and also, as of most recently, state space models (Schiff et al., 2024), for which similar principles apply. However, even with those recent methodological advances, sequences of billions, and not just millions in length, cannot be processed.

In the domain of diffusion models, works have been presented, for example “Stable Diffusion” (Rombach et al., 2022), that analogously adopt the paradigm of no longer working directly on the raw data, but rather on appropriately embedded versions of it. We adopt this paradigm.

3 METHODS

In the following, we describe the data representations that reflect human genomes, i.e. the genotype profiles that correspond to them, and the computation of embeddings for these genotype profiles, as well as the architectural choices for the diffusion model. For more details on diffusion models and the human Genotype, see the Appendix.

3.1 DATA

We deal with two data sets of human genotypes:

ALS Data. The individual genotype profiles that we use for training (and testing) in the following, were raised in the frame of Project MinE pro (2018), which is concerned with the study of amyotrophic lateral sclerosis (ALS). As a disease, ALS is of particular interest to AI based applications, because ALS is driven by complex, still insufficiently understood mutation patterns that escape the grasp of human-understandable approaches. For exactly these reasons, also earlier studies (Auer et al., 2012; Dolzhenko et al., 2017) focus on ALS, using data gathered through Project MinE.

While Project MinE establishes a data resource that is exemplary in terms of size and comprehensiveness, access to its data is subject to strict safety regulations. This is the reason why we exclusively deal with a Dutch cohort of people, for which we were provided access, while not with cohorts of genotypes referring to other countries. The Dutch cohort we worked with consisted of 3292 individuals affected by ALS and 7213 individuals known not to be affected by ALS, by ancestral relationships.

Accordingly, for this data, labels y used to steer the generation of new samples, refer to individuals affected with ALS and without ALS.

1000 Genomes (1KG). We also consider the 2504 individuals sequenced in Stage 3 of the 1000 Genomes project Consortium et al. (2015). For these 2504 individuals, alleles were assigned to ancestors (referred to as “phased” in genetics), such that one obtains two *haplotypes* instead of one genotype for each individual, where a haplotype is a binary-valued vector of length N where 0 reflects that the reference allele applied at the particular position whereas 1 reflects to observe the alternative allele at the respective SNP site. Adding up the two haplotypes, entry by entry, results in the genotype of the individual. Because, unlike genotypes, the haplotypes assign alleles to ancestors, they carry more information. This is considerably more valuable for genetics, because they provide immediate insight into the ancestral relationships affecting genomes.

When dealing with 1KG data, we seek to generate haplotype profiles instead of merely genotype profiles. Unlike with Project MinE, no particulars about the corresponding phenotypes are known in the frame of the 1KG project. The only known phenotype is the population from which they stem. So, the additional conditioning input y refers to these 26 population labels when working with 1KG data.

3.2 EMBEDDING GENOTYPES / HAPLOTYPES.

In the following, refer to Figure 1 for an illustration the embedding procedure. See also (Luo et al., 2023) for details on the following.

Embedding refers to turning ternary-valued (genotypes, ALS) or binary-valued (haplotypes, 1KG) vectors of length approximately 3-5 millions into real-valued vectors, whose dimension is in the tens of thousands. This does not only reduce the dimensionality of the data, but also ensures consistency in terms of arranging the data for appropriate processing by the diffusion model.

For that transformation, we consider the genes recorded for the ALS and 1KG datasets, amounting to 18279 and 26624 genes, respectively. Based on approved principles, we assign each SNP site to one of the genes. The number of SNP sites per gene can vary quite substantially. Depending on length and location in the genome, a gene can collect roughly between 5 and 100 SNP sites. In other words, each gene reflects a ternary- (ALS) or binary-valued (1KG) vector of length between 5 and 100. Following the approach described in (Luo et al., 2023), we apply principal component analysis (PCA) to each of the 18279 (ALS) or 26624 binary-valued (1KG) vectors of length between 5 and 100, for each gene separately. This amounts to 18279 (ALS) resp. 26624 (1KG) PCAs, each one applied to the vector segments referring to one of the genes. The result is a collection of principal components for each of the genes, both in the case of ALS and the case of 1KG. Depending on the number of SNP sites per gene, we pick between 1 and 8 principal components (PCs) for each of the 18 279 (ALS) or 26624 (1KG) genes. For consistency and due to architectural reasons, we pad any such vector of length less than 8 (corresponding to less than 8 PCs for the particular gene) with zeros to extend it to length 8.

For both ALS and 1KG, the reduction of dimension comes at minimal compression loss ($< 1\%$) as shown in Luo et al. (2023). This strongly implies that one can decompress the PC embedded data into the original genotypes or haplotypes at only a minor loss of information.

Observing that $18432 = 2^{11} \times 9$, we further pad embeddings $x \in \mathbb{R}^{18279 \times 8}$ with further zeros, extending individual genotype embeddings into elements of $\mathbb{R}^{18432 \times 8}$, which aims at efficiency gains with respect to modern hardware architecture. To remedy the issue that zero padded position add additional failure points all values at padded positions are clamped to zero during training and the generation process. We apply the same procedure for the 26624×8 - dimensional vectors referring to the 1KG data.

3.3 MODEL ARCHITECTURE

We base our Diffusion Model on the popular U-Net Architecture suggested by Ronneberger et al. (2015), while applying modifications that account for the sequential nature of the genetic data. We note that the order of the genes along the genome implies a natural order of the different 8-dimensional gene vectors.

We explore different variants of the basic U-Net architecture by replacing the 2D convolutional layers with their 1D counterparts, or with multi-layer perceptrons (MLPs).

We also explore a transformer encoder structure similar to the one presented in Devlin et al. (2019), but with learnable positional embeddings, and two additional tokens accounting for time steps t and class labels y , further equipped with an embedding layer similar to the one used for images in (Dosovitskiy et al., 2020). This serves the purpose of reducing the number of input tokens which is essential for reducing the computational cost. In Figure 2, we visualize the UnetMLP architecture that we propose, it is similar to the convolutional Unet, but does not include multi-headed attention in the intermediate layers. For visualizations of the Transformer and UnetCNN architectures see the Appendix.

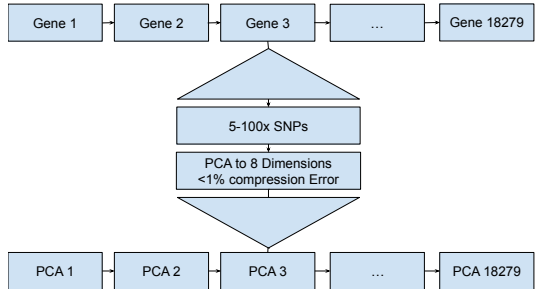


Figure 1: Pre-processing pipeline

In general, we want to point out that the approach using 1D convolutions prioritizes short- to medium-length interactions within the genome, but does only allow for limited long-range interactions. On the other hand, this greatly reduces the amount of parameters to be learned, which offsets the disadvantages from a practical point of view. However not including any kind of spatial information does lead to problems on the presented task, due to the high sensitivity of information with regards towards the position. I.e. it is highly important that the model has positional information about the PCA it processes. This is not the case for the CNN architecture.

In contrast, models solely incorporating dense layers are not subject to sequential biases. However, as in other domains of applications of neural networks, fully connected layers tend to fail to find suitable solutions due to the over-parameterization.

Fully attention based models, for example, models based on transformer encoder architectures do not have any inherent spatial bias, the positional encoding used induces the kind of bias. Following, they should be in theory applicable for this kind of data. Therefore, we also explore such architectures here.

Combining Models Since CNN and MLP based models focus on different aspects of the structure of the genome, we suggest combining them into a single network that benefits from the strengths of the two architectural choices, and synthesizes their advantages. We refer to this combination as CNN + MLP. We combine the two separate models $MLP(x, t, y)$ and $CNN(x, t, y)$ during training, and predict the noise (that the diffusion model has added to the input) accordingly:

$$(MLP + CNN)(x, t, y) = (1 - \lambda(t)) \cdot MLP(x, t, y) + \lambda(t) \cdot CNN(x, t, y) \tag{1}$$

where $\lambda(t), t \in [0, 1]$ reflects a learnable function, realized by a straightforward 2-layer-perceptron receiving the noise schedule t as input.

Overall, we explore 4 different diffusion model architectures: "Unet MLP", "Unet CNN", "Unet MLP + CNN" and "Transformer". We perform extensive hyper-parameter tuning on all of these model types and present the best results in the evaluation.

3.4 EVALUATION

Evaluation of synthetically generated genomes requires careful consideration. The driving underlying principles are realism, on the one hand, and diversity on the other hand. While realism refers to synthetic genomes being likely to stem from the true distribution of genomes, diversity is concerned with sampled synthetic genomes being sufficiently far away from the true training data points. In image generation common scores to measure these metrics are the Fréchet inception distance (FID) (Heusel et al., 2017) or, the Inception score (Salimans et al., 2016), for example. While human eyesight does not apply in genomics for obvious reasons, the FID and IS cannot be computed either because of the integration of pre-trained large scale networks. In fact, this scenario does not apply in genomics/genetics for exactly the reasons that motivate our work: the lack of available (accessible) large-scale data hampers conventional ML practice. Due to these reasons we are forced to rely on metrics which while proven are a bit more unconventional for the image generation domain namely: Adversarial Accuracy and Classifier Performance.

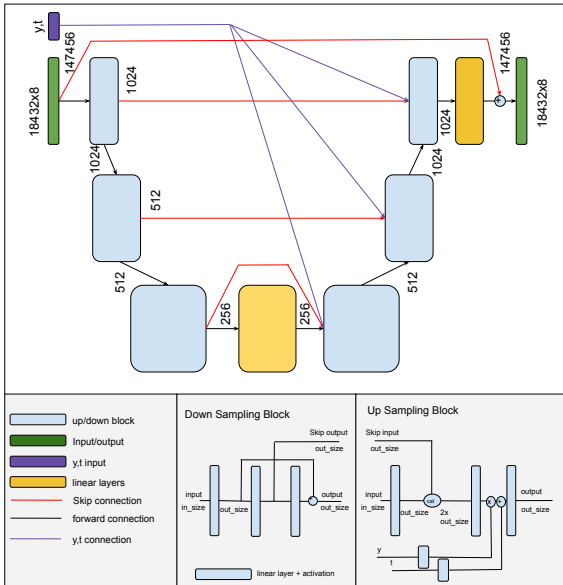


Figure 2: A structural overview of the architecture of the MLP diffusion model.

216
217
218
219
220
221
222
223
224
225
226
227
228
229
230
231
232
233
234
235
236
237
238
239
240
241
242
243
244
245
246
247
248
249
250
251
252
253
254
255
256
257
258
259
260
261
262
263
264
265
266
267
268
269

We note that all datasets we have access to are, although fairly large from the point of view of biomedicine, considerably limited in terms of ML concerns (number of samples at most 10 405). So, one cannot expect the diffusion model to perform at the level of realism observed in other complex domains (such as images and text). This explains why the evaluation relates to exploring the upper limits of possibilities in our context. Note however that our approach virtually serves the purpose of training diffusion models on large-scale data, as hosted by large, access restricted databases, in a safe, access-restricted environments. These databases could then provide safe, privacy-preserving large-scale synthetic data on demand, by drawing samples from the diffusion model, without having to publish neither real data nor the diffusion model trained on real data.

3.4.1 RECOVERY RATE

Employing synthetic data for training reflects using samples from a distribution that was estimated using empirical data. Since all knowledge captured by that distribution stems from the real, empirical data from which it was estimated, any samples drawn from that distribution can, at most, convey the knowledge that contributed to its estimation. In terms of classifier performance, this means that performance rates achieved when using real data for training establish an upper bound for the performance rates that one can achieve when using synthetic data which was generated by observing the same real data for training.

Somewhat more formally, consider a classifier C and a generator G , both of which are trained on the same real data D_r . While G explicitly addresses to approximate the distribution that governs D_r , C implicitly approximates it in order to establish sufficiently accurate classification boundaries. Sampling synthetic data corresponds to drawing data D_s from the distribution approximated by G . So, using D_s , the synthetic data, instead of D_r , the real data, for training C cannot lead to gains in performance, because of the additional layer of approximation that G introduced.

Any improvements that one observes when using D_s instead of D_r are not due to systematic principles, but can only reflect artifacts (such as overfitting of C when using D_r) or random effects (implying that C may not be able to approximate the distribution when using D_r as well as when using D_s). If the generator G is pre-trained on another data set this upper limit does no longer exist.

In summary, it makes sense to evaluate the quality of generated data in terms of how much of the accuracy of the classifiers achieved on real data one can recover when replacing the real data with synthetic data. We perform this evaluation by establishing *recovery rate* $R(a_r, a_s)$ as per the following definition:

$$R(a_r, a_s) = \frac{a_s}{a_r} \quad (2)$$

where a_r is the test accuracy when a classifier is trained on the full real training data and a_s is the test accuracy when the same classifier is trained on a synthetic data set. Based on the above reasoning, one can expect that $R(a_r, a_s) \in [0, 1]$, unless artifacts or random effects disturb the training processes in general.

Note already here that we demonstrate that the generated data do not merely reproduce the real data using other metrics (see just below for the definitions, and Section 4 for the corresponding experiments).

3.4.2 NEAREST NEIGHBOUR ADVERSARIAL ACCURACY

It was shown that diffusion models, when provided with too little training data, tend to reproduce training data instead of generating fresh samples (Somepalli et al., 2023). To quantify at what rate we are affected by such effects, we follow the approach presented by Yale et al. (2019).

$$\begin{aligned}
AA_{truth} &= \frac{1}{n_{truth}} \sum_{i=1}^{n_{truth}} \mathbf{1}(d_{truth,syn}(i) > d_{truth,truth}(i)) \\
AA_{syn} &= \frac{1}{n_{syn}} \sum_{i=1}^{n_{syn}} \mathbf{1}(d_{syn,truth}(i) > d_{syn,syn}(i)) \\
PrivacyLoss &= AA_{truth_{tr},syn} - AA_{truth_{te},syn}
\end{aligned} \tag{3}$$

where $\mathbf{1}$ is the indicator function. Scores of $AA_{truth} = 0.5$ and $AA_{syn} = 0.5$ mean that this metric cannot distinguish *syn* from *truth* (and vice versa). Scores closer to 0 reflect over-fitting, whereas scores closer to 1 reflect under-fitting. Using true training and held out test data $truth_{tr}, truth_{te}$ one can compute privacy loss as $AA_{truth_{tr},syn} - AA_{truth_{te},syn}$. We do not report $AA_{truth,syn} = \frac{1}{2}(AA_{truth} + AA_{syn})$ as in the original paper since underfitting on AA_{truth} and overfitting on AA_{syn} can mutually cancel each other, which leads to deceptively good scores despite the poor models that lead to these scores. For further details we refer the interested reader to the original paper Yale et al. (2019).

3.4.3 UMAP

Another common way of evaluating quality of generated data is the visualization of the neighbourhood structure using algorithms like UMAP (McInnes et al., 2018) or T-SNE (van der Maaten & Hinton, 2008). We employ these visualization tools in this paper.

3.4.4 CLASSIFIER TRAINING

The most important question that we would like to answer is to what degree replacing true training data with synthetically generated training data leads to losses in prediction. For obtaining answers, we consider two scenarios.

First, we focus on predicting the prevalence of the genetic disease Amyotrophic Lateral Sclerosis (ALS), for which the first whole-genome based classifier was presented only recently (Luo et al., 2023). There, training data was selected from 10 405 individual genotypes, referring to 3192 cases, that is individuals affected by ALS, and 7213 controls, that is individuals not affected by ALS, as determined by medical professionals. Further, ALS is known to be a complex and hard to disentangle genetic disease, which means that reliable classification does not depend on a small set of genes. This was documented in (Luo et al., 2023), by showing that good performance could only be established when employing at least on the order of 10^3 genes. Unlike in (Luo et al., 2023), we train a common multilayer perceptron (MLP) as a binary (ALS or not) classifier. Note that performance rates achieved here exceed the performance rates displayed in (Luo et al., 2023).

Second, we consider the 2504 genotypes provided through the 1000 Genomes (1KG) project (Consortium et al., 2015) (stage 3), and the one of 26 population labels assigned to the genotypes, which gives rise to a classification task referring to one of 26 different labels. Again, we establish our primary classifier as an MLP, which parallels the situation for the ALS data.

To demonstrate that favorable usage of synthetically generated data does not depend on a particular type of classifier, we further experiment with a transformer based ("Transformer") and a convolutional neural network ("CNN") based classifier.

We evaluate each of the classifiers, trained with true data on the one hand, and synthetic data, as generated by the Diffusion Model, on the other hand, on held out test data (ALS: balanced, 520 positive/520 negative genotypes; 1KG: 10% of total data, unbalanced, haplotypes). We follow the experimental protocol presented in (Luo et al., 2023) for the ALS data.

4 EXPERIMENTS

In this chapter, we will evaluate the Diffusion Model according to the metrics outlined before.

378
 379
 380
 381
 382
 383
 384
 385
 386
 387
 388
 389
 390
 391
 392
 393
 394
 395
 396
 397
 398
 399
 400
 401
 402
 403
 404
 405
 406
 407
 408
 409
 410
 411
 412
 413
 414
 415
 416
 417
 418
 419
 420
 421
 422
 423
 424
 425
 426
 427
 428
 429
 430
 431

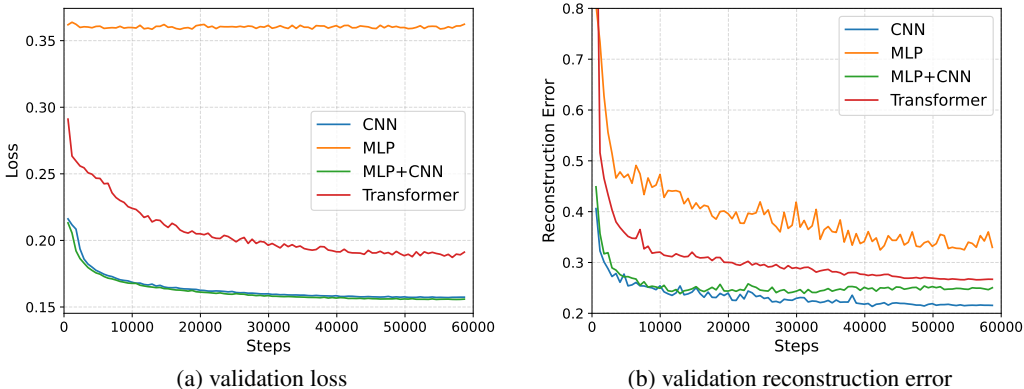


Figure 3: We display validation loss (a) and reconstruction error (b) e.g. $\|x_p - x\|$ during training using single shot denoising.

4.1 TRAINING METRICS

First, we show loss curves during training on validation data for the different diffusion model types, see Figure 3. We observe that none of the models over-fit and all of them reduce the reconstruction error $\|x - x_p\|$ (x_p is the one shot denoising result, see the Appendix for more details) as well as the loss continuously during training.

A closer look at the reconstruction error of the model, visualized in Figure 3 b), shows some interesting results. The MLP model performs only slightly worse compared to the CNN model in reconstruction error, and even though only small drops in loss during training can be observed for the MLP, the reconstruction error keeps improving. The Transformer based architecture seems to be performing well in terms of loss and reconstruction error. We note that reconstruction error is closely related to loss, but scaled by the noise schedule t , which means this error focuses more on large values of t . For further analysis we refer the interested reader to the Appendix for a discussion of the diffusion process.

4.2 EVALUATION

Disease and population classification First, we evaluate different classifiers trained on synthetic data in terms of recovering performance rates that one can achieve on true data—we recall that performance rates achieved on true data establish upper bounds for synthetic data generating mechanisms. See Table 2 for the corresponding results.

We observe that the combined MLP + CNN (U-Net) type architecture outperforms other generator architectures on both 1KG and ALS data, with near-perfect recovery rates on the ALS data. For ALS, the difference between MLP and MLP + CNN generated data is small, but on 1KG data, MLP + CNN clearly outperforms the other architectures. Synthetic data generated by Transformer and CNN based U-Net architectures perform poorly when used for training classifiers. See the Appendix for the accuracy values on which recovery rates are based.

Secondly, we consider the (ubiquitous) scenario where a translational geneticist is forced to restrict her-/himself to a set of available true genotypes that is too small to train reliable classifiers. Here, we evaluate how augmenting small sets of real "seed" training data with larger amounts of synthetically generated data—which may be easily and cheaply available for the particular user—improves performance rates.

See Table 3 for the corresponding results. For example, augmenting only 5% of the real training data (leading to 70% accuracy when used in isolation for training) with synthetically generated data to the overall full amount of data nearly re-establishes the accuracy when using the full, real training data set. This means that the availability of sufficiently large synthetic data sets may rescue efforts

of researchers that remain with too little training data sets due to, for example, budget constraints or restrictive access regulations.

Table 2: Recovery rates on a hold out test set of true genotypes after training different ALS or 1KG population classifiers (MLP, Transformer or CNN) on different synthetically generated data types (generated by: MLP, Transformer, CNN, MLP + CNN). The best synthetic data for each classifier type is marked in **bold**.

	Classifier	CNN	MLP	MLP + CNN	Transformer
ALS data	MLP	71.51	96.69	99.49	73.77
	Transformer	66.06	93.44	99.41	69.30
	CNN	69.88	91.72	91.46	68.72
1KG data	MLP	15.58	65.80	93.02	13.28
	Transformer	16.23	62.99	84.98	8.38
	CNN	19.52	56.57	77.54	21.21
Average (all)		43.17	78.06	90.98	42.56

Table 3: Accuracy improvements by integration of best synthetic data for best performing classification architecture.

	amount of real data	5%	10%	20%	50%
ALS Data	no synthetic data	70.96	76.50	80.90	84.60
	with synthetic data	84.83	85.01	85.34	85.70
1KG data	no synthetic data	29.01	43.99	71.52	85.19
	with synthetic data	83.98	86.93	87.11	87.50

Nearest Neighbour Adversarial Accuracy (NNAA) We further evaluate the nearest neighbour adversarial accuracy and Privacy Loss (see Eq. 3), in Table 4. We observe that the combined MLP + CNN (U-Net) architecture performs well in terms of both nearest neighbour adversarial accuracy, and Privacy Loss. Of note, also Transformer and CNN generated data points deliver similar but slightly worse performance in terms of these metrics. We draw two conclusions from this:

Interpreting the experiments, we conclude that the generated data is of sufficiently good quality, in particular that for the combined MLP + CNN architecture, documented by most scores being sufficiently close to 0.5. Improvements are conceivable, however, because the amount of training data used for training generators is likely at the lower limit of quantities required for sound estimation of such high dimensional and complex distributions.

To further quantify the preservation of privacy, we calculated L1, L2, and cosine distances between synthetic and real data points. Thereby, we can confirm that none of the synthetic data points matches any of the real data points. Note that this finding is crucial for maintaining the integrity of diffusion models in terms of privacy (i.e. diversity from a general perspective, which is also reflected in the NNAA score).

In summary, we conclude that the combined MLP + CNN U-Net architecture leads to synthetic genotypes/haplotypes that not only re-establish excellent performance in terms of classification, but also preserve the privacy of the real data used for training the generators to a sufficiently reliable degree.

Table 4: Result of Nearest Neighbour Adversarial Accuracy for generated datasets on the ALS and 1KG data; For AA the closer to 0.5 the better, For Privacy Loss the closer to 0 the better; best performance is **highlighted**.

			CNN	MLP	MLP + CNN	Transformer	
ALS data	test data	AA_{truth}	0.735	0.255	0.485	0.92	
		AA_{syn}	0.68	1.0	0.93	0.66	
	train data	AA_{truth}	0.81	0.005	0.405	0.93	
		AA_{syn}	0.67	1.0	0.92	0.69	
	Privacy Loss			0.0325	0.125	0.0475	0.02
	1KG data	test data	AA_{truth}	0.76	0.0	0.63	0.345
AA_{syn}			0.995	1.0	0.94	0.92	
train data		AA_{truth}	0.765	0.0	0.385	0.285	
		AA_{syn}	1.0	0.99	0.74	0.82	
Privacy Loss			0.05	-0.005	-0.2225	0.08	

5 CONCLUSION

In this work, we have presented, to the best of our knowledge, the first diffusion model based approach by which to generate full-length human genotypes. In this, by standard expansion of genotypes using human genome reference systems, we have also presented an approach by which to generate full-length human genomes at the level of DNA.

In our experiments we have demonstrated, that the synthetically generated genotypes are realistic and that they do not just reproduce the real human genotypes used as input for training.

To demonstrate the practical usefulness of synthetic genotypes, we have employed synthetically generated genotypes as training data for disease- or population-related classifiers. We have shown that such practice re-establishes original performance rates to a degree that justifies their usage in translational genetics research.

Improvements of the diffusion models are readily conceivable by increasing the amount of training data. Note that although limited, one can expect amounts of training data available for training generative models to be larger in real world settings. The reason is that generators can be trained in safe environments, for example as part of the databases that host large numbers of genotype cohorts, which implies that none of the real data has to be shared. Including differential privacy mechanisms may further open up opportunities for usage of generative models in (e.g. federated learning) settings, where sharing parameters of the generative models may be beneficial.

We publish all code at [anonymized for review].

REPRODUCIBILITY STATEMENT

We publish all code on github and in the supplementary materials used during training and evaluation of the models. The datasets used in the paper were the 1000 Genome Project Consortium et al. (2015) and Project MinE pro (2018). The 1000 Genome data is freely available, while the Project MinE data is only available to credited researchers after a review process.

ETHICS STATEMENT

Working with human genetic data involves significant privacy and ethical challenges. Our approach aims to mitigate privacy issues by generating synthetic data using diffusion models. However, there remains a potential risk that synthetic samples could inadvertently reveal information about original

540 data. While we conducted experiments to ensure that our model does not reproduce exact copies of
 541 real samples, we cannot fully guarantee that original data cannot be inferred from the synthetic out-
 542 puts. Future work could incorporate differential privacy mechanisms to provide theoretical privacy
 543 guarantees, although this may compromise the model’s performance.

544 We evaluate our diffusion model on real-world classification tasks from previous studies, including
 545 the identification of ALS patients, a task that holds promise for future therapeutic advancements. We
 546 also use the 1KG Genome ethnicity prediction task, which, while not directly useful to real-world
 547 scenarios, serves as a benchmark for evaluating the model’s performance on genome-level data.

548 In general we envisioned the diffusion model being trained in a secure environment and the synthet-
 549 ically generated privatised data being released for further research. To obtain provable privacy, we
 550 suggest mechanisms which lead to provable guarantees, such as differential privacy or multiparty
 551 computation. As the transfer of SNP data in between different locations might harm privacy, such
 552 methods could be combined with federated learning technologies.

553 A further challenge is a possible bias which can occur due to a skewed training set. Evaluation
 554 whether biases exist can be based on downstream tasks. Bias mitigation could be based on resam-
 555 pling strategies for the training data.

556 Finally, the same as all AI models, we expect that the method is vulnerable to attacks such as data
 557 poisoning or adversarial attacks of downstream tasks. Thus, the credibility of data sources and
 558 robustness of downstream models needs to be assured. We think, however, that the implementation
 559 of these variants is out of the scope of the current paper.

562 REFERENCES

- 563 Project mine: study design and pilot analyses of a large-scale whole-genome sequencing study in
 564 amyotrophic lateral sclerosis. *European Journal of Human Genetics*, 26(10):1537–1546, 2018.
- 565 Shaked Ahronoviz and Ilan Gronau. Genome-ac-gan: Enhancing synthetic genotype generation
 566 through auxiliary classification. *bioRxiv*, pp. 2024–02, 2024.
- 567 Paul L Auer, Jill M Johnsen, Andrew D Johnson, Benjamin A Logsdon, Leslie A Lange, Michael A
 568 Nalls, Guosheng Zhang, Nora Franceschini, Keolu Fox, Ethan M Lange, et al. Imputation of
 569 exome sequence variants into population-based samples and blood-cell-trait-associated loci in
 570 african americans: Nhlbi go exome sequencing project. *The American Journal of Human Genet-*
 571 *ics*, 91(5):794–808, 2012.
- 572 Pavel Avdeyev, Chenlai Shi, Yuhao Tan, Kseniia Dudnyk, and Jian Zhou. Dirichlet diffusion score
 573 model for biological sequence generation. In *International Conference on Machine Learning*, pp.
 574 1276–1301. PMLR, 2023.
- 575 Shekoofeh Azizi, Simon Kornblith, Chitwan Saharia, Mohammad Norouzi, and David J. Fleet. Syn-
 576 thetic data from diffusion models improves imagenet classification. *Transactions on Machine*
 577 *Learning Research*, 2023. ISSN 2835-8856. URL [https://openreview.net/forum?](https://openreview.net/forum?id=DlRsoxjyPm)
 578 [id=DlRsoxjyPm](https://openreview.net/forum?id=DlRsoxjyPm).
- 579 Callum Burnard, Alban Mancheron, and William J Ritchie. Generating realistic artificial human
 580 genomes using adversarial autoencoders. *bioRxiv*, pp. 2023–12, 2023.
- 581 Jun Cheng, Guido Novati, Joshua Pan, Clare Bycroft, Akvilė Žemgulytė, Taylor Applebaum,
 582 Alexander Pritzel, Lai Hong Wong, Michal Zielinski, Tobias Sargeant, et al. Accurate proteome-
 583 wide missense variant effect prediction with alphamissense. *Science*, 381(6664):eadg7492, 2023.
- 584 1000 Genomes Project Consortium et al. A global reference for human genetic variation. *Nature*,
 585 526(7571):68, 2015.
- 586 Meihua Dang, Anji Liu, Xinzhu Wei, Sriram Sankararaman, and Guy Van den Broeck. Tractable
 587 and expressive generative models of genetic variation data. *bioRxiv*, 2023.
- 588 Jacob Devlin, Ming-Wei Chang, Kenton Lee, and Kristina Toutanova. BERT: pre-training of deep
 589 bidirectional transformers for language understanding. In Jill Burstein, Christy Doran, and

- 594 Tamar Solorio (eds.), *Proceedings of the 2019 Conference of the North American Chapter of*
595 *the Association for Computational Linguistics: Human Language Technologies, NAACL-HLT*
596 *2019, Minneapolis, MN, USA, June 2-7, 2019, Volume 1 (Long and Short Papers)*, pp. 4171–
597 4186. Association for Computational Linguistics, 2019. doi: 10.18653/V1/N19-1423. URL
598 <https://doi.org/10.18653/v1/n19-1423>.
- 599 Prafulla Dhariwal and Alexander Nichol. Diffusion models beat gans on image synthesis. *Advances*
600 *in neural information processing systems*, 34:8780–8794, 2021.
- 602 Egor Dolzhenko, Joke Jfa Van Vugt, Richard J Shaw, Mitchell A Bekritsky, Marka Van Blitterswijk,
603 Giuseppe Narzisi, Subramanian S Ajay, Vani Rajan, Bryan R Lajoie, Nathan H Johnson, et al. De-
604 tection of long repeat expansions from pcr-free whole-genome sequence data. *Genome research*,
605 27(11):1895–1903, 2017.
- 607 Alexey Dosovitskiy, Lucas Beyer, Alexander Kolesnikov, Dirk Weissenborn, Xiaohua Zhai, Thomas
608 Unterthiner, Mostafa Dehghani, Matthias Minderer, Georg Heigold, Sylvain Gelly, et al. An
609 image is worth 16x16 words: Transformers for image recognition at scale. *arXiv preprint*
610 *arXiv:2010.11929*, 2020.
- 611 Zhiye Guo, Jian Liu, Yanli Wang, Mengrui Chen, Duolin Wang, Dong Xu, and Jianlin Cheng.
612 Diffusion models in bioinformatics and computational biology. *Nat Rev Bioeng*, 2(2):136–154,
613 October 2023.
- 615 Martin Heusel, Hubert Ramsauer, Thomas Unterthiner, Bernhard Nessler, and Sepp Hochreiter.
616 Gans trained by a two time-scale update rule converge to a local nash equilibrium. *Advances in*
617 *neural information processing systems*, 30, 2017.
- 618 Jonathan Ho and Tim Salimans. Classifier-free diffusion guidance. *arXiv preprint*
619 *arXiv:2207.12598*, 2022.
- 621 Jonathan Ho, Ajay Jain, and Pieter Abbeel. Denoising diffusion probabilistic models. 2020.
- 623 John Jumper, Richard Evans, Alexander Pritzel, Tim Green, Michael Figurnov, Olaf Ronneberger,
624 Kathryn Tunyasuvunakool, Russ Bates, Augustin Žídek, Anna Potapenko, et al. Highly accurate
625 protein structure prediction with alphafold. *Nature*, 596(7873):583–589, 2021.
- 626 Zehui Li, Yuhao Ni, William AV Beardall, Guoxuan Xia, Akashaditya Das, Guy-Bart Stan, and
627 Yiren Zhao. Discdiff: Latent diffusion model for dna sequence generation. *arXiv preprint*
628 *arXiv:2402.06079*, 2024.
- 630 Xiao Luo, Xiongbin Kang, and Alexander Schönhuth. Predicting the prevalence of complex genetic
631 diseases from individual genotype profiles using capsule networks. In *Nature Machine Intelli-*
632 *gence*. Nature Publishing, 2023. ISBN 978-3-319-24574-4.
- 633 Leland McInnes, John Healy, Nathaniel Saul, and Lukas Großberger. Umap: Uniform manifold
634 approximation and projection. *Journal of Open Source Software*, 3(29):861, 2018. doi: 10.
635 21105/joss.00861. URL <https://doi.org/10.21105/joss.00861>.
- 637 Eric Nguyen, Michael Poli, Marjan Faizi, Armin Thomas, Callum Birch-Sykes, Michael Wornow,
638 Aman Patel, Clayton Rabideau, Stefano Massaroli, Yoshua Bengio, Stefano Ermon, Stephen A.
639 Baccus, and Chris Ré. Hyenadna: Long-range genomic sequence modeling at single nucleotide
640 resolution, 2023.
- 641 Maria Perera, Daniel Mas Montserrat, Míriam Barrabés, Margarita Geleta, Xavier Giró-i Nieto, and
642 Alexander G Ioannidis. Generative moment matching networks for genotype simulation. In *2022*
643 *44th Annual International Conference of the IEEE Engineering in Medicine & Biology Society*
644 *(EMBC)*, pp. 1379–1383. IEEE, 2022.
- 646 Robin Rombach, Andreas Blattmann, Dominik Lorenz, Patrick Esser, and Björn Ommer. High-
647 resolution image synthesis with latent diffusion models. In *Proceedings of the IEEE/CVF confer-*
ence on computer vision and pattern recognition, pp. 10684–10695, 2022.

- 648 Olaf Ronneberger, Philipp Fischer, and Thomas Brox. U-net: Convolutional networks for biomedical
649 image segmentation. In Nassir Navab, Joachim Hornegger, William M. Wells, and Alejandro F. Frangi (eds.), *Medical Image Computing and Computer-Assisted Intervention – MICCAI 2015*, pp. 234–241, Cham, 2015. Springer International Publishing. ISBN 978-3-319-24574-4.
- 652 Tim Salimans, Ian Goodfellow, Wojciech Zaremba, Vicki Cheung, Alec Radford, and Xi Chen.
653 Improved techniques for training gans. *Advances in neural information processing systems*, 29,
654 2016.
- 656 Anirban Sarkar, Ziqi Tang, Chris Zhao, and Peter Koo. Designing dna with tunable regulatory
657 activity using discrete diffusion. *bioRxiv*, pp. 2024–05, 2024.
- 658 Yair Schiff, Chia-Hsiang Kao, Aaron Gokaslan, Tri Dao, Albert Gu, and Volodymyr Kuleshov.
659 Caduceus: Bi-directional equivariant long-range dna sequence modeling. *arXiv preprint*
660 *arXiv:2403.03234*, 2024.
- 662 Simon Senan, Aniketh Janardhan Reddy, Zach Nussbaum, Aaron Wenteler, Matei Bejan, Michael I
663 Love, Wouter Meuleman, and Luca Pinello. Dna-diffusion: Leveraging generative models for
664 controlling chromatin accessibility and gene expression via synthetic regulatory elements. In
665 *ICLR 2024 Workshop on Machine Learning for Genomics Explorations*, 2024.
- 666 Xiaoxi Shen, Chang Jiang, Yalu Wen, Chenxi Li, and Qing Lu. A brief review on deep learn-
667 ing applications in genomic studies. *Frontiers in Systems Biology*, 2, 2022. ISSN 2674-0702.
668 doi: 10.3389/fsysb.2022.877717. URL [https://www.frontiersin.org/articles/
669 10.3389/fsysb.2022.877717](https://www.frontiersin.org/articles/10.3389/fsysb.2022.877717).
- 671 Jascha Sohl-Dickstein, Eric Weiss, Niru Maheswaranathan, and Surya Ganguli. Deep unsupervised
672 learning using nonequilibrium thermodynamics. In *International conference on machine learn-
673 ing*, pp. 2256–2265. PMLR, 2015.
- 674 Gowthami Somepalli, Vasu Singla, Micah Goldblum, Jonas Geiping, and Tom Goldstein. Under-
675 standing and mitigating copying in diffusion models. *Advances in Neural Information Processing
676 Systems*, 36:47783–47803, 2023.
- 677 Jiaming Song, Chenlin Meng, and Stefano Ermon. Denoising diffusion implicit models, 2022.
- 679 Antoine Szatkownik, Cyril Furtlehner, Guillaume Charpiat, Burak Yelmen, and Flora Jay. Towards
680 creating longer genetic sequences with gans: Generation in principal component space. In *Ma-
681 chine Learning in Computational Biology*, pp. 110–122. PMLR, 2024.
- 682 Laurens van der Maaten and Geoffrey Hinton. Visualizing data using t-sne. *Journal of Ma-
683 chine Learning Research*, 9(86):2579–2605, 2008. URL [http://jmlr.org/papers/v9/
684 vandermaaten08a.html](http://jmlr.org/papers/v9/vandermaaten08a.html).
- 686 Sophie Wharrie, Zhiyu Yang, Vishnu Raj, Remo Monti, Rahul Gupta, Ying Wang, Alicia Martin,
687 Luke J O’Connor, Samuel Kaski, Pekka Marttinen, Pier Francesco Palamara, Christoph Lip-
688 pert, and Andrea Ganna. HAPNEST: efficient, large-scale generation and evaluation of syn-
689 thetic datasets for genotypes and phenotypes. *Bioinformatics*, 39(9):btad535, 08 2023. ISSN
690 1367-4811. doi: 10.1093/bioinformatics/btad535. URL [https://doi.org/10.1093/
691 bioinformatics/btad535](https://doi.org/10.1093/bioinformatics/btad535).
- 692 Felix Wong, Erica J Zheng, Jacqueline A Valeri, Nina M Donghia, Melis N Anahtar, Satotaka Otori,
693 Alicia Li, Andres Cubillos-Ruiz, Aarti Krishnan, Wengong Jin, et al. Discovery of a structural
694 class of antibiotics with explainable deep learning. *Nature*, 626(7997):177–185, 2024.
- 696 Andrew Yale, Saloni Dash, Ritik Dutta, Isabelle Guyon, Adrien Pavao, and Kristin P Bennett. Pri-
697 vacy preserving synthetic health data. In *ESANN 2019-European Symposium on Artificial Neural
698 Networks, Computational Intelligence and Machine Learning*, 2019.
- 700 Burak Yelmen, Aurélien Decelle, Linda Ongaro, Davide Marnetto, Corentin Tallec, Francesco Mon-
701 tinaro, Cyril Furtlehner, Luca Pagani, and Flora Jay. Creating artificial human genomes using
generative neural networks. *PLoS genetics*, 17(2):e1009303, 2021.

702 Burak Yelmen, Aurélien Decelle, Leila Lea Boulos, Antoine Szatkownik, Cyril Furtlehner, Guil-
703 laume Charpiat, and Flora Jay. Deep convolutional and conditional neural networks for large-scale
704 genomic data generation. *PLoS Computational Biology*, 19(10):e1011584, 2023.
705
706 Daoan Zhang, Weitong Zhang, Bing He, Jianguo Zhang, Chenchen Qin, and Jianhua Yao. Dnagpt: a
707 generalized pretrained tool for multiple dna sequence analysis tasks. *bioRxiv*, pp. 2023–07, 2023.
708
709
710
711
712
713
714
715
716
717
718
719
720
721
722
723
724
725
726
727
728
729
730
731
732
733
734
735
736
737
738
739
740
741
742
743
744
745
746
747
748
749
750
751
752
753
754
755

A APPENDIX

UMAP VISUALIZATIONS

In Figure 4, data points from real, train and test, as well as synthetically generated data points are visualized using UMAP (McInnes et al., 2018). We observe that almost all generated data points, with the exception of the MLP generated points, are well distributed i.e. hard to differentiate from both test and training data.

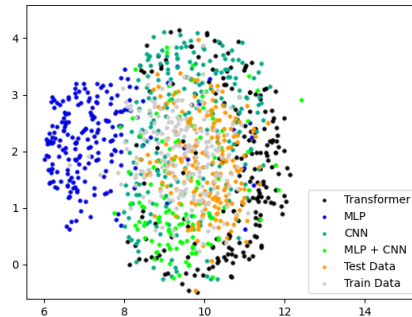


Figure 4: UMAP visualizations of data of different origins. Orange and grey points are test and train data (real). Blue, bright green, black and dark green are MLP, MLP + CNN, Transformer and CNN respectively.

ANALYZING THE RECONSTRUCTION ERROR OF DIFFERENT MODELS

To further analyze the reconstruction error of different models, we visualize the reconstruction error curves vs amount of noise added of the different diffusion models, in Figure 5. We observe that while the CNN has an overall better performance it focuses more on the fine detail of the structure e.g. recovering from small noise values, while the MLP architecture is more focused on recovering rough structure (e.g. high noise). Furthermore the MLP model does observe almost no improvement during training in recovering fine-details, but solely in recovering rough structure.

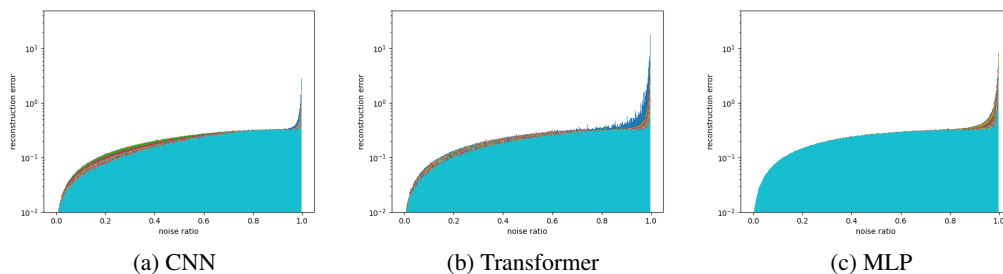


Figure 5: Reconstruction error vs noise curves during training for different diffusion model backbones. Note the logarithmic scaling of the y-axis. The different colors indicate different epochs during training. The final result has the color light blue. This highlights the improvements during training that the model experiences.

We also see this confirmed when looking at the λ parameter found for the MLP+CNN network version, see Figure 6 (a). We observe in Figure 6 (b) how the two parts of the network complement each other especially after longer training times.

810
811
812
813
814
815
816
817
818
819
820
821
822
823
824
825
826
827
828
829
830
831
832
833
834
835
836
837
838
839
840
841
842
843
844
845
846
847
848
849
850
851
852
853
854
855
856
857
858
859
860
861
862
863

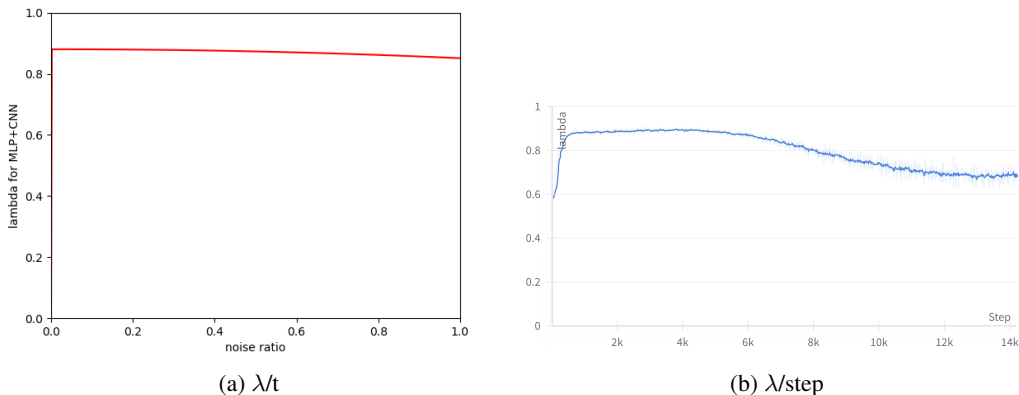


Figure 6: On the left, λ vs t (noise amplitude) curves after training for the MLP + CNN diffusion model backbones. Higher lambda results in more weight on the CNN part of the network. On the right, λ averaged over noise levels on y-axis and training step on x-axis during training for the MLP + CNN diffusion model backbones. Higher lambda results in more weight on the CNN part of the network.

In Figure 7 we analyze various bottlenecks of our diffusion models. The MLP based diffusion model as depicted in Figure 7 a) has trouble finding much structure in the data and only clearly differentiates MLP + CNN generated data from the rest.

The CNN based diffusion model as depicted in Figure 7 b) finds more structure and difference between data of synthetic origin and real data.

Both models on their own are not very accurate at separating the Transformer based data or their own generated data from the original data. We postulate that this might be a reason why they complement each other well in the hybrid MLP + CNN architecture.

However, it is unclear why both models have trouble separating the Transformer based data from the train and test data. This would generally indicate that the Transformer based data is of high quality, which other metrics (NNA, Recovery Rates) in this paper disagree with.

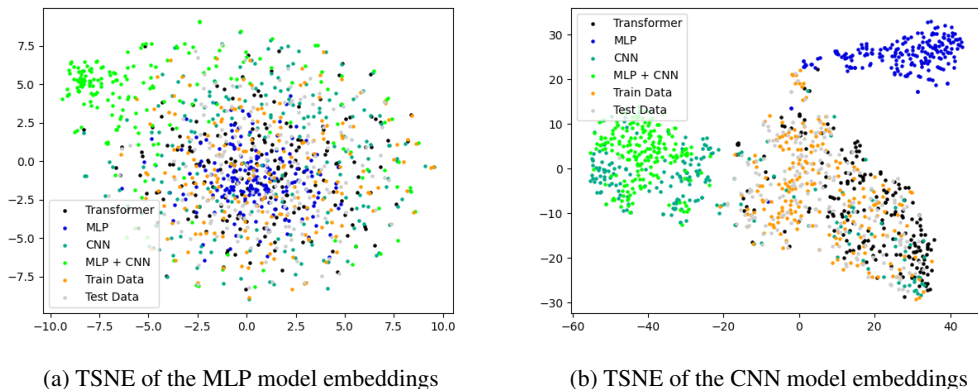


Figure 7: On the left, a TSNE dimension reduction of the bottleneck of the MLP diffusion model of 200 genomes from various data sources. The same on the right for the CNN diffusion model.

ARCHITECTURE DETAILS

In this section we present some technical details in more depth than possible in the main paper.

Table 5: Technical details for all generative models. Tflops (10^{12} flops) were determined using Pytorchs profiler. Training time was measured on a single Quadro 5000 RTX in seconds for the ALS data set. Units are given in brackets ().

	CNN	MLP	MLP + CNN	Transformer
Training time (s)	45.000	8.000	52.000	58.000
Parameter count	18.5561.68	310.094.848	328.651.401	135.280.130
Tflops per genome (Tflops)	1.73	0.62	2.35	49.29

CNN

The CNN diffusion model we use is very similar to the one used in stable diffusion (Rombach et al., 2022) and looks structurally similar to the MLP Unet. It is made up of 1D Convolutions instead of 2D, has 8 downsampling blocks with channel multipliers (1,1,1,1,2,2,3,4) and multi head attention at blocks 5 and 6. The base filter size used in our CNN experiments is 64. See Figure 8 for more details. A very deep convolutional architecture was used with a relatively large number of down and up blocks to account for the large sequence length of 18,000 of the genome data compared to typical image sizes of 1,000. In particular we want to point out that we experimented with a linear layer for each PCA in front of the CNN architecture. This was done to project the Genomes PCA into a shared embedding-space. Sadly this did not achieve superior performance.

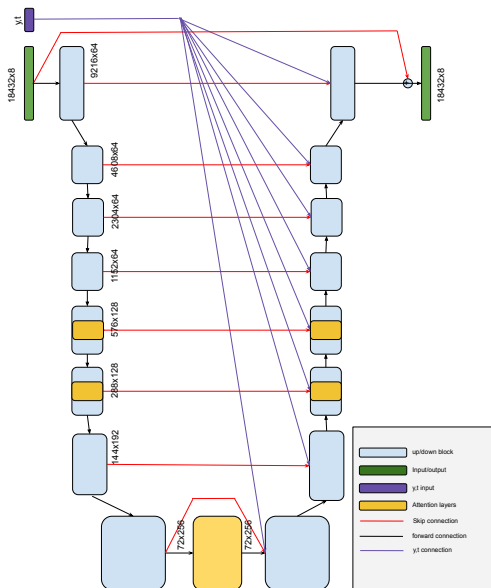


Figure 8: An overview of the CNN diffusion model architecture.

MLP

See Figure 2 in the main paper, for a good overview of the MLP diffusion model.

Transformer

The transformer architecture we use is very similar to the one used in ViT (Dosovitskiy et al., 2020), we use 12 encoder layers with feature size of 384, see Figure 9 for more details. The conditioning in the form of t and y is injected using 2 additional tokens. The first and last linear projection layers don't have shared weights for every position as in ViT, but rather are unique to every position. This was done as in contrast to the image domain where pixels always encode the same information, SNPs do encode different genetic mutations depending on position.

918
919
920
921
922
923
924
925
926
927
928
929
930
931
932
933
934
935
936
937
938
939
940
941
942
943
944
945
946
947
948
949
950
951
952
953
954
955
956
957
958
959
960
961
962
963
964
965
966
967
968
969
970
971

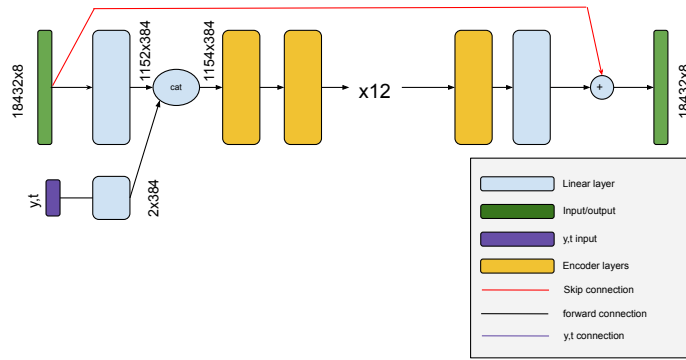


Figure 9: An overview of the transformer diffusion model architecture.

ADDITIONAL RESULTS

Here we display the results from section 4 not as recovery rates but as accuracy's.

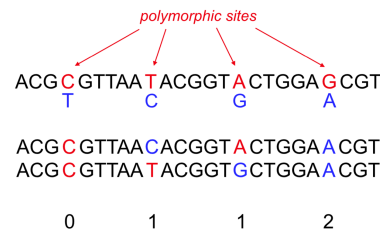
Table 6: Accuracy on a hold out test set after training different ALS or 1KG population classifiers (MLP, Transformer or CNN) on different synthetically generated data types (generated by: MLP, Transformer, CNN, MLP + CNN or Baseline). The best synthetic data for each classifier type is marked in **bold**.

Classifier	Real Data	CNN	MLP	MLP + CNN	Transformer
ALS data					
MLP	87.60	62.64	84.71	87.15	64.62
Transformer	82.11	54.23	76.73	81.63	56.92
CNN	73.17	51.15	67.11	66.92	50.29
1KG data					
MLP	90.23	14.06	59.38	83.98	11.98
Transformer	74.61	12.11	47.01	63.41	6.25
CNN	77.34	15.10	43.75	59.99	16.41
Average (all)	80.84	34.89	63.12	72.94	34.41

DISCUSSION ON HUMAN GENOTYPES.

The human genome is a sequence of approximately 3 billion letters $\{A, C, G, T\}$, reflecting the nucleotides that form the basis of DNA. The vast majority of the 3 billion letters in human genomes are identical across all individual genomes; only approximately 3-5 million positions, referred to as *polymorphic sites*, vary. Although all types of variations can be observed at such polymorphic sites, the vast majority of such sites exhibit single nucleotide polymorphisms (SNPs), defined by exchanges of single letters. In the following, we will only deal with SNP sites, and neglect polymorphic sites characterized by other types of mutations. Restricting oneself to studying SNPs is well justified, because the vast majority of other sites are in *linkage disequilibrium* (*LD*) with SNP sites. That is, in other words, they are statistically strongly correlated with SNP patterns nearby, such that one can safely infer non-SNP site contents based on knowing the SNPs of an individual.

Further, the great majority of SNP sites are *biallelic*, which means that only two letters happen to appear in human genomes; for example, a particular site may be defined by an *A* showing in some genomes, and a *T* showing in the other genomes. The letter that appears in the



18
Figure 10: The (unphased) genotype counts the number of alternative alleles (blue) in the two ancestral genome copies each individual inherits.

majority of people is referred to as *reference allele*, whereas the letter showing in the other, minor fraction of people is referred to as *alternative allele*. Every human genome consists of two copies, of which one is inherited from the mother, and the other from the father. Of course, the contents of the two copies can vary at the SNP sites: the reference allele can show in both copies, referred to as *homozygous for the reference allele*, the reference allele can show in one copy while the alternative allele shows in the other copy, referred to as *heterozygous* (for example, while the mother genome copy exhibits the reference allele G , the father copy exhibits the alternative allele C , or vice versa) or the alternative allele can show in both copies, referred to as *homozygous for the alternative allele*.

Let N be the number of SNP sites in human genomes. The *genotype profile* G is a vector of length N over the entries $\{0, 1, 2\}$, where 0, 1, 2 refer to alternative allele counts at the SNP sites. That is, 0 reflects a polymorphic site that is homozygous for the alternative allele, 1 refers to a heterozygous site, and so on. For example, let $i \in \{1, \dots, N\}$ refer to the i -th SNP site, and let G be the genotype of an individual, then $G[i] = 1$ reflects that the individual that gives rise to G inherited the reference allele from one of the ancestors, while having inherited the alternative allele from the other ancestor.

Note that expanding the genotype G for an individual into a full-length genome over the alphabet $\{A, C, G, T\}$ corresponds to a straightforward operation: for any non-SNP polymorphic site, insert the by LD principles statistically most likely variant, and for any non-polymorphic site, insert the only applicable reference letter. While not necessarily matching the real genome that gave rise to G in all places, the genome resulting from this expansion operation is highly likely to reflect true genetic sequence in the great majority of places.

In our work, the input to our diffusion model are individual genotype profiles $G \in \{0, 1, 2\}^N$, where N is the number of SNP sites. Individual genotype profiles are the by far predominant way of representing human genomes in the frame of genome wide association studies (GWAS). In the meantime, large databases have been filled up with individual genotypes of this kind. Of course, access to such genotype profiles is subject to stringent access regulations, because it is straightforward to match genotype profiles with individuals in a unique manner—in fact, already the content of 50 (sufficiently distant) SNP sites suffices to uniquely identify single individuals.

Note that untreated genotype profiles are still too large to serve as input to diffusion models. Preferably, one can trim down the length of the input from several (3-5) millions to only several tens of thousands, that is by a factor of 100. Below, we describe how to embed genotypes into real-valued vector spaces of dimension in the tens of thousands.

DIFFUSION MODELS

In this section we provide a brief overview of the diffusion process that we have implemented. For details on diffusion processes in general, please see Ho et al. (2020); Song et al. (2022).

TRAINING

Although originally presented in Sohl-Dickstein et al. (2015), diffusion models have only recently gained popularity. As generative models, they reflect frameworks that learn probability distributions from data that are too complex to draw samples from in other ways. The training procedure is driven by iteratively adding Gaussian noise to known examples and, simultaneously, learn the way back by means of a neural network (NN). After training, sampling reflects to sample pure noise, and predict the way back to the real data distribution by means of the trained NN. Formally, one iteration adds Gaussian noise $\epsilon = N(\mu, \sigma)$ with variance σ and mean $\mu = 0$ to real data $x \in D$ sampled from Distribution D , thereby generating a sequence of ever noisier $x_t, t \in (0, T]$, referred to as the *forward process*. Eventually, x_T can no longer be distinguished from pure Gaussian noise; in parallel, the NN seeks to learn how to return from x_t to x_{t-1} for $t \in (0, T]$, which is referred to as the *reverse process*.

We introduce the auxiliary variables

$$\alpha(t) = 1 - \beta(t); \bar{\alpha}(t) = \prod_{s=1}^t \alpha_s; \tilde{\beta}(t) = \frac{1 - \bar{\alpha}(t-1)}{1 - \bar{\alpha}(t)} \beta(t) \quad (4)$$

where here β_t corresponds to the variance that refers to the noise added when moving from x_{t-1} to x_t . In practice (see Ho et al. (2020)), the β_t are pre-determined and follow a linear schedule, increasing from $\beta(1) = 10^{-4}$ to $\beta(T) = 0.02$. Steps x_{t-1} to x_t can be summarized into one formula, yielding

$$x_n(t) = \sqrt{\bar{\alpha}(t)} \cdot \epsilon + \sqrt{1 - \bar{\alpha}(t)} \cdot x \quad (5)$$

The process is designed for a data distribution D with mean $\sigma = 1$ and variance $\mu = 0$, which can easily be achieved by pre-processing steps.

In practice, the NN, E is only required to predict the noise ϵ_t that was added in each step $t \in (0, T]$; we refer to the predicted noise as ϵ_p . Formally, E , upon having received timestep t and the noisy example $x(t)$ as input, predicts

$$\epsilon_p = E(t, x(t)) \quad (6)$$

which when subtracting ϵ_p from $x(t)$ yields a denoised version of $x(t)$. Recovering the the original data x in a single step can be computed according to:

$$x_p(t, x) = \frac{x_n(t) - (1 - \sqrt{\bar{\alpha}(t)}) \cdot \epsilon_p}{\sqrt{\bar{\alpha}(t)}} \quad (7)$$

The loss function $L(x)$ of our neural network reflects to correctly predict the added noise:

$$L(x) = \|\epsilon - \epsilon_p(x)\| \quad (8)$$

where, in our case, $\|\cdot\|$ reflects the L2 norm, which is a common choice. Sampling t during training follows a uniform distribution over drawing $t \in \{1, \dots, T\}$

DATA GENERATION

Generating new data is done analogously to recovering the original data. Starting from complete noise $x_{n,T} = N(0, I)$, the NN iteratively predicts the noise to be removed, which leads to a data point $x_n = x_{n,0}$ that stems from the original distribution:

$$x_{n,t-1} = \frac{1}{\sqrt{\alpha(t)}} \cdot (x_{n,t} - \frac{1 - \alpha(t)}{\sqrt{1 - \bar{\alpha}(t)}} \epsilon_p(x_{n,t}, t)) + \sqrt{\tilde{\beta}(t)} N(0, I) \quad (9)$$

This general process, as described in detail in (Ho et al., 2020), has been repeatedly pointed out as being successful in generating realistic artificial images, for example. Applying this process to generating genotype profiles (or their embeddings), which one can easily expand into full-length genomes, establishes a novelty.

In addition to this basic process, one can further incorporate conditioning information y by changing Eq. equation 6 to

$$\epsilon_p = E(t, x_n(t), y) \quad (10)$$

Typically y reflects an input vector that specifies additional information about the data sample x . For example, in the case of an image x , this could be the caption of the image, or whether or not the image contains particularly labeled elements, such as, for example, trees or beaches as part of the

1080 image. Providing y along with x drives the generation process towards the generation of new data
1081 that takes the additional information into account, so, when following our example further, generates
1082 images that contain trees or beaches. In our work, y refers to labels that characterize human genomes
1083 in terms of population or disease phenotypes.
1084

1085
1086
1087
1088
1089
1090
1091
1092
1093
1094
1095
1096
1097
1098
1099
1100
1101
1102
1103
1104
1105
1106
1107
1108
1109
1110
1111
1112
1113
1114
1115
1116
1117
1118
1119
1120
1121
1122
1123
1124
1125
1126
1127
1128
1129
1130
1131
1132
1133