Supplementary material for the preprint Coincidence Detection Is All You Need, submitted for a double-blind peer review to 36th Conference on Neural Information Processing Systems (NeurIPS 2022). Do not distribute.

Classification by coincidence detection via Sample-Standardized Softmax regression (S3-classifier)

Here we introduce the S3-classifier, which was inspired by the theory of coincidence detection, and show it to be competitive to ResNet-26 based on data from

Ho, CS. et al. Rapid identification of pathogenic bacteria using Raman spectroscopy and deep learning. Nat Commun 10, 4927 (2019). https://doi.org/10.1038/s41467-019-12898-9

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The main results were first obtained using Wolfram Mathematica by importing the dataset as matrices using the ReadNumpy package by Luca Robbiano, and running the following analysis

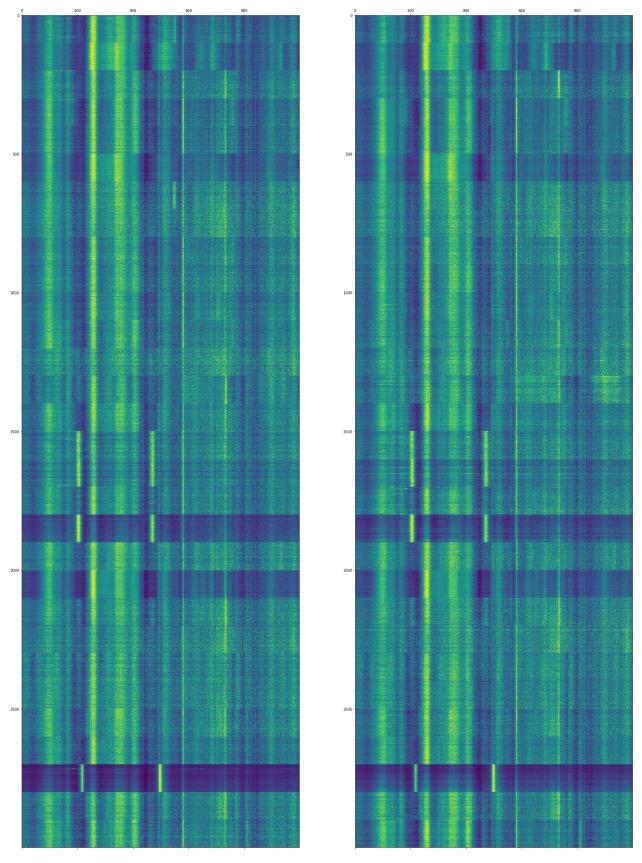
```
# Wolfram Language code to be run on the imported matrices xTrain and xTest
ssxTrain = Table[ Standardize@MeanFilter[xTrain[[L]], 1], {L, 3000}];
trainingset = Catenate@Table[ssxTrain[[(i - 1)*100 + j]] -> i, {i, 30}, {j, 100}];
c = Classify[trainingset, Method -> "LogisticRegression", PerformanceGoal -> "DirectTraining"];
ssxTest = Table[ Standardize@MeanFilter[xTest[[L]], 1] , {L, 3000}];
testset = Catenate@Table[ssxTest[[(i - 1)*100 + j]] -> i, {i, 30}, {j, 100}];
ClassifierMeasurements[c, testset]
```

Python code is provided below for a more detailed analysis, as it is presently more popular among the machine learning community.

In []: from time import time t00 = time()import numpy as np from sklearn.preprocessing import scale from sklearn.linear_model import LogisticRegression,LogisticRegressionCV import matplotlib.pyplot as plt %matplotlib inline

Visualize the train and test dataset

```
In [ ]: X_train = np.load('./drive/MyDrive/X_finetune.npy') #3000 samples of Raman spectra with 1000 Lines each
           y_train = np.load('./drive/MyDrive/y_finetune.npy') #3000 samples of bacterial classes from 0-29
X_test = np.load('./drive/MyDrive/X_test.npy')
y_test = np.load('./drive/MyDrive/y_test.npy')
           fig, (ax1,ax2) = plt.subplots(1,2,figsize=(30,50))
           ax1.matshow(X_train)
           ax2.matshow(X_test)
           plt.show()
```



Fit a softmax (multinomial logistic) regression model to the sample-standardized training dataset

In []:

sX_train = scale(X_train,axis=1) #note that standardization is performed across samples instead of across features reg = LogisticRegression(penalty='l2',C=1e4,max_iter=100,solver='lbfgs',multi_class='multinomial').fit(sX_train, y_train) reg.score(sX_train, y_train)

/usr/local/lib/python3.7/dist-packages/sklearn/linear_model/_logistic.py:818: ConvergenceWarning: lbfgs failed to converge (status=1): STOP: TOTAL NO. of ITERATIONS REACHED LIMIT.

Increase the number of iterations (max_iter) or scale the data as shown in: https://scikit-learn.org/stable/modules/preprocessing.html Please also refer to the documentation for alternative solver options: https://scikit-learn.org/stable/modules/linear_model.html#logistic-regression extra_warning_msg=_LOGISTIC_SOLVER_CONVERGENCE_MSG,

Out[]: 1.0

Evaluate the model on the standardized and smoothened test dataset

```
In []: #standardization is performed across samples instead of across features
sX_test = scale(X_test,axis=1)
#smooth dataset to account for horizontal shifts in the line spectra
def smooth(mat):
    return np.hstack((mat[::,0:1]+mat[::,1:2],mat[::,0:-2]+mat[::,1:-1]+mat[::,2:],mat[::,-1:]+mat[::,-2:-1]))
ssX_test = smooth(sX_test)
print(reg.score(ssX_test, y_test))
```

0.8293333333333334

The accuracy of our **theory-inspired model** is 82.9% and **outperforms** the 82.2% obtained by the ResNet-26 **deep learning** method.

In []: # prepare results to be plotted

y=y_test
y_hat=reg.predict(ssX_test)

The remaining analysis employs the code by Chi-Sing Ho to produce confusion matrix plots of the same style as in Ho, CS. et al (2019).

Plotting confusion matrix for bacterial isolates

We use the predictions to plot a version of the confusion matrix see in Figure 2 of the paper (Ho *et al.* 2019). Each row represents the true class and each columen represents the predicted class. The entries of the confusion matrix are normalized so that the rows sum to 100% (differences from rounding). The accuracy for each class can be seen in the diagonal entries.

```
In [ ]:
          import seaborn as sns
          from sklearn.metrics import confusion_matrix
          ORDER = [16, 17, 14, 18, 15, 20, 21, 24, 23, 26, 27, 28, 29, 25, 6, 7, 5, 3, 4,
                     9, 10, 2, 8, 11, 22, 19, 12, 13, 0, 1]
          STRAINS = {}
          STRAINS[0] = "C. albicans"
STRAINS[1] = "C. glabrata"
          STRAINS[2] = "K. aerogenes"
STRAINS[3] = "E. coli 1"
          STRAINS[4] = "E. coli 2"
          STRAINS[5] = "E. faecium"
          STRAINS[6] = "E. faecalis 1"
          STRAINS[7] = "E. faecalis 2"
          STRAINS[8] = "E. cloacae"
STRAINS[8] = "K. pneumoniae 1"
          STRAINS[10] = "K. pneumoniae 2"
STRAINS[11] = "P. mirabilis"
          STRAINS[12] = "P. aeruginosa 1"
STRAINS[13] = "P. aeruginosa 2"
          STRAINS[14] = "MSSA 1"
          STRAINS[15] = "MSSA 3"
          STRAINS[16] = "MRSA 1 (isogenic)"
          STRAINS[17] = "MRSA 2"
          STRAINS[18] = "MSSA 2"
          STRAINS[19] = "S. enterica"
          STRAINS[20] = "S. epidermidis"
          STRAINS[21] = "S. lugdunensis"
          STRAINS[22] = "S. marcescens"
          STRAINS[23] = "S. pneumoniae 2"
          STRAINS[24] = "S. pneumoniae 1"
          STRAINS[25] = "S. sanguinis"
          STRAINS[26] = "Group A Strep."
          STRAINS[27] = "Group B Strep."
          STRAINS[28] = "Group C Strep."
          STRAINS[29] = "Group G Strep."
```

MRSA 1 (isogenic) – [©]		 – MRSA 2 		MSSA 2	0,	 –S. epidermidis 		-S. pneumoniae 1	1	\triangleleft	m			o – S. sanguinis		-	o – E. faecium	o – E. coli 1	o −E. coli 2	 –K. pneumoniae 1 	 –K. pneumoniae 2 	 –K. aerogenes 		○ – P. mirabilis			 – P. aeruginosa 1 	<mark>₀</mark> – P. aeruginosa 2	 –C. albicans 	<mark>o –</mark> C. glabrata		- 100
MRSA 2 – 0		53	1	0	4	0	29	0	0	2	0	0	0	0	0	1	0	0	0	1	0	1	0	2	0	0	5	1	0	0		
MSSA 1 -		0	100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
MSSA 2 – 2		0	0	93	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
MSSA 3-4		1	0	0	59	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
S. epidermidis –			0	0	0	100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
S. lugdunensis – 🤉			0	0	1	0	97	0	0	0			0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0		- 80
S. pneumoniae 1 – o	D	0	0	0	0	0	0	93	6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0		
S. pneumoniae 2 – 🤉			0	0	0	0	0	13	80	1	0	0	0	5	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
Group A Strep. – 🤉	D	0	0	0	0	0	0	0	0	96	0	0	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
Group B Strep. – 🤉	D	0	0	0	0	0	0	0	0	0	99	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0		
Group C Strep. – 🤉		0	0	0	0	0	0	0	0	0	0	100			0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		- 60
Group G Strep. – 🤉		0	0	0	0	0	0	0	0	0	0	0	100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		00
S. sanguinis – o		0	0	0	0	0	0	0	0	1	0	1		94	0	0	0	0	0	0	0	0	0	0	2	0	2	0	0	0		
E. faecalis 1 – o			0	0	0	0	0	1	0	2	0	0		7				0	0	1	3	0	0	1	0	2	0	1	0	0		
E. faecalis 2 – o			0	0		0	1	1	0	0	0	2				77		0	0	2	0	0	1	0	3	0	2	0	0	0		
E. faecium – o		0	0	0	0	0	0	0	0	0	0	0	0	0		0			0	0	0	0	0	0	0	0	0	0	0	0		
E. coli 1 – c		0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		100		0	0	0	0	0	0	0	0	0	0	0		- 40
E. coli 2 – c			0		0	0	0	0	0	0	0	0	0	0	0	0	0		50		0	0	0	15	0	0	0	0	0	0		10
K. pneumoniae 1 – o			0			0	0	0	0	0	0	3	0	0	0	0	0		25		_	20	0	2	1	0	0	0	0	0		
K. pneumoniae 2 – C		0			0		0	0	0	0	0	0	2	0	0	0	0	2	0	1	92	0	0	2	0	1	0	0	0	0		
K. aerogenes – C E. cloacae – C		0 0	0		0 0		0	0	0	0	0	0	0	0	0	0	0	0	5	0	0	64 7		0 0	15	0	0	0	0	0		
P. mirabilis –		0	1				2		0	0	0	0	1	0 1	0	3	0	1	1	1	2		73 16	26	12 23	0	4	1	0	0		
S. marcescens –		0	0				0			0	0	0	0	0	0	4	0	-	1	0	0	2	5		86	1	4	0	0	0		- 20
S. enterica		0	0				0			0	0	0	0	0	0	4	0	0	0	0	0	2	0		0			0	0	0		
P. aeruginosa 1 – o		0	0				0		0	0	0	0	-	0	0	0	0	0	0	1	0	1	0	0	0	100	95	2	0	0		
P. aeruginosa 2 – 0		0	0				0		0			0				0	0	0	0	0	0	0	0	0	0	-	7	2 93	0	0		
C. albicans –			0			0	0	0	0	0		0			0		0	0	0	0	0	0	0	0	0	0	0	0	97			
C. glabrata – o						0	0	0	0	0		0					-	-	0	0	0	0	0	0	0	0	0	0		100	þ	
																																-0

Plotting confusion matrix for antibiotic groupings

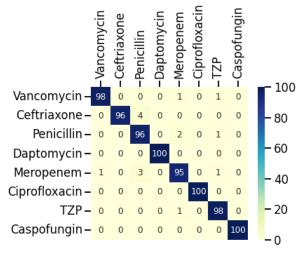
Finally, we can combine predictions into antibiotic groupings to estimate treatment accuracy. The entries of the confusion matrix are normalized so that the rows sum to 100% (differences from rounding). The accuracy for each antibiotic group can be seen in the diagonal entries.

```
In [ ]: ATCC_GROUPINGS = {3: 0,
                                 4: 0,
                                 9: 0,
10: 0,
                                 2:0,
                                 8:0,
                                 11: 0,
                                 22: 0,
12: 2,
                                 13: 2,
                                 14: 3, # MSSA
                                 18: 3, # MSSA
                                 15: 3, # MSSA
                                 20: 3,
                                 21: 3,
                                 16: 3, # isogenic MRSA
                                 17: 3, # MRSA
                                 23: 4,
                                 24: 4,
                                 26: 5,
                                  27: 5,
                                 28: 5,
                                  29: 5,
                                 25: 5,
                                 6: 5,
                                 7:5,
                                 5: 6,
19: 1,
                                 0: 7,
1: 7}
           ab_order = [3, 4, 5, 6, 0, 1, 2, 7]
antibiotics = {}
antibiotics[0] = "Meropenem" # E. coli
```

```
antibiotics[1] = "Ciprofloxacin" # SalmoneLLa
antibiotics[2] = "TZP" # PSA
antibiotics[3] = "Vancomycin" # Staph
antibiotics[4] = "Ceftriaxone" # Strep pneumo
antibiotics[5] = "Penicillin" # Strep + E. faecalis
antibiotics[6] = "Daptomycin" # E. faecium
antibiotics[7] = "Caspofungin" # Candidas
```

In []: # Mapping predictions into antibiotic groupings
y_ab = np.asarray([ATCC_GROUPINGS[i] for i in y])
y_ab_hat = np.asarray([ATCC_GROUPINGS[i] for i in y_hat])
Computing accuracy
acc = (y_ab_hat == y_ab).mean()
print('Accuracy: {:0.1f}%'.format(100*acc))

Accuracy: 96.9%



This demo was completed in: 27.12s