

# ROLL-AE: A SPATIOTEMPORAL INVARIANT AUTOENCODER FOR UNCOVERING NEURONAL ELECTROPHYSIOLOGICAL PATTERNS

**Rounak Dey, Tommaso Dreossi, Emily Fox & Daphne Koller**

insitro

South San Francisco, CA 94080, USA

{rdey, tommaso, emily, daphne}@insitro.com

## ABSTRACT

Micro-electrode array (MEA) assays enable high-throughput recording of the electrophysiological activity in biological tissues, both in vivo and in vitro. While various classical and deep learning models have been developed for MEA signal analysis, the majority focus on in vivo experiments or specific downstream applications in vitro. Consequently, extracting relevant features from in vitro MEA recordings has remained largely dependent on particular curated features known as neural metrics. In this work, we introduce Roll-AE, a novel autoencoder designed to extract meaningful spatiotemporally invariant features from in vitro MEA recordings. We demonstrate that 1) Roll-AE’s embeddings outperform those from standard autoencoders across various classification tasks, and 2) Roll-AE’s embeddings effectively characterize electrophysiological phenotypic traits in induced Pluripotent Stem Cells (iPSC)-derived neuronal cultures.

## 1 INTRODUCTION

In vitro micro-electrode array (MEA) assays provide a unique opportunity to obtain rich high-throughput electro-physiological data from induced Pluripotent Stem Cell (iPSC) (Takahashi & Yamanaka, 2006)-derived neuronal cultures (Fukushima et al., 2016; Kayama et al., 2018). MEA enables monitoring and recording of the extra-cellular action potentials in a non-invasive manner and provides valuable insights into the development and organization of neuronal networks (Novellino et al., 2011; Maeda et al., 2016). MEA has been particularly useful in the context of disease modeling, where excitability phenotypes are used to quantify the effects of genetic mutations and potential treatments of Parkinson’s disease, Amyotrophic Lateral Sclerosis (ALS), Tuberous Sclerosis Complex (TSC), etc. (Woodard et al., 2014; Wainger et al., 2014; Winden et al., 2019)

Although MEAs on in-vitro cell cultures have increased in popularity over the past decade, existing methodologies to analyse this type of data remain limited. Most existing methods are based on a collection of hand-crafted features called neural metrics (Mossink et al., 2021; Wainger et al., 2014; Kim et al., 2020). These methods (Mack et al., 2014; Bryson et al., 2022; Kapucu et al., 2022; Passaro et al., 2021) apply principal component analysis, factor analysis, or similar dimension reduction techniques to distill and analyze electro-physiological recordings. Neural metrics generally include descriptive statistics on different activity patterns such as sporadic neuronal firing, synchronous firing across electrodes, rapid consecutive firings (bursts), and bursts across multiple electrodes (network bursts). Fig. 6a lists the neural metrics curated by the commonly used Axion Biosystems (Biosystems, 2024) algorithm. Analysis of these neural metrics comes with several limitations. First, a typical MEA recording involves action-potential data on 6-96 wells per plate and 8-64 electrodes per well at a millisecond sampling rate. This high-resolution data is then compressed to 30-40 predefined metrics, many of which are redundant because they are functionally linked to other metrics. This compression often leads to significant loss of information which adversely impacts the quality of both phenotypes and disease models. Second, these neural metrics depend on multiple manually picked hyperparameters (e.g., burst threshold). This may lead to high variance with respect to different experimental conditions (e.g., culture media, seeding cell density, etc.).

Over the past decade, deep learning models have revolutionized the fields of computer vision, natural language processing, video and speech recognition (LeCun et al., 2015; Chai et al., 2021; Chai & Li, 2019). Recent applications of deep learning in neuroscience have shown promising advancement especially in processing electroencephalogram (EEG) data or in vivo recordings (van Leeuwen et al., 2019; Schirrneister et al., 2017; Buccino et al., 2018). Several recent statistical and machine-learning methods have been developed to understand the neuronal biology from live electro-physiological recordings of mouse-brains (Wu et al., 2017; Williams et al., 2020; Valente et al., 2022; Keeley et al., 2020). On passive MEA recordings of in vitro tissue cultures however, such applications are still limited. Recent works (Matsuda et al., 2022; Zhao et al., 2019), mainly deriving inspiration from computer vision, use of end-to-end convolutional neural networks (CNN) (Krizhevsky et al., 2012) on rasterized MEA signals for classifying gene knockouts and drug responses. Beyond models developed for specific tasks, foundational models such as autoencoders, or in general, feature extraction through self-supervised learning models, remains unexplored.

The MEA data modality is different from the image data modality and requires different calibration techniques. An important requirement for MEA is that the recordings remain invariant to shifts in time or changes in the orientation of the electrodes. When learning features from MEA recordings, it is important for a model to learn features that do not change if the recording is moved in time by a fixed amount, nor should they change if the spatial arrangement of the electrodes is permuted keeping the inter-electrode distances intact. These invariances are crucial as any temporal shift is determined by when the recording started and they have no biological relevance. Similarly, different spatial arrangements of the electrodes only reflect the arbitrary order in which they were arranged in the data matrix. Classical point process models (Snyder & Miller, 2012; Deutsch & Pfeifer, 1981) for analyzing spatiotemporal time-series data, in theory, can address this invariance issue. However, these models often require specific parametric assumptions, which can be overly restrictive for feature learning. Additionally, there is no widely adopted point process model for the analysis of in vitro MEA data; instead, in the broader field of neuronal electrophysiology, ad-hoc models (Amarasingham et al., 2006; Bogaard et al., 2009) are typically devised based on specific research hypotheses.

The spatiotemporal invariance for MEA recordings is analogous to orientational (rotation, translation, scale, mirror-flip) invariance of images for computer vision, where data augmentation is a standard technique of choice for model calibration (Perez & Wang, 2017). However, data augmentation does not guarantee that the encoded embeddings generated from an original image and a differently oriented image will be the same. Recent works (Burgess et al., 2024; Lohit & Trivedi, 2020) have proposed autoencoder architectures that guarantee such orientational invariance for 2D and spherical (3D) images by using orientation-equivariant convolution layers and a spatial pooling layer. Moreover, theoretical developments in the deep set literature (Zaheer et al., 2017) and group-equivariant methods (Cohen & Welling, 2016) provided necessary building blocks in understanding these invariance principles. Our work is deeply inspired by these recent developments.

In this paper, we show that augmentation techniques, even though slightly improve the model performances, still fail to learn important features for detecting subtle and complex firing patterns. For this reason, we propose Roll-AE, an novel autoencoder architecture that explicitly and completely calibrates for spatiotemporal invariance while extracting relevant features from MEA recordings. Roll-AE constructs invariant sets from given recordings and learns a set-to-set mapping with a low-dimensional bottleneck. Roll-AE is intended to be a foundational model for in vitro MEA recordings and its learned features can be used for multiple downstream tasks. We first demonstrate on a synthetic dataset that the Roll-AE embeddings have superior performance in identifying unique and complex firing patterns compared to standard autoencoders with augmentation. Then, we demonstrate multiple downstream applications of Roll-AE embeddings on an real iPSC-derived neuronal culture to illustrate that the proposed architecture captures meaningful multi-dimensional biological phenotypes useful for disease modeling and treatment discovery.

## 2 ROLL-AE

Let  $x \in \Omega \subseteq \mathbb{R}^D$  be a  $D$ -length time-series (or *spike-train*) from a feature space  $\Omega$  and let  $x_i$  be the  $i$ -th element of  $x$ . Let  $\mathbb{N}_D = \{0, 1, \dots, D-1\}$ , and  $\pi_i : \Omega \rightarrow \Omega$  for  $i \in \mathbb{N}_D$ , be a cyclic permutation function where  $\pi_i(x) = (x_{D-i+1}, x_{D-i+2}, \dots, x_D, x_1, x_2, \dots, x_{D-i})$ . Intuitively,  $\pi_i(x)$  cyclically shifts  $x$ 's elements by  $i$  positions, and we can define such cyclic permutations as

*shifts*. Let  $\Pi(x) = \{\pi_i(x) : i \in \mathbb{N}_D\}$  be the set of all  $D$  shifts of  $x$  and  $\Pi(\Omega) = \{\Pi(x) : x \in \Omega\}$  be the set of all  $\Pi(x)$  where  $x \in \Omega$ .

The standard autoencoder on the input feature space  $\Omega$  is a map  $f_{\theta, \phi} : \Omega \rightarrow \Omega$  with the encoder  $g_{\theta} : \Omega \rightarrow \mathbb{R}^k$ , decoder  $h_{\phi} : \mathbb{R}^k \rightarrow \Omega$ , and loss function  $\ell : \Omega \times \Omega \rightarrow \mathbb{R}$ . Here,  $\theta$  and  $\phi$  are parameters of the encoder and decoder, respectively. Let the output of  $g_{\theta}(x)$  to be the encoded embedding of  $x \in \Omega$  and  $\mathbb{E} = \{g_{\theta}(x) : x \in \Omega\}$  to be the embedding space.

The goal of an autoencoder is to reconstruct a given input. Specifically, the network is trained to minimize a reconstruction loss  $\ell(x, \hat{x})$  where  $\hat{x} = f_{\theta, \phi}(x) = h_{\phi}(g_{\theta}(x))$  is the forward propagation reconstruction of a given input  $x \in \Omega$ . When  $\Omega = \mathbb{R}^D$ , a commonly adopted loss function is the mean-squared loss, defined as  $\ell(x, \hat{x}) = \frac{1}{D} \sum_{i=1}^D (x_i - \hat{x}_i)^2$ .

A limitation of the standard autoencoder is that it suffers from the lack of shift-invariance. This means that two spike-trains  $x$  and  $\pi_i(x)$ , which are just shifted versions of each other, are interpreted as different spike-trains and encoded into different embeddings in the embedding space, i.e.,  $g_{\theta}(x) \neq g_{\theta}(\pi_i(x))$ . In the electro-physiology context, a shift is purely determined by when the recording started and has no biological relevance. Hence, any difference between the embeddings of two shifted spike-trains should only represent noise or potentially confounding information.

Shift-invariance in time-series data is analogous to the rotational invariance of autoencoders applied in image processing, where we want the network to learn the same encoded embeddings from different rotations of the same image. A standard way to handle such invariances is to apply augmentations (Perez & Wang, 2017; Caron et al., 2021). In the context of MEA, this means, at each training iteration, a randomly selected shifted spike-train  $\pi_i(x)$  is used as the input instead of  $x$ , and the reconstruction loss is calculated with respect to the original input  $x$ . Notice that even if augmentation encourages the network to reconstruct the original spike-train  $x$ , it does not guarantee that  $g_{\theta}(x) = g_{\theta}(\pi_i(x))$  for all  $i$ , which means the invariance problem is only partially tackled by augmentation. Drawing inspiration from recently proposed methods (Burgess et al., 2024; Lohit & Trivedi, 2020) that achieves orientation-invariance for 2D and spherical (3D) images, in the following section we introduce a novel architecture that enforces shift-invariance directly in the encoding process, ensuring that  $g_{\theta}(x) = g_{\theta}(\pi_i(x))$ .

## 2.1 ROLL-AE

A *shift-invariant* loss  $\rho : \Omega \rightarrow \Omega$  is defined such that the distance  $\rho(\pi_i(x), \pi_j(x))$  between two shifts  $\pi_i(x)$  and  $\pi_j(x)$  of the same spike-train is zero for any  $i, j \in \mathbb{N}_D$ , or in other words, the shifted spike-trains are treated as the same spike-train. This can be achieved by defining the distance metric to be  $\rho(x, x') = L(\Pi(x), \Pi(x'))$  where  $L$  is a set-based loss such as Chamfer loss (Zhang et al., 2019), Linear assignment loss (Zhang et al., 2020), etc. Therefore, shift-invariance can be achieved in an autoencoder architecture by modifying the objective, specifically, by reconstructing entire sets  $\Pi(x)$  instead of single spike-trains and back-propagating on the set-based loss function  $L$ . Since  $L$  is invariant to the ordering of the elements in a set, it guarantees  $\rho(\pi_i(x), \pi_j(x)) = L(\Pi(x), \Pi(x)) = 0$ . Formally, Roll-AE can be defined as a network  $f_{\theta, \phi} : \Pi(\Omega) \rightarrow \Pi(\Omega)$  with the encoder  $g_{\theta} : \Pi(\Omega) \rightarrow \mathbb{R}^k$ , decoder  $h_{\phi} : \mathbb{R}^k \rightarrow \Pi(\Omega)$ , and loss  $L$ .

**Encoder** Since Roll-AE is a set-to-set mapping, the encoder is constructed based on ideas from the Deep Set literature (Zaheer et al., 2017; Soelch et al., 2019; Zhang et al., 2019). Explicitly, for any  $\Pi(x) \in \Pi(\Omega)$ , the encoder is defined as  $g_{\theta}(\Pi(x)) = a(\{\tilde{g}_{\theta}(x') : x' \in \Pi(x)\})$ , where  $\tilde{g}_{\theta} : \Omega \rightarrow \mathbb{R}^k$  is a Multi-layer Perceptron (MLP) and  $a$  is an aggregation function that aggregates the set of  $k$ -dimensional outputs from  $\tilde{g}_{\theta}$  into a single  $k$ -dimensional embedding. The purpose of the aggregation function is to make the learned embeddings invariant of the shifts. Notice that  $a(\{\tilde{g}_{\theta}(x') : x' \in \Pi(x)\}) = a(\{\tilde{g}_{\theta}(x') : x' \in \Pi(\pi_i(x))\})$ , and hence  $g_{\theta}(\Pi(x)) = g_{\theta}(\Pi(\pi_i(x)))$ . Typically, the average function is used as the aggregator, although other methods can be defined (Soelch et al., 2019). Roll-AE uses average as aggregation function.

**Decoder** The decoder is defined as  $h_{\phi}(e) = \Pi(\tilde{h}_{\phi}(e))$  for  $e \in \mathbb{R}^k$ , where  $\tilde{h}_{\phi} : \mathbb{R}^k \rightarrow \Omega$  is an MLP. A crucial challenge in constructing Deep Set autoencoders is finding a suitable mapping from the output of  $\tilde{h}_{\phi}$  to the space of sets. Roll-AE does not face this challenge as this mapping is

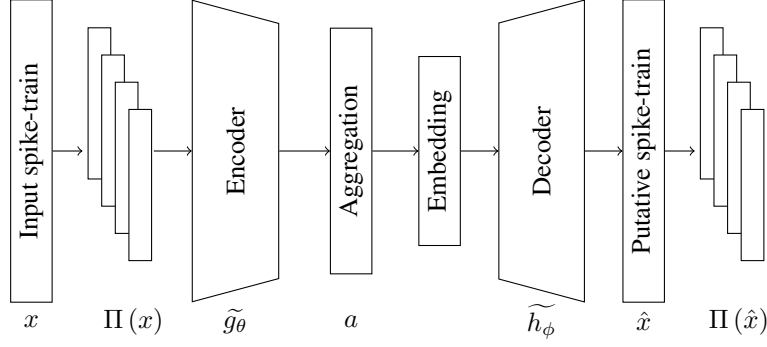


Figure 1: Roll-AE architecture. The input spike-train  $x$  is converted into its cyclic permutation set  $\Pi(x)$  which is passed to the encoder  $\tilde{g}_\theta$ . The encoder outputs are grouped by the aggregation function  $a$  into a single embedding. The decoder  $\tilde{h}_\phi$  reconstructs the putative spike-train  $\hat{x}$  which is converted into its cyclic permutation set  $\Pi(\hat{x})$ . The reconstruction loss is computed between  $\Pi(x)$  and  $\Pi(\hat{x})$ .

deterministic and known. Therefore, the forward propagation of the overall Roll-AE architecture is defined as the mapping  $f_{\theta,\phi}(\Pi(x)) = \Pi(\tilde{h}_\phi(a(\{\tilde{g}_\theta(x') : x' \in \Pi(x)\})))$ . Fig. 1 shows Roll-AE architecture. We note that the trainable parameters in Roll-AE are all contained in the encoder MLP  $\tilde{g}_\theta$  and the decoder MLP  $\tilde{h}_\phi$ , which are functions that simply map  $\Omega$  onto  $\mathbb{R}^k$  and back. As a result, the number of trainable parameters in Roll-AE remains the same as a standard autoencoder.

**Reconstruction Loss** Roll-AE uses the Linear Assignment loss as the reconstruction loss between  $X = \Pi(x)$  and  $X' = f_{\theta,\phi}(\Pi(x))$ . Specifically, for any arbitrary ordering of the elements  $X = \{x^{(1)}, x^{(2)}, \dots, x^{(D)}\}$  and  $X' = \{x'^{(1)}, x'^{(2)}, \dots, x'^{(D)}\}$ , and denoting  $\Psi$  to be the set of all possible permutations (not just cyclic) of  $(1, 2, \dots, D)$ , the Linear Assignment loss is defined as  $L(X, X') = \min_{\psi \in \Psi} \sum_{i \in \mathbb{N}_D} \ell(x^{(i)}, x'^{\psi(i)})$ .

Computing the Linear assignment loss, in general, is extremely expensive with complexity  $O(s^3)$  using the Hungarian algorithm, where  $s$  is the cardinality of  $X$  (in this context,  $s = D$ ). However, in our problem, since both the sets  $X$  and  $X'$  are closed under the cyclic permutation operation, the computation can be substantially improved to  $O(r)$  complexity where  $r$  is the order of the cyclic permutation operation (in this context,  $r = s = D$ ). Lemma A.1 then simplifies this reconstruction loss as  $L(\Pi(x), \Pi(\hat{x})) = D[\min_{i \in \mathbb{N}_D} \ell(\pi_i(x), \hat{x})]$ , where  $\hat{x} = \tilde{h}_\phi(a(\{\tilde{g}_\theta(x') : x' \in \Pi(x)\}))$  is the putative output train. From Lemma A.1, the above expression also applies to Chamfer loss.

**Stochastic Shift-invariance** Roll-AE further implements stochastic shift-invariance, i.e., instead of passing all possible shifted spike-trains in  $\Pi(x)$  on each forward propagation iteration, it uniformly samples spike-trains from  $\Pi(x)$  with sampling rate  $\tau$  and uses the set of sampled spike-trains as inputs. Once the model is trained, the final embeddings can then be calculated by running a final forward propagation of the encoder with the entire  $\Pi(x)$  as input. This ensures the invariance in the final embeddings remains intact. In Appendix C, we demonstrate through extensive simulation studies that stochastic shift-invariance can perform as good or sometimes even better than complete shift-invariance ( $\tau = 1$ ) while reducing the memory requirement and computation time.

**Generic Permutations** Roll-AE is not limited to temporal shifts or permutation of input spike-trains. The above definitions and properties hold as long as  $\Pi(x)$  is a closed group with respect to any class of cyclic permutations. This allows Roll-AE to handle other interesting classes of permutation that are relevant for our biological assays. For instance, mirror symmetries can be used to permute electrodes in a well and enforce spatial invariance on our embeddings. When multiple spike-trains are simultaneously recorded by different electrodes placed on the same cell culture, they need to be arranged and inputted into the model in a specific reference order. However, this order is arbitrary, and any permutation should be biologically equivalent to the reference order as long as the inter-electrode distances are preserved. Such permutations are given by the mirror flip operation. Fig. 2a shows an image of an actual eight-electrode well used to record neuronal electrophysiological activity using an

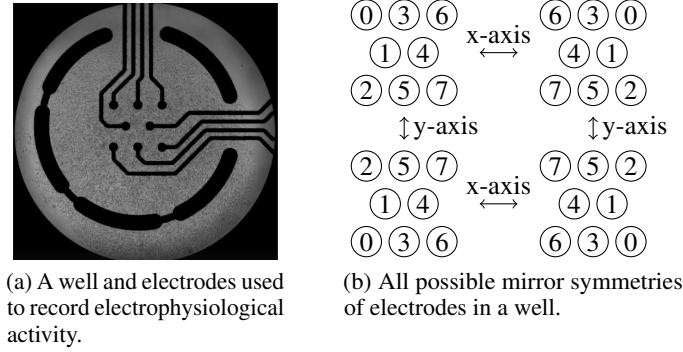


Figure 2: Electrophysiological activity and electrode symmetry in a well. Left: Axon Biosystems image showing 3-2-3 electrode arrangement. Right: Possible electrode symmetries with indices. Electrode labels follow the top left pattern.

Axon Biosystems (Biosystems, 2024) MEA machine. Fig. 2b shows all possible electrode orderings with the top-left graph matching the reference orientation. Notice that the four permutations defined by mirror symmetry form a closed group under the mirror flip operation which further is a special case of cyclic permutations.

### 3 EVALUATIONS

#### 3.1 SYNTHETIC DATA EVALUATION

We simulate recordings with spike-trains on eight electrodes. Our synthetic dataset has four tunable parameters that determine the level of activity along four different types of source-events, namely sporadic spikes (spike), sporadic single-channel bursts (burst), cyclic bursts (cycle), and network bursts (network). These parameters are formally defined as follows: *spike*: the probability of a sporadic firing event at a particular time instance; *burst*: the probability of a sequence of firing events starting at a particular time instance for a random duration; *cycle*: presence or absence of a repeating pattern of sequence of firing events with a random phase and duration; *network*: presence or absence of network bursts where sequences of firing events are recorded simultaneously across multiple electrodes. Each network burst starts with an originating electrode, and the probability of observing firing events in other electrodes depend on their proximity to the originating electrode. Fig. 7 depicts some examples of firing patterns where one type of firing event was kept at a high probability (or present) and the others are kept at a low probability (or absent). Details of the data generating model for the synthetic data are presented in Appendix B.

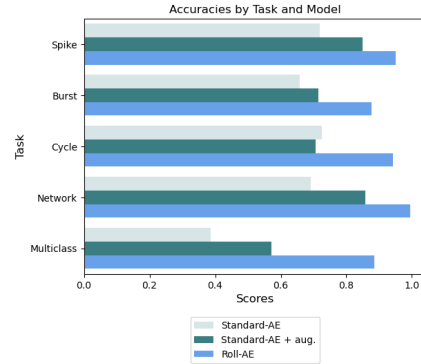


Figure 3: Roll-AE consistently outperforms standard autoencoders (both with and without augmentation) on all binary and multiclass classification tasks. Accuracy comparison between autoencoders across tasks. Roll-AE achieves highest accuracy across all binary and multiclass tasks.

**Model Training** We trained three models: a standard autoencoder, a standard autoencoder with augmentation, and Roll-AE on the synthetic dataset. The hyperparameters (such as training batch-size, embedding dimension etc.) for each model were selected based on a two-step training/validation scheme (see Appendix D for training details). The embeddings generated by the trained models were then compared based on downstream classification tasks.

We trained five classifiers (with 70/30% training/validation data split) using the embeddings from each autoencoder: four binary classifiers (e.g., high vs low probability of spikes, high vs low probability of a bursts, etc.) and a multi-class classifier with 16 classes encompassing all combinations of our synthetic dataset parameters (e.g., low spike, low burst, no cycle, no network vs low spike, low burst, no cycle, present network vs low spike, low burst, present cycle, present network, and so on). Specifically, we trained logistic regressors with L2 regularization with the penalty parameter determined using a 4-fold cross-validation. The best trained logistic regressor was then used to make predictions on the validation data.

**Results** Fig. 3 reports the obtained accuracies together with the confusion matrices for the multi-class task. Overall, Roll-AE outperforms the standard autoencoders across all classification tasks (see Fig. 3). Remarkably, Roll-AE achieved a +30% accuracy on the hardest multi-class task. It is also worth noticing, that augmentation improves the standard autoencoder across all tasks (except *cycle*), while still being sensibly inferior to Roll-AE. Fig. 9 reports the confusion matrices of the three models for the multiclass task. These plots highlight how Roll-AE embeddings lead to a high predictive accuracy in our downstream classification tasks while the two standard autoencoders tend to misclassify similar classes in a multi-class regime.

### 3.2 siRNA TREATMENT EVALUATION

As a second case study, we applied the three models on a real electrophysiological dataset obtained from induced Pluripotent Stem Cells (iPSC) (Takahashi & Yamanaka, 2006) derived neurons subject to Small Interfering RNA (siRNA) (Fire et al., 1998) treatments. Technically, iPSCs are cells that have been reprogrammed from skin or blood cells to become other types of cells, in our case, neurons. siRNAs are artificially synthesized RNA molecules commonly used in molecular biology for silencing genes of interest. In our case, we apply a double siRNA treatment: the first to silence a gene to trigger the disease, the second to investigate a possible cure to counteract the effect of the first intervention.

Our neuronal culture was organized on two 96-well plates. Half of the sample set was subject to a siRNA knockdown (siKD) of the gene of interest, mimicking the effect of the considered disease, while the other half was subject to a non-targeting control siRNA sequence (NTS) designed to target no known genes. The NTS treatment is our negative control, i.e., a condition that does not affect the neuron state. Both siKD and NTS samples have been then treated with 24 different siRNA potential cures. Among these, there is an additional negative control NTS and a positive control CTRL+ known to reduce neuronal excitability. For each condition, we cultured 4 replicates recorded at nine different days up to 24 days in vitro. In total we obtained  $2 \text{ (siKD or NTS)} \times 24 \text{ (siRNA treatments)} \times 4 \text{ (replicates)} \times 9 \text{ (days in vitro)} \times 8 \text{ (electrodes)} = 13824$  raw spike-trains. Each spike-train was recorded with one milli-second sampling rate for 10 minutes. We bucketized the raw spike-trains to 500 milli-second bins to end up with spike-train length of  $D = 1200$ . To train the models, we performed a hyperparameter selection of training batch-size, embedding dimension, learning rate, and shift-sampling rate (see Appendix D). Models with the selected hyperparameters were then trained to generate the final embeddings. First, we compare the embeddings on an siKD phenotyping task.

**Results - siKD Phenotyping** We compared the embeddings from the three models based on their performance in the classification of siKD wells from NTS wells on each of the 24 siRNA treatments. Biologically, the effect of siKD is very subtle, and discovering a classifier with good classification accuracy helps uncover subtle phenotypes in our disease model. We applied a leave-one-well-out approach, where at each iteration, we left out all the recordings from a particular well with a particular siRNA treatment, trained a siKD vs NTS classifier on the rest of the wells, and predicted the recordings of the left-out well. Logistic regressors with L2 regularization and 4-fold cross-validation were used as the classifiers.

Classification accuracies are presented in Fig. 4. Roll-AE embeddings achieved the highest accuracy in 15 out of the 24 treatments, and most crucially, Roll-AE exhibited the highest accuracy for NTS. It is important to highlight that the effect of siKD is particularly subtle and has only been previously identified in NTS. For other siRNA knockdowns, the influence of siKD remains uncertain. Ideally, when targeting a gene with potential as a drug target, the knockdown should mitigate the effects of siKD, restoring the condition to the NTS level. Next, we demonstrate the application of the Roll-AE embeddings on two downstream tasks: treatment clustering and metrics credentialing.

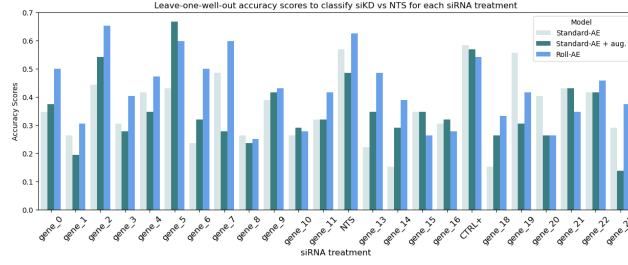


Figure 4: Accuracy comparison between autoencoders for classification of siKD vs NTS on different siRNA treatments. Roll-AE has the highest accuracy for 15 out of the 24 treatments.

**Results - Treatment Clustering** For this study, we used Roll-AE embeddings to extract biological insights and characterize treatment similarities. To do so, we considered cells cultured for 24 days in vitro, reduced the dimension of our embeddings to 10 principal components explaining at least 95% of the variance, computed the centroids of each treatment cluster, and calculated the pairwise distances between treatment centroids. The obtained results for NTS and siKD treated cells are organized in the two dendrograms shown in Fig. 5a and Fig. 5b, respectively.

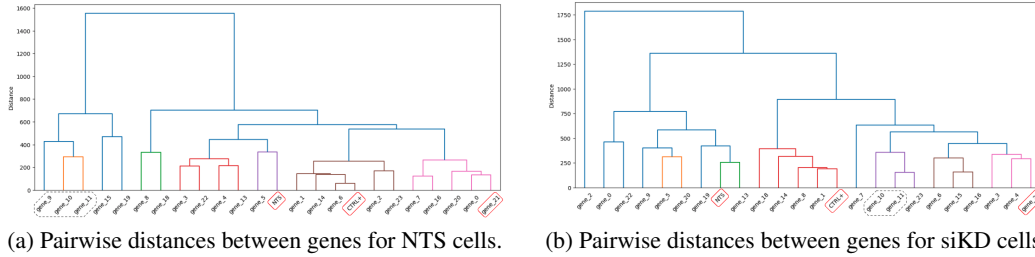


Figure 5: Dendrograms of embedding pairwise distances between gene centroids applied to unperturbed (NTS) and diseased (siKD) cells.

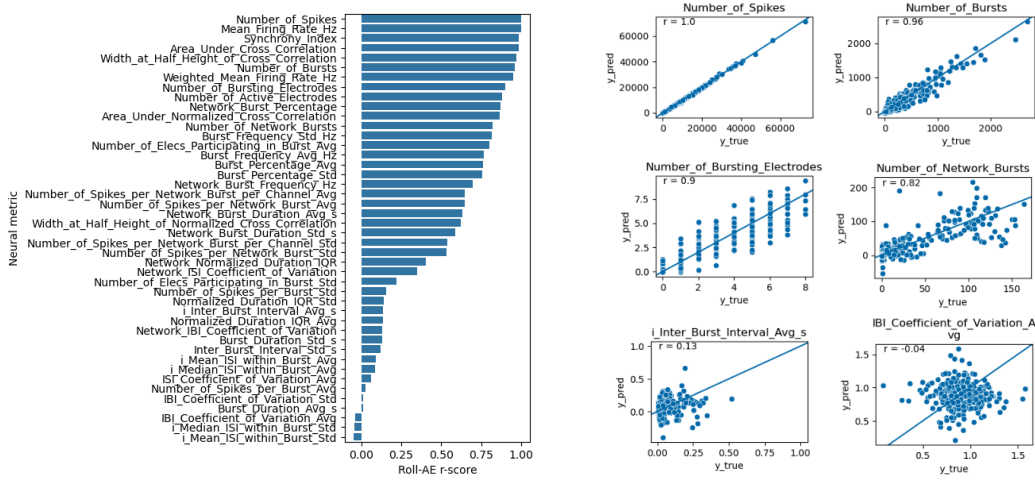
For cells with NTS, we observe a clear distinction between CTRL+ and NTS treatments, indicating that the model effectively differentiates CTRL+ (known to reduce cell excitability) from unperturbed NTS cells. On the other hand, gene\_21, which clusters far from CTRL+, is known for inducing hyperexcitability and stimulating neuronal activity. gene\_9, gene\_10, and gene\_11, which clustered together, are all components of the same signaling pathway involved in stress responses.

For cells with siKD, we observe again that NTS and CTRL+ treatments form distinct clusters, with gene\_21 clustering on the opposite side of the hyperexcitability spectrum of the dendrogram. Interestingly, gene\_10 and 11 are still clustering together but gene\_9 does not. Additionally, gene\_3 and gene\_4, which were grouped as hyperexcitable in the NTS cells study, now cluster among hypoexcitable treatments. This suggests that these genes interact differently with the induced disease state, leading to different types of neuronal activity depending on the disease state of the cell.

**Results - Neural Metrics Credentialing** Finally, we verify whether the obtained embeddings retain relevant information on the curated metrics provided by the Axon Biosystems instrument that are explicitly computed from raw recordings. The full metric list is reported in Fig. 6a.

To assess the ability of our embeddings to retain relevant information on these curated neural metrics, for each metric, we trained a Ridge regressor with an  $L2$  penalty of 1.0. This was trained on 80% of the entire dataset and then used to predict each metric on the remaining 20%. The correlation (r-score) between the predicted and observed neuronal metrics from the validation data is shown in Fig. 6a. The metrics are ranked from the highest to the lowest correlation.

Roll-AE embeddings predict most metrics with high correlation. Out of 44 regressed metrics, 25 had r-score above 0.75, 9 between 0.25 and 0.75, and 10 between -0.1 and 0.25. Fig. 6b illustrates some scatter plots of actual metric values against the predicted ones. We observed that metrics



(a) r-scores between observed and predicted neural metrics from Roll-AE embeddings. High r-scores indicate Roll-AE’s embedding ability to capture explicit neural metrics.

(b) Examples of observed vs predicted neural metrics from Roll-AE embeddings with high (top row) and low (bottom row) r-scores.

Figure 6: Credentialing Roll-AE by evaluating its performance in predicting the neural metrics.

useful for phenotypic analyses, including firing, spike, or burst counts and rates, are accurately captured and predicted. However, certain metrics, such as those related to the inter-burst interval (IBI coefficient) (Di Credico et al., 2021), demonstrated lower correlations. We hypothesize that this discrepancy may be attributed to the selected binning size during the compression of the raw signal, which can potentially eliminate inter-burst information.

## 4 CONCLUSION

We presented Roll-AE, a novel spatiotemporal invariant autoencoder for feature extraction from passive recordings of MEA assays. By imposing invariances, Roll-AE extracts features that are relevant for identifying unique firing patterns. On synthetic data, we demonstrated that the Roll-AE embeddings far outperform standard autoencoders (with or without augmentation) in discriminating different individual source-events. Roll-AE is particularly accurate in the multi-class classification task and showed a +30% accuracy gain compared to standard autoencoders suggesting Roll-AE’s ability to identify features relevant to complex and subtle phenotypes. On the siRNA experiment data, we demonstrated the superior performance of Roll-AE embeddings in discriminating siKD from NTS highlighting its use in phenotype discovery. The concordance of the relative clustering of treatments with biological evidences supports the validity of these machine learnt features. Finally, we showed that these embeddings retain explicit metrics and can be used to predict manually curated features.

The original formulation of Roll-AE came with a few limitations, mainly in regard to computational efficiency. Constructing the entire set of cyclic permutations and evaluating the Linear Assignment loss can be computationally demanding. To tackle this issue, we have proposed the stochastic shift-invariance approach (Appendix C). In our siRNA experiment example, we found that the embeddings were not able to predict some neural metrics well, predominantly those with inter-spike interval-related metrics. This could be a consequence of the adopted 500 milli-seconds bin size. While we have demonstrated its efficacy using in vitro MEA data, the autoencoder design could be easily generalized for other types of data where such spatiotemporal invariance is relevant. Other potential use cases for such architectures would be to identify arrhythmia from ECG data and anomaly detection from sensors.

In conclusion, Roll-AE provides a foundational model for extracting features from in vitro MEA recordings. The features from Roll-AE enables better identification of unique electro-physiological activity patterns from MEA recordings, and can be used for a multitude of downstream applications.



## MEANINGFULNESS STATEMENT

In vitro micro-electrode array (MEA) offers powerful insights into neuronal network dynamics, especially for disease modeling. By providing high-throughput monitoring of electrophysiological activity, MEA serves as a link between neuroscience and therapeutic research. Our paper introduces Roll-AE, a novel autoencoder architecture that addresses spatiotemporal invariance in MEA recordings. This approach enhances feature extraction, enabling the representation of multidimensional biological phenotypes essential for disease modeling and treatment exploration. The embeddings generated by Roll-AE offer a robust representation of complex neuronal behaviors, making them an effective tool for understanding complex biological processes in neurons.

## REFERENCES

- Asohan Amarasingham, Ting-Li Chen, Stuart Geman, Matthew T Harrison, and David L Sheinberg. Spike count reliability and the poisson hypothesis. *J. Neurosci.*, 26(3):801–809, January 2006.
- Axion Biosystems. Axion biosystems, 2024. URL <https://www.axionbiosystems.com/>. [Online; accessed 22-April-2024].
- Andrew Bogaard, Jack Parent, Michal Zochowski, and Victoria Booth. Interaction of cellular and network mechanisms in spatiotemporal pattern formation in neuronal networks. *J. Neurosci.*, 29(6):1677–1687, February 2009.
- Alexander Bryson, Dulini Mendis, Emma Morrisroe, Christopher A. Reid, Saman Halgamuge, and Steven Petrou. Classification of antiseizure drugs in cultured neuronal networks using multielectrode arrays and unsupervised learning. *Epilepsia*, 63(7):1693–1703, 2022. doi: <https://doi.org/10.1111/epi.17268>. URL <https://onlinelibrary.wiley.com/doi/abs/10.1111/epi.17268>.
- Alessio P Buccino, Torbjorn V Ness, Gaute T Einevoll, Gert Cauwenberghs, and Philipp D Hafliger. A deep learning approach for the classification of neuronal cell types. *Annu. Int. Conf. IEEE Eng. Med. Biol. Soc.*, 2018:999–1002, July 2018.
- James Burgess, Jeffrey J Nirshchl, Maria-Clara Zanellati, Alejandro Lozano, Sarah Cohen, and Serena Yeung-Levy. Orientation-invariant autoencoders learn robust representations for shape profiling of cells and organelles. *Nature Communications*, 15(1):1022, February 2024.
- Mathilde Caron, Hugo Touvron, Ishan Misra, Hervé Jégou, Julien Mairal, Piotr Bojanowski, and Armand Joulin. Emerging properties in self-supervised vision transformers, 2021.
- Junyi Chai and Anming Li. Deep learning in natural language processing: A state-of-the-art survey. In *2019 International Conference on Machine Learning and Cybernetics (ICMLC)*, pp. 1–6, 2019. doi: 10.1109/ICMLC48188.2019.8949185.
- Junyi Chai, Hao Zeng, Anming Li, and Eric W.T. Ngai. Deep learning in computer vision: A critical review of emerging techniques and application scenarios. *Machine Learning with Applications*, 6: 100134, 2021. ISSN 2666-8270. doi: <https://doi.org/10.1016/j.mlwa.2021.100134>. URL <https://www.sciencedirect.com/science/article/pii/S2666827021000670>.
- Taco Cohen and Max Welling. Group equivariant convolutional networks. In Maria Florina Balcan and Kilian Q. Weinberger (eds.), *Proceedings of The 33rd International Conference on Machine Learning*, volume 48 of *Proceedings of Machine Learning Research*, pp. 2990–2999, New York, New York, USA, 20–22 Jun 2016. PMLR. URL <https://proceedings.mlr.press/v48/cohen16.html>.
- Stuart Jay Deutsch and Phillip E. Pfeifer. Space-time arma modeling with contemporaneously correlated innovations. *Technometrics*, 23(4):401–409, 1981. ISSN 00401706. URL <http://www.jstor.org/stable/1268231>.
- Andrea Di Credico, Giulia Gaggi, Pascal Izzicupo, Laura Ferri, Laura Bonanni, Giovanni Iannetti, Angela Di Baldassarre, and Barbara Ghinassi. Real-time monitoring of levetiracetam effect on the electrophysiology of an heterogenous human ipsc-derived neuronal cell culture using microelectrode array technology. *Biosensors*, 11(11):450, 2021.

- Andrew Fire, SiQun Xu, Mary K Montgomery, Steven A Kostas, Samuel E Driver, and Craig C Mello. Potent and specific genetic interference by double-stranded rna in *caenorhabditis elegans*. *nature*, 391(6669):806–811, 1998.
- Kazuyuki Fukushima, Yuji Miura, Kohei Sawada, Kazuto Yamazaki, and Masashi Ito. Establishment of a human neuronal network assessment system by using a human neuron/astrocyte co-culture derived from fetal neural stem/progenitor cells. *J. Biomol. Screen.*, 21(1):54–64, January 2016.
- Fikret Emre Kapucu, Andrey Vinogradov, Tanja Hyvärinen, Laura Ylä-Outinen, and Susanna Narkilahti. Comparative microelectrode array data of the functional development of hPSC-derived and rat neuronal networks. *Scientific Data*, 9(1):120, March 2022.
- Tasuku Kayama, Ikuro Suzuki, Aoi Odawara, Takuya Sasaki, and Yuji Ikegaya. Temporally coordinated spiking activity of human induced pluripotent stem cell-derived neurons co-cultured with astrocytes. *Biochem. Biophys. Res. Commun.*, 495(1):1028–1033, January 2018.
- Stephen L Keeley, David M Zoltowski, Mikio C Aoi, and Jonathan W Pillow. Modeling statistical dependencies in multi-region spike train data. *Current Opinion in Neurobiology*, 65:194–202, 2020. ISSN 0959-4388. doi: <https://doi.org/10.1016/j.conb.2020.11.005>. URL <https://www.sciencedirect.com/science/article/pii/S0959438820301720>. Whole-brain interactions between neural circuits.
- Byung Woo Kim, Jiwon Ryu, Ye Eun Jeong, Juhyun Kim, and Lee J Martin. Human motor neurons with SOD1-G93A mutation generated from CRISPR/Cas9 gene-edited iPSCs develop pathological features of amyotrophic lateral sclerosis. *Front. Cell. Neurosci.*, 14:604171, November 2020.
- Diederik P. Kingma and Jimmy Ba. Adam: A method for stochastic optimization, 2017.
- Alex Krizhevsky, Ilya Sutskever, and Geoffrey E Hinton. Imagenet classification with deep convolutional neural networks. In F. Pereira, C.J. Burges, L. Bottou, and K.Q. Weinberger (eds.), *Advances in Neural Information Processing Systems*, volume 25. Curran Associates, Inc., 2012. URL [https://proceedings.neurips.cc/paper\\_files/paper/2012/file/c399862d3b9d6b76c8436e924a68c45b-Paper.pdf](https://proceedings.neurips.cc/paper_files/paper/2012/file/c399862d3b9d6b76c8436e924a68c45b-Paper.pdf).
- Yann LeCun, Yoshua Bengio, and Geoffrey Hinton. Deep learning. *Nature*, 521(7553):436–444, May 2015.
- Suhas Lohit and Shubhendu Trivedi. Rotation-invariant autoencoders for signals on spheres, 2020.
- Cina M. Mack, Bryant J. Lin, James D. Turner, Andrew F.M. Johnstone, Lyle D. Burgoon, and Timothy J. Shafer. Burst and principal components analyses of mea data for 16 chemicals describe at least three effects classes. *NeuroToxicology*, 40:75–85, 2014. ISSN 0161-813X. doi: <https://doi.org/10.1016/j.neuro.2013.11.008>. URL <https://www.sciencedirect.com/science/article/pii/S0161813X13001800>.
- Hiroshi Maeda, Tomohiro Chiyonobu, Michiko Yoshida, Satoshi Yamashita, Masashi Zuiki, Satoshi Kidowaki, Kenichi Isoda, Kazuhiro Yamakawa, Masafumi Morimoto, Tatsutoshi Nakahata, Megumu K Saito, and Hajime Hosoi. Establishment of isogenic iPSCs from an individual with SCN1A mutation mosaicism as a model for investigating neurocognitive impairment in dravet syndrome. *J. Hum. Genet.*, 61(6):565–569, June 2016.
- N Matsuda, A Odawara, K Kinoshita, A Okamura, T Shirakawa, and I Suzuki. Raster plots machine learning to predict the seizure liability of drugs and to identify drugs. *Scientific Reports*, 12(1): 2281, February 2022.
- Britt Mossink, Anouk H A Verboven, Eline J H van Hugte, Teun M Klein Gunnewiek, Giulia Parodi, Katrin Linda, Chantal Schoenmaker, Tjitske Kleefstra, Tamas Kozicz, Hans van Bokhoven, Dirk Schubert, Nael Nadif Kasri, and Monica Frega. Human neuronal networks on micro-electrode arrays are a highly robust tool to study disease-specific genotype-phenotype correlations in vitro. *Stem Cell Reports*, 16(9):2182–2196, September 2021.

- A Novellino, Bibiana Scelfo, T Palosaari, A Price, Tomasz Sobanski, T J Shafer, A F M Johnstone, G W Gross, A Gramowski, O Schroeder, K Jügel, M Chiappalone, F Benfenati, S Martinoia, M T Tedesco, E Defranchi, P D' Angelo, and M Whelan. Development of micro-electrode array based tests for neurotoxicity: assessment of interlaboratory reproducibility with neuroactive chemicals. *Front. Neuroeng.*, 4:4, April 2011.
- Austin P Passaro, Onur Aydin, M Taher A Saif, and Steven L Stice. Development of an objective index, neural activity score (NAS), reveals neural network ontogeny and treatment effects on microelectrode arrays. *Scientific Reports*, 11(1):9110, April 2021.
- Luis Perez and Jason Wang. The effectiveness of data augmentation in image classification using deep learning. *arXiv preprint arXiv:1712.04621*, 2017.
- Robin Tibor Schirrmester, Jost Tobias Springenberg, Lukas Dominique Josef Fiederer, Martin Glasstetter, Katharina Eggensperger, Michael Tangermann, Frank Hutter, Wolfram Burgard, and Tonio Ball. Deep learning with convolutional neural networks for EEG decoding and visualization. *Hum. Brain Mapp.*, 38(11):5391–5420, November 2017.
- D.L. Snyder and M.I. Miller. *Random Point Processes in Time and Space*. Springer Texts in Electrical Engineering. Springer New York, 2012. ISBN 9781461231660. URL [https://books.google.com/books?id=c\\_3UBwAAQBAJ](https://books.google.com/books?id=c_3UBwAAQBAJ).
- Maximilian Soelch, Adnan Akhundov, Patrick van der Smagt, and Justin Bayer. *On Deep Set Learning and the Choice of Aggregations*, pp. 444–457. Springer International Publishing, 2019. ISBN 9783030304874. doi: 10.1007/978-3-030-30487-4\_35. URL [http://dx.doi.org/10.1007/978-3-030-30487-4\\_35](http://dx.doi.org/10.1007/978-3-030-30487-4_35).
- Kazutoshi Takahashi and Shinya Yamanaka. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *cell*, 126(4):663–676, 2006.
- Adrian Valente, Jonathan W. Pillow, and Srdjan Ostojic. Extracting computational mechanisms from neural data using low-rank RNNs. In Alice H. Oh, Alekh Agarwal, Danielle Belgrave, and Kyunghyun Cho (eds.), *Advances in Neural Information Processing Systems*, 2022. URL <https://openreview.net/forum?id=M12autRxeeS>.
- K G van Leeuwen, H Sun, M Tabaeizadeh, A F Struck, M J A M van Putten, and M B Westover. Detecting abnormal electroencephalograms using deep convolutional networks. *Clin. Neurophysiol.*, 130(1):77–84, January 2019.
- Brian J Wainger, Evangelos Kiskinis, Cassidy Mellin, Ole Wiskow, Steve S W Han, Jackson Sandoe, Numa P Perez, Luis A Williams, Seungkyu Lee, Gabriella Boulting, James D Berry, Robert H Brown, Jr, Merit E Cudkowicz, Bruce P Bean, Kevin Eggan, and Clifford J Woolf. Intrinsic membrane hyperexcitability of amyotrophic lateral sclerosis patient-derived motor neurons. *Cell Rep.*, 7(1):1–11, April 2014.
- Alex H. Williams, Anthony Degleris, Yixin Wang, and Scott W. Linderman. Point process models for sequence detection in high-dimensional neural spike trains, 2020.
- Kellen D Winden, Maria Sundberg, Cindy Yang, Syed M A Wafa, Sean Dwyer, Pin-Fang Chen, Elizabeth D Buttermore, and Mustafa Sahin. Biallelic mutations in TSC2 lead to abnormalities associated with cortical tubers in human iPSC-derived neurons. *J. Neurosci.*, 39(47):9294–9305, November 2019.
- Chris M Woodard, Brian A Campos, Sheng-Han Kuo, Melissa J Nirenberg, Michael W Nestor, Matthew Zimmer, Eugene V Mosharov, David Sulzer, Hongyan Zhou, Daniel Paull, Lorraine Clark, Eric E Schadt, Sergio Pablo Sardi, Lee Rubin, Kevin Eggan, Mathew Brock, Scott Lipnick, Mahendra Rao, Stephen Chang, Aiqun Li, and Scott A Noggle. ipsc-derived dopamine neurons reveal differences between monozygotic twins discordant for parkinson’s disease. *Cell Rep.*, 9(4): 1173–1182, November 2014.
- Anqi Wu, Nicholas A Roy, Stephen Keeley, and Jonathan W Pillow. Gaussian process based nonlinear latent structure discovery in multivariate spike train data. *Adv. Neural Inf. Process. Syst.*, 30:3496–3505, December 2017.

Manzil Zaheer, Satwik Kottur, Siamak Ravanbakhsh, Barnabas Poczos, Russ R Salakhutdinov, and Alexander J Smola. Deep sets. In I. Guyon, U. Von Luxburg, S. Bengio, H. Wallach, R. Fergus, S. Vishwanathan, and R. Garnett (eds.), *Advances in Neural Information Processing Systems*, volume 30. Curran Associates, Inc., 2017. URL [https://proceedings.neurips.cc/paper\\_files/paper/2017/file/f22e4747dalaa27e363d86d40ff442fe-Paper.pdf](https://proceedings.neurips.cc/paper_files/paper/2017/file/f22e4747dalaa27e363d86d40ff442fe-Paper.pdf).

Yan Zhang, Jonathon Hare, and Adam Prugel-Bennett. Deep set prediction networks. *Advances in Neural Information Processing Systems*, 32, 2019.

Yan Zhang, Jonathon Hare, and Adam Prügel-Bennett. Fspool: Learning set representations with featurewise sort pooling. In *International Conference on Learning Representations*, 2020. URL <https://openreview.net/forum?id=HJgBA2VYwH>.

Yun Zhao, Elmer Guzman, Morgane Audouard, Zhuowei Cheng, PaulK. Hansma, Kenneth S. Kosik, and Linda Petzold. A deep learning framework for classification of in vitro multi-electrode array recordings, 2019.

## A MATHEMATICAL RESULTS

**Lemma A.1.** For two sets  $\Pi(x)$  and  $\Pi(y)$ ;  $x, y \in \Omega$ ,

$$\min_{\psi \in \Psi} \sum_{i \in \mathbb{N}_D} \ell(\pi_i(x), \pi_{\psi(i)}(y)) = D \times \left[ \min_{i \in \mathbb{N}_D} \ell(\pi_i(x), y) \right].$$

*Proof.* First, we note that, for any  $i \in \mathbb{N}_D, \psi \in \Psi$ ,

$$\ell(\pi_i(x), \pi_{\psi(i)}(y)) \geq \min_{j, j' \in \mathbb{N}_D} \ell(\pi_j(x), \pi_{j'}(y)),$$

which implies

$$\min_{\psi \in \Psi} \sum_{i \in \mathbb{N}_D} \ell(\pi_i(x), \pi_{\psi(i)}(y)) \geq D \times \left[ \min_{j, j' \in \mathbb{N}_D} \ell(\pi_j(x), \pi_{j'}(y)) \right]. \quad (1)$$

Lets denote  $(j^*, j'^*) = \arg \min_{j, j'} \ell(\pi_j(x), \pi_{j'}(y))$ . Further, let the permutation  $\psi^* \in \Psi$  to be a cyclic permutation that maps  $j^*$  to  $j'^*$ , explicitly,  $\psi^*(j^*) = j'^*$ . Then,

$$\begin{aligned} D \times \left[ \min_{j, j' \in \mathbb{N}_D} \ell(\pi_j(x), \pi_{j'}(y)) \right] &= D \ell(\pi_{j^*}(x), \pi_{j'^*}(y)) \\ &= D \ell(\pi_{j^*}(x), \pi_{\psi^*(j^*)}(y)) = D \ell(\pi_{i^*}(x), \pi_{\psi^*(i^*)}(y)). \end{aligned}$$

The above is true for any  $i^* \in \mathbb{N}_D$ . Therefore, for  $\psi = \psi^*$ , the equality holds in 1, i.e.,

$$\min_{\psi \in \Psi} \sum_{i \in \mathbb{N}_D} \ell(\pi_i(x), \pi_{\psi(i)}(y)) = D \ell(\pi_{i^*}(x), \pi_{\psi^*(i^*)}(y)) \quad \forall i^* \in \mathbb{N}_D.$$

The final part of the proof holds by selecting  $i^*$  such that  $\psi^*(i^*) = 0$ ,

$$\ell(\pi_{i^*}(x), \pi_0(y)) = \min_{i \in \mathbb{N}_D} \ell(\pi_i(x), y).$$

□

## B SYNTHETIC DATA GENERATION

The synthetic data are simulated as normalized binary tensors. Lets denote the synthetic data corresponding to the tunable parameters  $\beta_s, \beta_b, \beta_c, \beta_n$  as  $X(\beta_s, \beta_b, \beta_c, \beta_n)$  with dimensions  $N \times E \times D$ , where  $N = 500$  is the number of recordings,  $E = 8$  is the number of electrodes, and  $D = 300$  is the recording time duration. The parameter  $\beta_s = \{0.02, 0.1\}$  represents the probability of a sporadic firing event. The parameter  $\beta_b = \{0.005, 0.025\}$  represents the probability of a sporadic

sequence of multiple firing event, or a burst on a single electrode. The parameter  $\beta_c = \{0, 1\}$  represents the absence or presence of cyclic burst firing pattern, and the parameter  $\beta_n = \{0, 1\}$  represents the absence or presence of network burst firing pattern. Let the indices  $n$ ,  $e$ , and  $d$  represent single instances of recordings, electrodes, and timepoints. Let us also denote the four different firing patterns sporadic single firing, sporadic single-channel burst, cyclic single-channel burst, and network burst as *source events*.

To simulate the synthetic recordings, first, four source-specific binary recordings were simulated corresponding to each of the four different source events, and then those recordings were combined using the binary OR ( $\vee$ ) operation. This means, at any given time-point on a given electrode, a neuronal firing can be observed due to any combination of the source events. The synthetic data simulation algorithm is as follows:

**Algorithm B.1** (Algorithm to simulate the synthetic recordings). *First, initialize the following parameters: Firing frequency within a burst  $\gamma_b = 0.9$ , probability of a network burst starting at a given time point  $\gamma_n = 0.035$ , and network decay factor  $\delta = 0.8$ .*

1. **Sporadic single firings (Spike):** Simulate  $Z_{n,e,d}^{(S)} \sim \text{Bernoulli}(\beta_s)$  i.i.d.
2. **Sporadic single-channel bursts (Burst):**
  - (a) Initialize  $Z^{(B)} = 0$ .
  - (b) Simulate burst initiation indicators  $S_{n,e,d} \sim \text{Bernoulli}(\beta_b)$  i.i.d.
  - (c) For each  $S_{n,e,d} = 1$ ,
    - i. Select a burst duration  $\Delta = \min(D, \Delta^*)$  where  $\Delta^* \sim \text{DiscUnif}(\{3, 4, 5\})$ , the discrete Uniform distribution.
    - ii. Simulate the source-specific firings  $Z_{n,e,i}^{(B)} \sim \text{Bernoulli}(\gamma_b)$  for  $i = d, \dots, d + \Delta$ .
3. **Cyclic single-channel bursts (Cycle):**
  - (a) Initialize  $Z^{(C)} = 0$ . If  $\beta_c = 0$ , then skip to step 4. Else, move to the next step.
  - (b) For each recording  $n$  and electrode  $e$ ,
    - i. Select a cycle period  $Q_c \sim \text{DiscUnif}(\{15, 16, \dots, 19\})$  and phase  $P_c \sim \text{DiscUnif}(\{0, 1, \dots, 14\})$ .
    - ii. Set the burst initiation indicators  $S_{n,e,d} = 1$  if  $(d - P_c)$  is divisible by  $Q_c$ , 0 otherwise.
    - iii. For each  $S_{n,e,d} = 1$ ,
      - A. Select a burst duration  $\Delta = \min(D, \Delta^*)$  where  $\Delta^* \sim \text{DiscUnif}(\{3, 4, 5\})$ , the discrete Uniform distribution.
      - B. Simulate the source-specific firings  $Z_{n,e,i}^{(C)} \sim \text{Bernoulli}(\gamma_b)$  for  $i = d, \dots, d + \Delta$ .
4. **Network bursts (Network):**
  - (a) If  $\beta_n = 0$ , set  $Z^{(N)} = 0$  and skip to step 5. Else, move to the next step.
  - (b) Simulate burst initiation indicators  $S_{n,e,d} \sim \text{Bernoulli}(\gamma_n/E)$  i.i.d.
  - (c) For each  $S_{n,e,d} = 1$ ,
    - i. Denote  $e$  to be the starting electrode, and  $d$  to be starting time-point for the network burst.
    - ii. Select a burst duration  $\Delta = \min(D, \Delta^*)$  where  $\Delta^* \sim \text{DiscUnif}(\{3, 4, 5\})$ , the discrete Uniform distribution.
    - iii. Simulate the source-specific firings  $Z_{n,e',i}^{(N)} \sim \text{Bernoulli}(\gamma_b \delta^{\alpha(e,e')})$  for  $i = d, \dots, d + \Delta$  and  $e' \in \{1, \dots, E\}$ . Here,  $\alpha(e, e')$  represents the physical distance between the electrodes  $e$  and  $e'$  assuming the distance between electrodes 0 and 1 in the configuration described in Fig. 2 to be one unit.
5. **Combine the recordings:** Obtain the combined recording  $Z = Z^{(S)} \vee Z^{(B)} \vee Z^{(C)} \vee Z^{(N)}$ .
6. **Normalize:** The final dataset  $X(\beta_s, \beta_b, \beta_c, \beta_n)$  is obtained by normalizing the dataset  $Z$ .

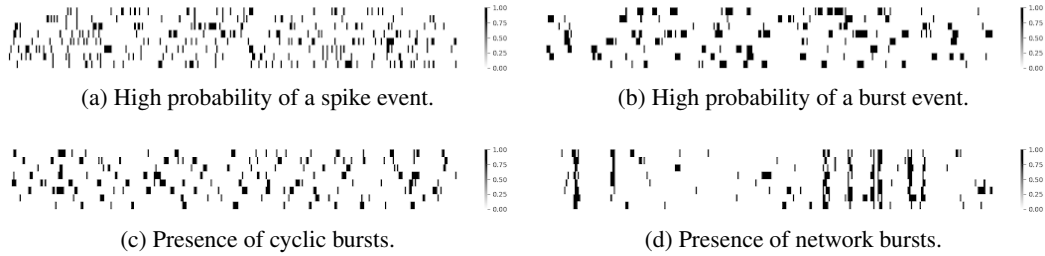


Figure 7: Examples of synthetically generated spike-trains from eight electrodes tuning the probabilities of spike, burst, cycle, or network firing events.

## C EVALUATING STOCHASTIC SHIFT-INVARIANCE

Here, we evaluate the accuracy, computation time, and memory requirements of the Roll-AE model under the stochastic shift-invariance strategy based on the synthetic data (generative model B). We trained the Roll-AE model (as well as standard autoencoders with and without augmentation) with different training batch-sizes, embedding dimensions ( $k$ ), and shift-sampling rates ( $\tau$ ). We evaluated the accuracy under each selection of hyperparameters using the same evaluation scheme outlined in D. All models were trained with learning rate 0.0001.

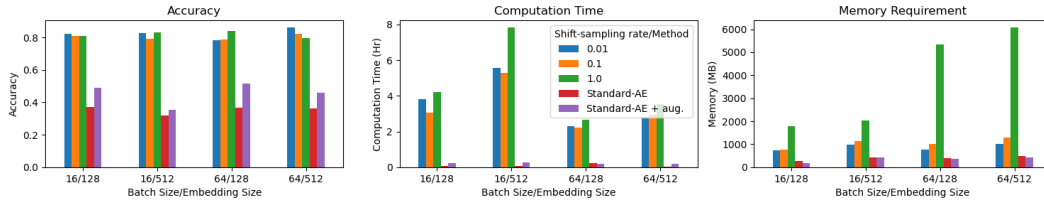


Figure 8: Accuracy, Computation time (Hour), and Memory requirement (MB) for different training batch-sizes and embedding dimensions ( $k$ ), with respect to different shift-sampling rates ( $\tau$ ) (for Roll-AE), and methods (Standard AE and Standard AE with augmentation). All models were trained on an NVIDIA® V100 GPU.

Fig. 8 shows the classification accuracy, computation time, and memory requirements for each selection of the hyperparameters and methods. The accuracies for Roll-AE were similar across all choices of hyperparameters, and except for the case with batch-size = 64 and embedding dimension  $k = 128$ , the shift-sampling rate  $\tau = 0.01$  resulted in the best accuracy for the 16-class classification task on the second-level validation dataset (see D). As expected, the computation time was the longest for  $\tau = 1$  which implies the entire set  $\Pi(x)$  was used in each pass of model training. However, the strongest contributor to the computation time was batch size, with batch-size 16 requiring  $\sim 1.5 - 2$  times more computation time than batch-size 64. The computation time for standard autoencoder was substantially lower, however, at the cost of a substantial drop in accuracy.

The most important benefit of the stochastic shift-invariance is in the memory requirement. Training with  $\tau = 0.01$  required half as much memory for batch-size = 16, and nearly one-seventh as much memory for batch-size 64 compared to  $\tau = 1$ . By requiring substantially lower memory than training with complete shift-invariance (entire  $\Pi(x)$ ), in large datasets, the stochastic shift-invariance strategy can allow for training with larger batch-sizes on limited GPU memory, which in turn can help reduce the computation time without impacting the accuracy.

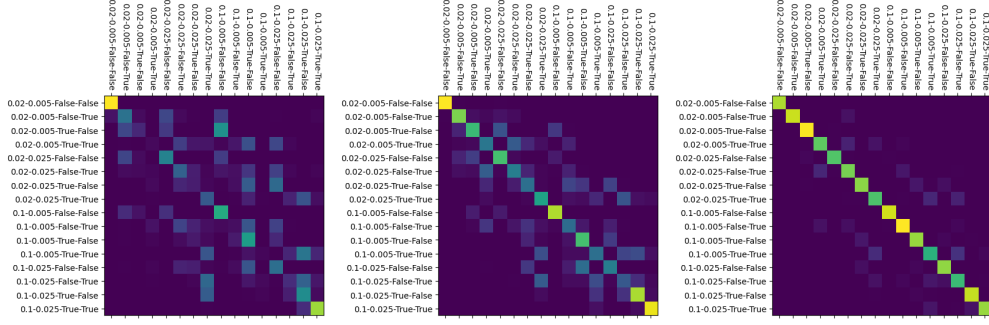


Figure 9: Roll-AE consistently outperforms standard autoencoders (both with and without augmentation) on all binary and multiclass classification tasks. Confusion matrices of standard (left), standard with augmentation (center), and Roll-AE (right) autoencoders for multi-class classification. The 16 class labels were each formatted as {spike probability}-{burst probability}-{cycle present}-{network present}.

## D MODEL TRAINING AND HYPERPARAMETER SELECTION

### D.1 MODEL TRAINING

For the synthetic data, we trained the three models with different hyperparameters (listed in Table 1) on 70% of the data (training) randomly selected, and then applied the trained model on the remaining 30% of the data (validation) to obtain the embeddings. The embeddings of the validation dataset were then further split into a second-level of 70%-30% training/validation data to evaluate the predictive accuracy of those embeddings for the 16-class classification task. For this purpose, a 16-class logistic regression classifier with L2-regularization was trained on the second-level training data, and then the classes were predicted on the second-level validation data. The penalty parameter for the L2-regularization was selected based on four-fold cross-validation. Using the two levels of training/validation data splits, we ensured that both the autoencoder model, and the downstream classifier are generalizable to previously unseen data. For each of the three models, whichever hyperparameters led to the highest predictive accuracy in the second-level validation data, were selected and the models with those selected hyperparameters were then trained on the entire dataset to generate the final embeddings.

Similar two-level validation approach was taken for selecting hyperparameters in the siRNA data. We first trained the three models with the same set of hyperparameters (listed in Table 1) and generated the embeddings based on a 70%-30% training/validation data split. Then, with the embeddings of the validation dataset, we evaluated a logistic regression classifier (with L2 regularization) of the siKD vs NTS samples based on a second-level 70% – 30% split of the data. The hyperparameters which led to the highest accuracy in the second-level validation data were selected for each model. Finally, we applied the three models with the selected hyperparameters on the entire dataset to generate the final embeddings.

### D.2 HYPERPARAMETERS

Table 1 lists all choices of the training parameters that were considered to train the three autoencoder models. For the standard autoencoder with augmentation, two augmentation sampling schemes were evaluated, namely Uniform and Half-mass. For the Uniform sampling scheme, on each epoch, the augmented recording was randomly selected from the set of all possible shifted (and mirror-flipped) recordings uniformly. On the contrary, for the Half-mass sampling scheme, with probability 0.5 the original recording was used as the augmented spike-train, and the with the rest 0.5 probability, the other shifted (and mirror-flipped) recordings were uniformly selected. The best training parameter choice for each autoencoder model was determined based on the performance of the embeddings in the multi-class classification task for synthetic data, or in the binary classification of siKD and NTS in the siRNA experiment data. These parameter choices are highlighted in Table 1 in bold.

Table 1: Choice of training parameters for the three autoencoder models. The best choice of parameters for each autoencoder model are highlighted in bold font (for the synthetic data) or with an asterisk (for the siRNA experiment data).

Training parameter	Standard AE	Standard AE + aug.	Roll-AE
Training batch-size	<b>8</b> , 16, 32, 64*	8, <b>16*</b> , 32, 64	8, 16, 32, <b>64*</b>
Embedding dimension ( $k$ )	128*, <b>256</b> , 512	128, <b>256*</b> , 512	128, 256, <b>512*</b>
Learning rate	0.0001, <b>0.001*</b> , 0.01, 0.1	<b>0.0001*</b> , 0.001, 0.01, 0.1	<b>0.0001*</b> , 0.001, 0.01, 0.1
Augmentation scheme	N/A	<b>Uniform</b> , Half-mass*	N/A
Shift-sampling rate ( $\tau$ )	N/A	N/A	<b>0.01*</b> , 0.05, 0.1, 1

All models had two hidden layers in each of their encoder and decoder MLPs. The hidden layers in the encoder MLPs had  $4k$  and  $2k$  neurons sequentially, where  $k$  is the embedding dimension. Conversely, the hidden layers in the decoder MLPs had  $2k$  and  $4k$  neurons sequentially. Counting the  $k$  parameters for the output layer of the encoder MLP and the  $ED$  parameters for the output layer of the decoder MLP, our models had a total of  $(13k + ED)$  trainable parameters. Mean-squared error loss was used for the standard autoencoders (with or without augmentation), and Linear assignment loss was used for Roll-AE. Adam optimizer Kingma & Ba (2017) was used for the back-propagation in all models. Each model was trained for 200 epochs.