Ridge regression baseline model outperforms deep learning method for cancer genetic dependency prediction

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9 Abstract

10 Accurately predicting genetic or other cellular vulnerabilities of unscreened, or difficult to 11 screen, cancer samples will allow vast advancements in precision oncology. We re-analyzed a 12 recently published deep learning method for predicting cancer genetic dependencies from their 13 omics profiles. After implementing a ridge regression baseline model with an alternative, 14 simplified problem setup, we achieved a model that outperforms the original deep learning 15 method. Our study demonstrates the importance of problem formulation in machine learning 16 applications and underscores the need for rigorous comparisons with baseline approaches.

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19 Precision oncology methods rely on the ability to accurately translate molecular measurements of 20 tumors to insights about their genetic dependencies or other cellular vulnerabilities, ultimately 21 dictating targeted therapeutics. Through genome-wide CRISPR-Cas9 knockout screens, the 22 Cancer Dependency Map(*1*–*5*) (DepMap) has characterized the genetic dependency profiles of 23 over 1000 cancer cell lines (CCLs). Using the DepMap 2018Q2 release, Chiu *et al.* recently 24 developed DeepDEP, a deep learning method for predicting genetic dependency profiles of CCLs 25 given their multi-omics(*6*). DeepDEP was reported to vastly outperform baseline conventional 26 machine learning models. However, we argue here that this result can be attributed to aspects of 27 its problem formulation.

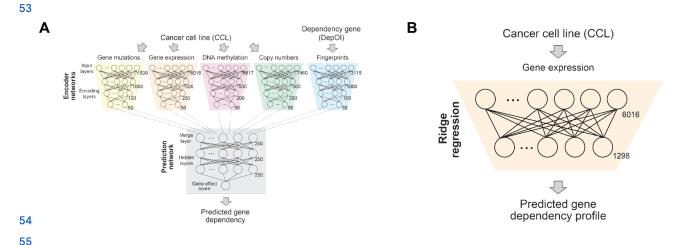
29 Notably, DeepDEP does not jointly predict the entire genetic dependency profile of an input 30 CCL at once. Instead, the model takes as input both multi-omics of a CCL and a functional 31 fingerprint of a single gene dependency of interest (DepOI, as abbreviated by Chiu *et al.*), and 32 outputs the predicted score of that specific gene DepOI for that CCL (Fig. 1A). Functional 33 fingerprints were defined as binary vectors encoding a gene's involvement in 3115 chemical and 34 genetic perturbations (CGPs) from MSigDB v6.2(7), potentially facilitating the model to learn 35 relationships between genes with functional similarities.

37 We recognized that this problem formulation, while likely beneficial for embedding prior 38 knowledge about a gene DepOI's function in a deep learning context, could be problematic for 39 simpler baseline models as it requires a model to learn highly non-linear relationships between 40 input omics features and dependency scores. Because this formulation requires a singular model to predict the score of any gene DepOI (given its functional fingerprint representation), the model is unable to directly relate input omics features to dependency scores. Instead, an optimal model must first generate intermediate representations composed of both a CCL's input omics features and the DepOI's fingerprint vector. Because Chiu *et al.* evaluated baseline model performances using this setup, we were skeptical about the degree by which DeepDEP truly outperforms baseline methods.

48 To evaluate this, we implemented a ridge regression baseline model using a simplified problem 49 formulation (Fig. 1B). Instead of outputting a single gene dependency score for a CCL and gene 50 DepOI combination, our baseline jointly outputs predictions of an input CCL's genetic 51 dependency profile across 1298 cancer relevant genes (defined by Chiu *et al.*), the same gene 52 DepOI set on which DeepDEP was trained and evaluated.

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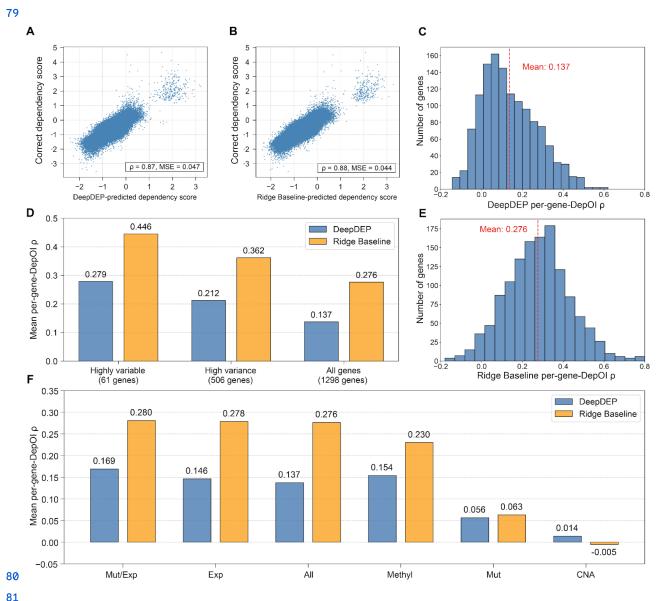
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56 **Fig. 1. Comparison of DeepDEP and baseline setups. (A)** DeepDEP architecture (taken directly from Chiu *et al.*). 57 DeepDEP takes as input a CCL's DNA mutation, gene expression, DNA methylation, and copy number alteration 58 profiles. In addition, a functional fingerprint of a gene dependency of interest (DepOI) is supplied. Dimensionality 59 of each input is displayed. DeepDEP performs dimensionality reduction using an autoencoder pretrained on 8238 60 TCGA tumors. The model then merges the dimensionality-reduced data into a prediction network to predict the 61 score of the gene DepOI (corresponding to the input functional fingerprint) for a given CCL. **(B)** Baseline model 62 setup. A multioutput ridge regression model takes as input omics data (in this illustration, just gene expression) from 63 a CCL, and predicts its genetic dependency profile across 1298 cancer relevant gene DepOIs, as defined by Chiu *et* 64 *al.*

We compared 10-fold cross validation results of DeepDEP and a ridge regression baseline using the aforementioned simplified setup. The ridge regression baseline here uses all input omics features, but no functional fingerprints. Across 278 CCLs and 1298 gene DepOIs, DeepDEP and the ridge regression baseline achieved similar predictive performances (DeepDEP: Fig. 2A, Pearson correlation coefficient $\rho = 0.87$; ridge regression baseline: Fig. 2B, $\rho = 0.88$). However, because of the existence of pan-essential genes, for which dependency score predictions are mostly constant across CCLs and thus easy to predict, analyzing correlation results across all

73 genes likely yields an overly-optimistic estimate of model performances. Thus, we next revaluated the results by computing correlations per-gene-DepOI. DeepDEP achieved a mean respectively per-gene-DepOI ρ of 0.137 (Fig. 2C), while the ridge regression baseline achieved a ρ of 0.276 (Fig. 2E). The ridge regression baseline (with this simplified setup) not only achieved a result respectively higher than that of any baseline machine learning methods evaluated by Chiu *et al.* (not shown here), but also outperformed the full DeepDEP model.



82 Fig. 2. Ridge regression baseline outperforms DeepDEP. (A and B) Scatterplots of DeepDEP (x-axis, A) and 83 ridge baseline (x-axis, B) predicted dependency scores vs the correct dependency scores (y-axis) across 278 CCLs 84 and 1298 gene DepOIs. 10-fold cross-validation, where CCLs are held out, was used to generate predictions for both 85 models. DeepDEP achieves $\rho = 0.87$ and mean squared error (MSE) = 0.047. The ridge baseline achieves $\rho = 0.88$ 86 and MSE = 0.044. (C and E) Histogram of per-gene-DepOI ρ for DeepDEP and the ridge baseline. DeepDEP 87 achieves a mean ρ of 0.137, and the ridge baseline achieves a mean ρ of 0.276. (D) Mean per-gene-DepOI ρ for (as 88 defined in Chiu *et al.*) highly variable dependency score genes (n = 61), high variance dependency score genes (n =

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89 506), and the entire gene set (n = 1298). The ridge regression baseline model outperforms DeepDEP on all gene sets.
90 (F) Simplified models trained on subsets of the omics data. Mut = mutation, Exp = gene expression, Methyl =
91 methylation, and CNA = copy number alteration. For all simplified models except that trained on just CNA data, the
92 ridge regression baseline achieves a higher mean per-gene-DepOI ρ than DeepDEP does.
93
94 Chiu et al. also analyzed mean per-gene-DepOI p on two subsets of genes that were observed to
95 have high variance dependency scores and thus were likely cancer-relevant genes. The ridge
96 regression baseline model outperformed DeepDEP on both of these gene sets (Fig. 2D).
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98 We next constructed several simplified ridge models using subsets of the omics data types (for
99 example, Fig. 1B depicts an expression-only ridge model), similarly as done in Chiu et al. These
100 simplified models were compared with DeepDEP results. In all instances except when using only
101 copy number alteration data, the ridge regression baseline achieves a higher mean
102 per-gene-DepOI ρ than DeepDEP does (Fig. 2F).
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104 These results demonstrate how machine learning problem formulations dramatically impact the
105 performance of baseline approaches. We show that a problem formulation that is convenient for
106 embedding information into deep learning models may not always be the ideal formulation for
107 baseline approaches. To truly evaluate the degree by which novel methods outperform baselines.
108 it is necessary to rigorously evaluate baselines using different, potentially simpler, problem
109 formulations. Moreover, many of our simplified ridge regression baselines, trained on subsets of
110 the data types (Mut/Exp, Exp, Methyl), outperformed DeepDEP trained on all input genomic
111 features. This demonstrates that the proposed deep learning model was not able to benefit from
112 integrating information from a diverse set of data modalities. We also demonstrate how the use
113 of gene functional fingerprints is not important for achieving an elevated prediction performance
114 over DeepDEP. These results bring into question the strength of the results obtained from
115 downstream analyses using the DeepDEP model, such as the pan-cancer tumor dependency map
116 that Chiu et al. generated by applying DeepDEP on TCGA data.
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118 Overall, we demonstrate the importance of conducting rigorous baselines when evaluating the
119 performance of novel methods, and the importance of considering the implications of different
120 machine learning problem formulations.
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122 Data and code availability
123 All data was obtained from the Code Ocean compute capsule accompanying Chiu et al., the
124 supplementary tables of Chiu et al., and from the DepMap portal. Machine learning tasks were
125 performed using Scikit-learn v1.2.2(8). All code and data for this analysis can be found at:
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126 https://github.com/danielchang2002/deepdep reanalysis.

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128 Methods

- 129 DeepDEP CCL dependency score predictions from 10-fold cross-validation (where CCLs are
- 130 held out) were obtained from Supplementary Table S8 of Chiu et al. Ground truth gene effect
- 131 scores of the 278 CCLs were obtained from the DepMap 2018Q2 release. The DeepDEP 10-fold
- 132 cross-validation dependency score predictions were compared with the ground truth in Fig. 2A
- 133 and 1C.

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- 135 DNA mutation, gene expression, DNA methylation, and copy number alteration data of the 278
- 136 CCLs were obtained from the Code Ocean compute capsule accompanying Chiu et al.
- 137 Functional fingerprints were not used in our analysis. Ridge regression
- 138 (sklearn.linear model.Ridge) with default parameters was then used to predict an input CCL's
- 139 genetic dependency profile across 1298 cancer relevant genes (defined by Chiu et al.), the same
- 140 gene DepOI set on which DeepDEP was trained and evaluated, given a flattened vector of the
- 141 four data types concatenated together for an input CCL. Ridge regression baseline prediction
- 142 scores were obtained via 10-fold cross-validation (where CCLs are held out). Notably, the CCL
- 143 fold partitioning is not identical to that used to generate DeepDEP cross-validation results in
- 144 Supplementary Table S8 of Chiu et al., but generated independently in this analysis
- 145 (sklearn.model selection.KFold; random state = 42). Ridge regression baseline 10-fold
- 146 cross-validation scores were compared with the ground truth in Fig. 2B and 2E.
- 148 The gene set of 61 "highly variable" genes was obtained by locating genes with dependency
- 149 score standard deviations greater than 0.3, as defined by Chiu et al. The gene set of 506 "high
- 150 variance" genes was obtained using the "Achilles_high_variance_genes.csv" file provided by the
- 151 DepMap portal. The 10-fold cross-validation results on these two gene subsets, for both the
- 152 Ridge regression baseline and DeepDEP, are detailed in Fig. 2D.
- 154 Results for simplified ridge regression baseline models were obtained using subsets of the CCL
- 155 omics data. This was performed identically as before, using 10-fold cross-validation. However,
- 156 for DeepDEP, the 10-fold cross-validation prediction scores were only available (in the
- 157 supplementary of Chiu et al.) for the model using all omics features (i.e. the "All" model), and
- 158 not for the simplified models. Thus, in Fig. 2F, simplified DeepDEP model performances were
- 159 obtained using data from Supplementary Fig. S5 of Chiu et al., which details per-gene-DepOI ρ
- 160 for simplified models using 10 independent train-test subsampling (i.e., a slightly different
- 161 evaluation method than 10-fold cross-validation).

163 References

- 164 1. R. M. Meyers, J. G. Bryan, J. M. McFarland, B. A. Weir, A. E. Sizemore, H. Xu, N. V. Dharia, P. G.
- Montgomery, G. S. Cowley, S. Pantel, A. Goodale, Y. Lee, L. D. Ali, G. Jiang, R. Lubonja, W. F.
- Harrington, M. Strickland, T. Wu, D. C. Hawes, V. A. Zhivich, M. R. Wyatt, Z. Kalani, J. J. Chang,
- M. Okamoto, K. Stegmaier, T. R. Golub, J. S. Boehm, F. Vazquez, D. E. Root, W. C. Hahn, A.

- Tsherniak, Computational correction of copy number effect improves specificity of CRISPR-Cas9
- essentiality screens in cancer cells. *Nat. Genet.* **49**, 1779–1784 (2017).
- 170 2. M. Ghandi, F. W. Huang, J. Jané-Valbuena, G. V. Kryukov, C. C. Lo, E. R. McDonald, J. Barretina,
- E. T. Gelfand, C. M. Bielski, H. Li, K. Hu, A. Y. Andreev-Drakhlin, J. Kim, J. M. Hess, B. J. Haas,
- F. Aguet, B. A. Weir, M. V. Rothberg, B. R. Paolella, M. S. Lawrence, R. Akbani, Y. Lu, H. L. Tiv, P.
- 173 C. Gokhale, A. de Weck, A. A. Mansour, C. Oh, J. Shih, K. Hadi, Y. Rosen, J. Bistline, K.
- Venkatesan, A. Reddy, D. Sonkin, M. Liu, J. Lehar, J. M. Korn, D. A. Porter, M. D. Jones, J. Golji,
- G. Caponigro, J. E. Taylor, C. M. Dunning, A. L. Creech, A. C. Warren, J. M. McFarland, M.
- Zamanighomi, A. Kauffmann, N. Stransky, M. Imielinski, Y. E. Maruvka, A. D. Cherniack, A.
- Tsherniak, F. Vazquez, J. D. Jaffe, A. A. Lane, D. M. Weinstock, C. M. Johannessen, M. P.
- Morrissey, F. Stegmeier, R. Schlegel, W. C. Hahn, G. Getz, G. B. Mills, J. S. Boehm, T. R. Golub, L.
- A. Garraway, W. R. Sellers, Next-generation characterization of the Cancer Cell Line Encyclopedia.
- *Nature.* **569**, 503–508 (2019).
- 181 3. C. Pacini, J. M. Dempster, I. Boyle, E. Goncalves, H. Najgebauer, E. Karakoc, D. van der Meer, A.
- Barthorpe, H. Lightfoot, P. Jaaks, J. M. McFarland, M. J. Garnett, A. Tsherniak, F. Iorio, Integrated
- cross-study datasets of genetic dependencies in cancer. *Nat. Commun.* 12, 1–14 (2021).
- 184 4. J. M. Dempster, J. Rossen, M. Kazachkova, J. Pan, G. Kugener, D. E. Root, A. Tsherniak, Extracting
- Biological Insights from the Project Achilles Genome-Scale CRISPR Screens in Cancer Cell Lines.
- *bioRxiv* (2019), p. 720243.
- 187 5. J. M. Dempster, I. Boyle, F. Vazquez, D. Root, J. S. Boehm, W. C. Hahn, A. Tsherniak, J. M.
- McFarland, Chronos: a CRISPR cell population dynamics model. bioRxiv (2021), p.
- **189** 2021.02.25.432728.
- 190 6. Y.-C. Chiu, S. Zheng, L.-J. Wang, B. S. Iskra, M. K. Rao, P. J. Houghton, Y. Huang, Y. Chen,
- 191 Predicting and characterizing a cancer dependency map of tumors with deep learning. *Science*
- *Advances* (2021), doi:10.1126/sciadv.abh1275.
- 193 7. A. Liberzon, A. Subramanian, R. Pinchback, H. Thorvaldsdóttir, P. Tamayo, J. P. Mesirov, Molecular
- signatures database (MSigDB) 3.0. *Bioinformatics*. **27**, 1739–1740 (2011).
- 195 8. Pedregosa, Varoquaux, Gramfort, Scikit-learn: Machine learning in Python. the Journal of machine
- 196 (available at
- https://www.jmlr.org/papers/volume12/pedregosa11a/pedregosa11a.pdf?ref=https://githubhelp.com).