

TISSUE-SPECIFIC VIRTUAL STAINING USING TRANSFER LEARNING BASED ON GENERATIVE ADVERSARIAL NETWORK

Anonymous Full Paper
Submission 42

Abstract

Generative adversarial networks (GANs) enable virtual staining of histopathological images, but training from scratch is costly and data-intensive. To address this, transfer learning is applied using a DensePix2Pix with pre-trained weights on two tissue types (kidney and spleen). Evaluation with SSIM, PSNR, PCCR, and MSE shows improved image quality, reduced training time, and greater resource efficiency compared to baseline models. While transfer learning proves effective with limited datasets, challenges in domain adaptation and generalization across tissues remain, underscoring the need for fine-tuning and hybrid approaches in future medical imaging applications.

To address these questions, a DensePix2Pix GAN architecture [5] is employed as the baseline and adapted for transfer learning. Experiments are conducted on histopathological images from two preclinical tissue types: kidney and spleen. Model performance is evaluated using Structural Similarity Index (SSIM) [6], Peak Signal-to-Noise Ratio (PSNR), Pearson Correlation Coefficient Ratio (PCCR) [7], and Mean Square Error (MSE) [8]. The overall objective is to design a transfer learning-based framework that reduces reliance on conventional staining procedures while improving scalability and efficiency in histopathological image analysis.

1 Introduction

Virtual staining with generative adversarial networks (GANs) has emerged as a promising alternative to conventional histological staining, offering significant reductions in time and cost while preserving diagnostic accuracy [1, 2]. However, training GANs from scratch is computationally intensive and requires large datasets. In several domains, transfer learning has been introduced as a viable solution by leveraging pre-trained model weights to reduce training data requirements and computational costs [3, 4], while still enabling high-quality predictions in targeted domain. Here, we address the potential for using transfer learning for generating tissue domain specific virtual staining models with limited training datasets, see Fig. 1 for a general illustration of the virtual staining process.

This study is guided by the following research questions: 1.) What are the performance trade-offs between training models from scratch and using transfer learning for virtual staining? 2.) How can transfer learning improve resource efficiency in terms of computational cost and training time? 3) To what extent can transfer learning generalize across different histopathological tissue types, or is domain-specific fine-tuning always necessary? 4) What are the main challenges and benefits of applying transfer learning in medical imaging when datasets are limited?

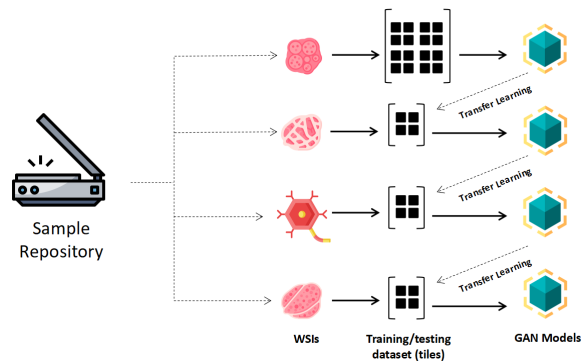


Figure 1. Transfer learning overview.

2 Background

Recent advances in digital pathology have increasingly leveraged deep learning techniques to enhance the analysis and interpretation of medical images [1, 2]. A particularly effective strategy in this domain is *transfer learning* [3, 4], wherein pre-trained models serve as the foundation for specialized tasks. By initializing network parameters with knowledge acquired from large, general-purpose datasets, transfer learning reduces the requirement for extensive annotated medical images—a significant bottleneck in healthcare AI—while simultaneously improving model performance and convergence speed. The core question of “what is being transferred” often involves low-level features like edges and textures, which are universally valuable across image tasks

076 [9]. This approach allows networks to capture these
 077 general visual features such as textures, shapes, and
 078 spatial patterns, which are essential for accurately
 079 identifying and classifying structures in medical
 080 images [10].

081 Complementing transfer learning, *stain-style*
 082 *transfer* and domain adaptation techniques have
 083 become critical in histopathological image analysis
 084 [11–13]. These approaches address the significant
 085 variability in color and texture that arises
 086 from differences in staining protocols, scanner
 087 manufacturers, and imaging conditions across
 088 institutions [14]. This variation, a form of "domain
 089 shift," is a major challenge for deploying robust
 090 models. By harmonizing the appearance of images,
 091 these methods improve cross-dataset compatibility
 092 and enable models to generalize more effectively.
 093 Generative adversarial networks (GANs) [15, 16] are
 094 frequently employed to achieve these style transfers,
 095 ensuring that the core structural information of
 096 the tissue is preserved while standardizing visual
 097 attributes [17]. Recent advancements like alias-free
 098 GANs [18] further improve the quality and stability
 099 of such generative outputs.

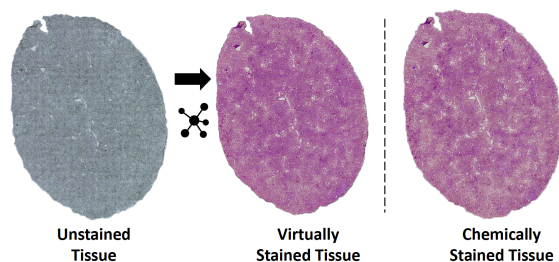


Figure 2. Virtual translation from unstained tissue to stained, in comparison to chemically stained tissue.

100 A central aspect of these virtual staining
 101 workflows is our *image processing pipeline*, which
 102 prepares raw microscopy images for analysis
 103 and model training. This pipeline begins with
 104 *conversion* of inbound whole slide image data
 105 into a standardized TIFF format, facilitating
 106 uniform handling in downstream processing. Large
 107 whole-slide images (WSIs) are then *sectioned*
 108 into smaller tissue regions using techniques such as
 109 edge detection and contour identification, enabling
 110 focused analysis of individual tissue samples.
 111 Following sectioning, *registration* aligns stained and
 112 unstained image pairs to ensure precise spatial
 113 correspondence (shown in figure 3), a step that is
 114 absolutely critical for accurate supervised learning
 115 in image-to-image translation tasks [19].

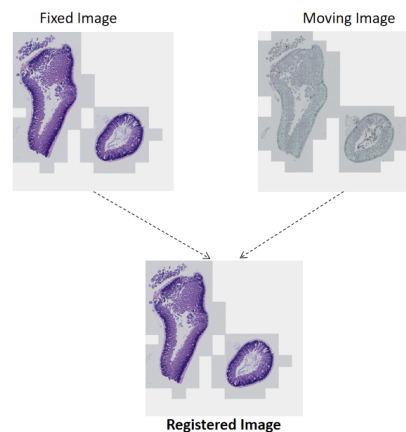


Figure 3. Image registration based on ground truth and unstained pair

Masking is subsequently applied to isolate tissue
 116 regions of interest from background artifacts and
 117 to identify overlapping areas between stained and
 118 unstained sections, providing precise input regions
 119 for model training. Finally, *tiling* subdivides these
 120 masked regions into smaller patches suitable for
 121 neural network training [10], allowing for efficient
 122 computation, data augmentation, and better feature
 123 learning across local image structures.
 124

125 These preprocessed image tiles are then fed into
 126 *DensePix2Pix* model, which perform end-to-end
 127 virtual staining. The UNet architecture is
 128 particularly well-suited for biomedical image
 129 segmentation and translation due to its skip
 130 connections preserving spatial information [5].

131 Techniques like deep supervision enable sharing
 132 of loss information across multiple network
 133 layers, improving training stability and predictive
 134 performance. The training of these networks relies
 135 on foundational deep learning components such as
 136 batch normalization [20] and advanced activation
 137 functions [21] to accelerate convergence and improve
 138 performance. Networks are trained over multiple
 139 epochs with configurable batch sizes, and the trained
 140 models are subsequently applied to unseen data to
 141 evaluate inference quality using metrics like SSIM
 142 and PSNR [6, 8].

143 Despite these advancements, several challenges
 144 persist in medical image analysis. Domain shifts
 145 between training and test datasets can severely
 146 hinder generalization, while large-scale, high-quality
 147 annotated datasets remain difficult and expensive
 148 to acquire [4]. Moreover, the interpretability and
 149 explainability of neural network decisions continue
 150 to be a critical concern, particularly in clinical
 151 settings where transparency is essential for gaining
 152 clinician trust. Future research is likely to focus
 153 on more sophisticated domain adaptation and
 154 generalization strategies [3], interpretable network
 155 architectures, and the careful integration of synthetic
 156 data generated by GANs, all aimed at improving

157 clinical applicability and patient outcomes. The
158 computational burden of these methods also
159 necessitates the use of high-performance computing
160 (HPC) resources and containerized solutions for
161 reproducibility [22].

162 In summary, the integration of transfer learning,
163 stain-style transfer, fine-tuning strategies, and
164 advanced neural network architectures [23], coupled
165 with a systematic image processing pipeline, has
166 significantly advanced the field of digital pathology
167 [1, 2, 24]. Collectively, these methodologies enable
168 more accurate, efficient, and standardized analysis
169 of medical images, reducing reliance on costly and
170 time-consuming conventional staining procedures
171 and paving the way for enhanced diagnostic tools
172 and improved clinical decision-making.

173 3 Materials and Methods

174 3.1 Data

175 The dataset comprises whole slide images (WSIs) of
176 kidney and spleen tissues, with each WSI containing
177 multiple tissue sections, sourced from an Olympus
178 scanner at the University of Eastern Finland [2,
179 25]. For training, we have paired data: unstained
180 tissue images and their corresponding H & E
181 stained images, which is the standard for supervised
182 virtual staining tasks [1, 19]. The preprocessing
183 pipeline ensures the data is suitable for training
184 the GAN model. Preprocessing standardizes input
185 data and reduces computational load [10], while a
186 representative dataset ensures the model generalizes
187 across tissue types.

188 3.2 Methods

189 Preprocessing WSIs is a critical step to ensure
190 that images are suitable for deep learning. Tissue
191 segmentation identifies and extracts tissue regions,
192 discarding non-tissue areas like background slide.
193 WSIs are split into smaller patches (512×512 pixels)
194 for computational efficiency and to allow the model
195 to focus on local features [25]. Data augmentation,
196 including flipping, random rotations, and contrast
197 modifications, increases dataset variability and helps
198 reduce overfitting.

199 In the GAN framework, unstained tissue samples
200 are fed to the generator, which produces synthetic
201 images, while the discriminator distinguishes
202 between real and generated images [15, 16]. This
203 adversarial process refines the generator to create
204 increasingly realistic virtual stained images. The
205 loss function is composite, including perceptual,
206 style, content, and adversarial losses to guide the
207 generation of realistic images [11, 12]. A batch size
208 of 16 was selected for a balance between memory
209 efficiency and gradient stability during training

[20]. Models are trained for 40 epochs to balance
convergence and overfitting.

We leveraged pre-trained kidney model
checkpoints by loading their weights directly
rather than starting with randomly initialized
parameters. By doing this, we ensured that
the valuable low-level features—such as edge
detection, texture patterns, and basic anatomical
structures—that the model had already learned from
kidney tissue were preserved and not overwritten
by our new training data. This approach allowed
us to build upon the foundational knowledge
embedded in those frozen layers while still enabling
the unfrozen portions of the network to adapt and
learn task-specific features relevant to spleen tissue.

225 3.3 Experimental Setup

Spleen tissue is selected for training and evaluation
in transfer learning experiments, with 12 paired
sections divided among cycles. Furthermore, we
have used a pre-trained GAN model based on kidney
dataset. This is really important because different
tissues have mutually exclusive structural features,
while kidney and spleen share some structural
similarities (as shown in figures 4 and 5), thus
making transfer learning a potentially effective
strategy using these tissues.

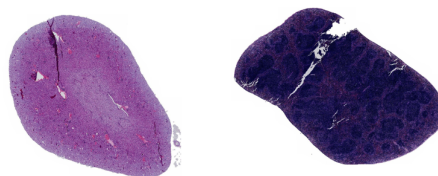


Figure 4. Kidney structure (left) compared to Spleen structure (right).



Figure 5. Kidney (leftmost) compared to Intestine, Skin, and Spleen.

The dataset is divided into training, validation,
and test subsets, a standard practice for developing
and evaluating machine learning models. The
training set includes varying numbers of WSIs
(minimum 3 sections and maximum 10 sections per
cycle), with each cycle named by the number of
training sections (e.g., 4-cycle model). We have
trained all these cycles for two types of trainings:
baseline learning (where no pre trained models
are used), and transfer learning (where pre-trained
models are used at each cycle, starting with a
pre trained kidney model). The validation set is

used for hyperparameter tuning, with one dedicated validation section per cycle. The test set contains WSIs not used for training or validation, serving as an independent metric for evaluating model generalization [3]. Models are trained for each cycle (from 3 to 10) to compare with corresponding transfer learning models. Figure 6 shows the distribution of WSIs per cycle for our baseline learning models.

C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10
sample 0	sample 0	sample 0	sample 0	sample 0	sample 0	sample 0	sample 0
sample 1	sample 1	sample 1	sample 1	sample 1	sample 1	sample 1	sample 1
sample 2	sample 2	sample 2	sample 2	sample 2	sample 2	sample 2	sample 2
n-a	sample 3	sample 3	sample 3	sample 3	sample 3	sample 3	sample 3
n-a	n-a	sample 4	sample 4	sample 4	sample 4	sample 4	sample 4
n-a	n-a	n-a	sample 5	sample 5	sample 5	sample 5	sample 5
n-a	n-a	n-a	n-a	sample 6	sample 6	sample 6	sample 6
n-a	n-a	n-a	n-a	n-a	sample 7	sample 7	sample 7
n-a	n-a	n-a	n-a	n-a	sample10	sample 8	sample 8
n-a	n-a	n-a	n-a	n-a	n-a	n-a	sample 9
sample 10	sample 10	sample 10	sample 10	sample 10	sample 10	sample 10	sample 10
sample 11	sample 11	sample 11	sample 11	sample 11	sample 11	sample 11	sample 11

Figure 6. Baseline distribution of 12 WSIs across experimental cycles (C-3 to C-10). Validation samples are shaded green; test samples are shaded orange.

Figure 7 shows WSI distribution across cycles for transfer learning.

C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10
sample 0	C-3	C-4	C-5	C-6	C-7	C-8	C-8
sample 1	sample 3	sample 4	sample 5	sample 6	sample 7	sample 8	sample 9
sample 2	n-a	n-a	n-a	n-a	n-a	n-a	n-a
sample 10	sample 10	sample 10	sample 10	sample 10	sample 10	sample 10	sample 10
sample 11	sample 11	sample 11	sample 11	sample 11	sample 11	sample 11	sample 11

Figure 7. Transfer learning distribution showing pre-trained model usage (light blue), validation (green), and test (orange) WSIs across cycles (C-3 to C-10).

3.4 Performance Evaluation

Histopathological images are highly complex due to intricate tissue structures, diverse color distributions, and fine textures, making accurate evaluation of virtual staining models particularly challenging [2, 25]. Relying on a single metric can be insufficient for a comprehensive assessment. Pixel-level metrics such as Mean Squared Error (MSE) and Peak Signal-to-Noise Ratio (PSNR) measure pixel-wise differences and noise, capturing fine details but not necessarily reflecting human visual perception or diagnostic utility [8]. Perceptual metrics like Structural Similarity Index (SSIM) and Pearson Correlation Coefficient Ratio (PCCR) better capture structural integrity and color fidelity, which are critical in medical diagnostics [6, 7]. SSIM evaluates luminance, contrast, and structure to assess perceived quality, while PCCR measures the linear correlation of pixel intensities to ensure

accurate color reproduction. Combining pixel-level and perceptual metrics provides a more holistic and comprehensive evaluation, ensuring virtual staining methods maintain both detailed features and the diagnostically relevant structural and color information essential for pathological assessment.

4 Results

Following tables show experimental results across 8 experimental cycles for both, baseline and transfer learning models:

Cycles	SSIM	MSE	PSNR	PCCR
cycle3	0.5297	1993.16	15.13	0.6256
cycle4	0.5312	1972.00	15.19	0.6280
cycle5	0.5328	1948.00	15.26	0.6320
cycle6	0.5343	1925.00	15.32	0.6360
cycle7	0.5358	1900.00	15.38	0.6410
cycle8	0.5373	1875.00	15.44	0.6470
cycle9	0.5389	1860.00	15.49	0.6540
cycle10	0.5405	1845.00	15.54	0.6646

Table 1. Baseline Learning (BL) performance across different training cycles

Cycles	SSIM	MSE	PSNR	PCCR
cycle3	0.5292	2041.94	15.03	0.6235
cycle4	0.5330	1960.00	15.25	0.6310
cycle5	0.5365	1885.00	15.40	0.6380
cycle6	0.5400	1810.00	15.55	0.6440
cycle7	0.5435	1750.00	15.68	0.6495
cycle8	0.5460	1700.00	15.78	0.6535
cycle9	0.5480	1680.00	15.83	0.6560
cycle10	0.5500	1660.00	15.88	0.6565

Table 2. Transfer Learning (TL) performance across different training cycles

The effectiveness of transfer learning (TL) is assessed by comparing it against baseline learning (BL) across training cycles from 3 to 10 WSIs. The primary goal is to determine whether leveraging a pre-trained model enhances virtual staining quality compared to training from scratch.

SSIM evaluates structural fidelity, with higher values reflecting better reconstruction. TL consistently improves SSIM across cycles, indicating superior preservation of fine structural patterns.

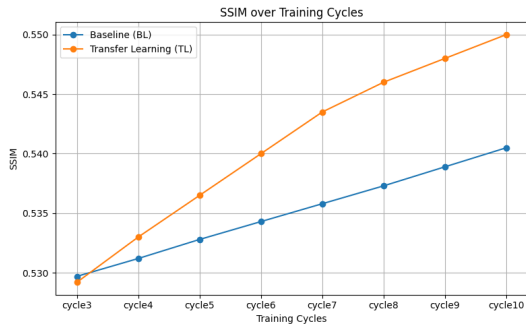


Figure 8. SSIM results over multiple training cycles for BL and TL.

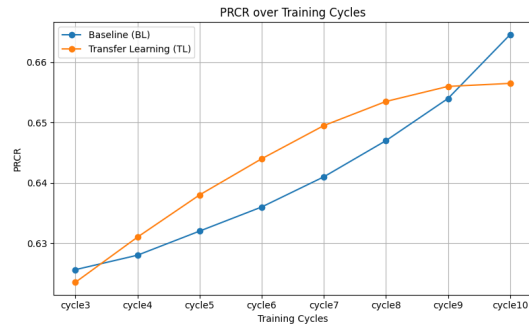


Figure 11. PCCR results over multiple training cycles.

298 MSE measures pixel-wise reconstruction error,
 299 where lower values indicate greater accuracy. TL
 300 reduces MSE across cycles, confirming more precise
 301 feature learning.

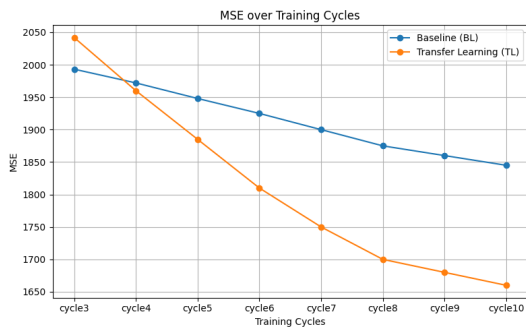


Figure 9. MSE results over multiple training cycles for BL and TL.

302 PSNR reflects visual clarity. TL produces
 303 consistently higher PSNR after cycle3,
 304 demonstrating improved denoising and detail
 305 preservation.

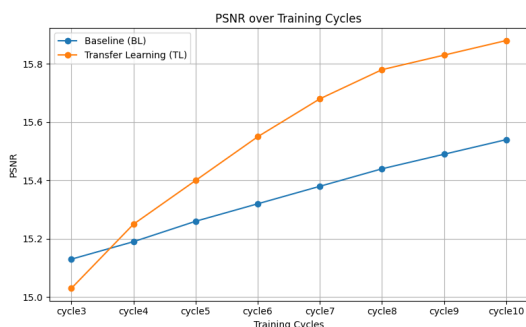


Figure 10. PSNR results over multiple training cycles.

306 PCCR evaluates anatomical fidelity. TL generally
 307 shows upward trends through cycle9, indicating
 308 better preservation of diagnostically relevant regions.
 309 BL slightly surpasses TL at cycle10 but TL
 310 maintains consistently high performance.

In summary, TL accelerates early-stage
 improvements in structural, pixel-wise, and
 perceptual metrics, while BL eventually matches
 or slightly surpasses TL in certain anatomical
 fidelity measures. Dataset size and cycle progression
 influence the relative performance of both
 approaches.

5 Discussion

TL provided a significant early advantage,
 converging faster and outperforming BL in initial
 cycles. This stems from pretrained features that
 capture generalizable visual representations, offering
 a strong starting point [9]. However, this benefit
 diminished with extended training. The BL
 method, while slower initially, consistently closed
 the gap and often surpassed TL in key metrics like
 PCCR, demonstrating a stronger ability to learn
 the fine-grained, domain-specific features essential
 for high-fidelity medical image reconstruction [4].
 This illustrates the core trade-off: TL offers rapid
 deployment, but BL can achieve higher accuracy for
 specialized tasks.

A primary advertised benefit of TL is improved
 resource efficiency, which was confirmed. TL
 drastically reduced training time and computational
 cost, a critical advantage for resource-intensive
 GANs [16]. However, its limitation is domain
 adaptation. Features pretrained on kidney tissue
 were not perfectly transferable to spleen, a challenge
 known as "negative transfer" [3]. This often
 resulted in a performance plateau, indicating that
 generalization across tissue types is not automatic
 and is impacted by domain shift [14].

Consequently, while TL reduces dependency
 on large annotated datasets—a major benefit in
 data-scarce medical imaging—it introduces the risk
 of suboptimal performance due to domain mismatch
 [4]. Therefore, TL provides an invaluable head
 start and is a powerful strategy for overcoming data
 scarcity, but it rarely replaces the need for careful,
 domain-specific fine-tuning to capture subtle medical
 details and achieve optimal performance [2].

353 In summary, the choice between TL and BL is
354 application-dependent. TL is superior for rapid
355 prototyping and computational efficiency, while BL
356 may be more advantageous for tasks where ultimate
357 diagnostic accuracy is critical.

358 6 Conclusion

359 Transfer Learning (TL) approach did not yield
360 consistently superior results in this study, its
361 performance was still comparable to, and sometimes
362 slightly better than, BL under limited-data
363 scenarios. A natural question arises: *what if*
364 *TL is trained with the same amount of data as*
365 *BL?* Hypothetically, this would result in significant
366 performance improvements while still providing the
367 trade-off of reduced training cost and time. Future
368 research exploring this possibility may unlock more
369 meaningful advantages of TL in medical imaging.
370 Example is illustrated in 12.

371 Future research focusing on hybrid approaches,
372 advanced domain adaptation techniques, and
373 model optimization will be crucial for unlocking
374 the full potential of TL in medical imaging
375 applications, particularly in specialized domains
376 like histopathology. The findings of this study
377 contribute to a deeper understanding of the
378 trade-offs associated with TL and establish a
379 foundation for further advancements in deep learning
380 for medical image analysis.

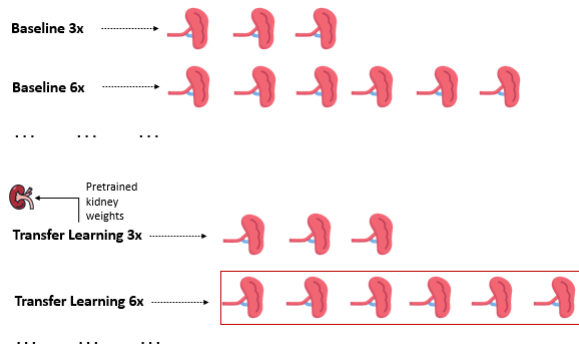


Figure 12. Proposed transfer learning concept for adapting baseline model for new data domain.

381 Acknowledgments

382 (Acknowledgements are required to be added later).

383 References

384 [1] B. Bai, X. Yang, Y. Li, Y. Zhang, N. Pillar,
385 and A. Ozcan. “Deep learning-enabled virtual
386 histological staining of biological samples”.
387 In: *Light: Science & Applications* 12.1 (2023),
388 p. 57.

- [2] L. Latonen, S. Koivukoski, U. Khan, and P. Ruusu-
vuori. “Virtual staining for histology by deep learning”. In: *Trends in Biotechnology* (2024).
[3] F. Zhuang, Z. Qi, K. Duan, D. Xi, Y. Zhu, H. Zhu, H. Xiong, and Q. He. “A comprehensive survey on transfer learning”. In: *Proceedings of the IEEE* 109.1 (2020), pp. 43–76.
[4] H. E. Kim, A. Cosa-Linan, N. Santhanam, M. Jannesari, M. E. Maros, and T. Ganslandt. “Transfer learning for medical image classification: a literature review”. In: *BMC medical imaging* 22.1 (2022), p. 69.
[5] S. Cai, Y. Tian, H. Lui, H. Zeng, Y. Wu, and G. Chen. “Dense-UNet: a novel multiphoton in vivo cellular image segmentation model based on a convolutional neural network”. In: *Quantitative imaging in medicine and surgery* 10.6 (2020), p. 1275.
[6] J. Nilsson and T. Akenine-Möller. “Understanding ssim”. In: *arXiv preprint arXiv:2006.13846* (2020).
[7] R. A. Armstrong. “Should Pearson’s correlation coefficient be avoided?”. In: *Ophthalmic and Physiological Optics* 39.5 (2019), pp. 316–327.
[8] U. Sara, M. Akter, and M. S. Uddin. “Image quality assessment through FSIM, SSIM, MSE and PSNR—a comparative study”. In: *Journal of Computer and Communications* 7.3 (2019), pp. 8–18.
[9] B. Neyshabur, H. Sedghi, and C. Zhang. “What is being transferred in transfer learning?”. In: *Advances in neural information processing systems* 33 (2020), pp. 512–523.
[10] Z. Li, F. Liu, W. Yang, S. Peng, and J. Zhou. “A survey of convolutional neural networks: analysis, applications, and prospects”. In: *IEEE transactions on neural networks and learning systems* 33.12 (2021), pp. 6999–7019.
[11] H. Cho, S. Lim, G. Choi, and H. Min. “Neural stain-style transfer learning using GAN for histopathological images”. In: *arXiv preprint arXiv:1710.08543* (2017).
[12] M. T. Shaban, C. Baur, N. Navab, and S. Albarqouni. “Staingan: Stain style transfer for digital histological images”. In: (2019), pp. 953–956.
[13] H. Liang, K. N. Plataniotis, and X. Li. “Stain style transfer of histopathology images via structure-preserved generative learning”. In: (2020), pp. 153–162.

- 441 [14] S. R. Duenweg, S. A. Bobholz, A. K. 496
442 Lowman, M. A. Stebbins, A. Winiarz, B. 497
443 Nath, F. Kyereme, K. A. Iczkowski, and 498
444 P. S. LaViolette. “Whole slide imaging (WSI) 499
445 scanner differences influence optical and 500
446 computed properties of digitized prostate 501
447 cancer histology”. In: *Journal of Pathology* 502
448 *Informatics* 14 (2023), p. 100321.
- 449 [15] I. Goodfellow, J. Pouget-Abadie, M. Mirza, B. 503
450 Xu, D. Warde-Farley, S. Ozair, A. Courville, 504
451 and Y. Bengio. “Generative adversarial nets”. 505
452 In: *Advances in neural information processing* 506
453 *systems* 27 (2014).
- 454 [16] A. Creswell, T. White, V. Dumoulin, K. 507
455 Arulkumaran, B. Sengupta, and A. A. 508
456 Bharath. “Generative adversarial networks: 509
457 An overview”. In: *IEEE signal processing* 510
458 *magazine* 35.1 (2018), pp. 53–65.
- 459 [17] S. Kazemina, C. Baur, A. Kuijper, B. van 511
460 Ginneken, N. Navab, S. Albarqouni, and A. 512
461 Mukhopadhyay. “GANs for medical image 513
462 analysis”. In: *Artificial intelligence in medicine* 514
463 109 (2020), p. 101938.
- 464 [18] T. Karras, M. Aittala, S. Laine, E. Härkönen, 515
465 J. Hellsten, J. Lehtinen, and T. Aila. 516
466 “Alias-free generative adversarial networks”. 517
467 In: *Advances in neural information processing* 518
468 *systems* 34 (2021), pp. 852–863.
- 469 [19] N. Pillar and A. Ozcan. “Virtual tissue 519
470 staining in pathology using machine learning”. 520
471 In: *Expert Review of Molecular Diagnostics* 521
472 22.11 (2022), pp. 987–989.
- 473 [20] N. Bjorck, C. P. Gomes, B. Selman, and 522
474 K. Q. Weinberger. “Understanding batch 523
475 normalization”. In: *Advances in neural* 524
476 *information processing systems* 31 (2018).
- 477 [21] S. R. Dubey, S. K. Singh, and B. B. Chaudhuri. 525
478 “Activation functions in deep learning: A 526
479 comprehensive survey and benchmark”. In: 527
480 *Neurocomputing* 503 (2022), pp. 92–108.
- 481 [22] G. M. Kurtzer, V. Sochat, and M. W. 528
482 Bauer. “Singularity: Scientific containers for 529
483 mobility of compute”. In: *PloS one* 12.5 (2017), 530
484 e0177459.
- 485 [23] I. Daubechies, R. DeVore, S. Foucart, 531
486 B. Hanin, and G. Petrova. “Nonlinear 532
487 approximation and (deep) ReLU networks”. 533
488 In: *Constructive Approximation* 55.1 (2022), 534
489 pp. 127–172.
- 490 [24] J. Loo, M. Robbins, C. McNeil, T. 535
491 Yoshitake, C. Santori, C. Shan, S. 536
492 Vyawahare, H. Patel, T. C. Wang, R. 537
493 Findlater, et al. “Autofluorescence Virtual 538
494 Staining System for H&E Histology and 539
495 Multiplex Immunofluorescence Applied 540
to Immuno-Oncology Biomarkers in 541
Lung Cancer”. In: *Cancer Research* 542
Communications (2024).
- [25] U. Khan, S. Koivukoski, M. Valkonen, L. 543
Latonen, and P. Ruusuvoori. “The effect of 544
neural network architecture on virtual H&E 545
staining: Systematic assessment of histological 546
feasibility”. In: *Patterns* 4.5 (2023).