

# INCORPORATING STRUCTURAL MOTIFS AS WEAK PRIORS IN MOLECULAR LEARNING

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006 Paper under double-blind review  
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standard benchmarks remains competitive, demonstrating that interpretability and task-adaptivity do not sacrifice prediction quality.

Our main contributions are:

1. **Weakly supervised task-adaptive pre-training framework:** We introduce a weak supervision approach on chemically important motifs paired with an open source library, ChemCaption, for molecular featurization.
2. **Chemical interpretability:** Functional group clustering, feature attribution, and task-conditioned attention routing demonstrate that learned representations align with chemical concepts and can be visualized to understand model reasoning per task.
3. **Semantic task reasoning capabilities:** Zero-shot inference (up to 68% on novel tasks) through interpretable task embeddings—capabilities categorically unavailable to task-agnostic pre-trained models that require fine-tuning on each new task.
4. **Data efficient competitive performance:** CILT outperforms other task-agnostic models on the MoleculeNet benchmark across multiple versions of the model, showing that structured weak supervision substitutes for scale without sacrificing prediction quality.

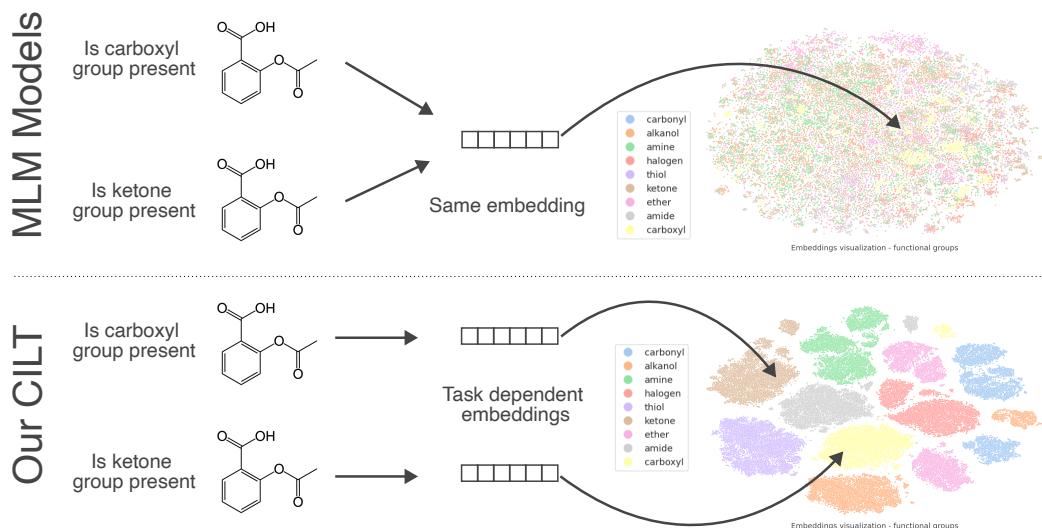


Figure 1: **Overview figure of different approaches.** Our CILT model can utilize the advantages of both deep learning and classical fingerprinting approaches.

## 2 BACKGROUND

**Group Contribution Methods** are a family of techniques for estimating molecular properties based on their substructural composition (Joback & Reid, 1987; Fredenslund et al., 1975). Molecules are decomposed into predefined structural groups, where each group has assigned empirically derived parameters that represent their contribution. These contributions are then combined, while accounting for the correction terms for group interactions, to form a property prediction. Chemists apply this method to this day to quickly and at scale estimate properties for mixture thermodynamics (Fredenslund et al., 1975), property estimation (Lydersen, 1955), drug discovery (Andrews et al., 1984), to name a few. Besides predictive power, thanks to hand-tuned features, predictions made with group-contribution approaches are very interpretable.

**Molecular Fingerprints** describe a molecule as a vector encoding the presence or count of predefined structural features. These fingerprints can then be used for fast similarity comparisons, forming the basis for structure-to-property predictive modeling. Machine learning models often offer

108 negligible gains compared to fingerprints while lacking interpretability and introducing additional  
 109 computational overhead (Praski et al., 2025; Boldini et al., 2024).  
 110

### 111 3 RELATED WORK 112

113 **Molecular Representation Learning** Molecular property prediction has been addressed through  
 114 diverse representation learning approaches. Sequence-based methods treat molecules as sequences,  
 115 typically using the SMILES notation (Weininger, 1988) or other line representations such as SELF-  
 116 IES (Krenn et al., 2022; 2020). Early work applied recurrent neural networks to SMILES (Segler  
 117 et al., 2018; Mayr et al., 2018; Goh et al., 2017), while more recent approaches use transformer  
 118 architectures with masked language modeling objectives (Ahmad et al., 2022; Chithrananda et al.,  
 119 2020; Ross et al., 2022; Fabian et al., 2020; Honda et al., 2019; Irwin et al., 2022; Born & Man-  
 120 icica, 2023). ChemBERTa (Chithrananda et al., 2020) adapts RoBERTa to molecular data, while  
 121 MolFormer (Ross et al., 2022) scales to more than a billion molecules using linear attention mecha-  
 122 nisms.  
 123

124 **Graph-based Approaches** One can represent molecules as molecular graphs with atoms as nodes  
 125 and bonds as edges. Message-passing neural networks (Gilmer et al., 2017; Scarselli et al., 2008)  
 126 form the foundation for many architectures. Self-supervised approaches include contrastive learning  
 127 methods like MolCLR (Wang et al., 2022b), GraphCL (You et al., 2020), GraphMAE (Hou et al.,  
 128 2022), and GROVER (Rong et al., 2020).  
 129

130 **Multi-Task and Auxiliary Supervision** Several approaches incorporate additional supervision  
 131 signals during pretraining. MolBERT (Fabian et al., 2020) combines masked language modeling  
 132 with auxiliary tasks such as descriptor prediction. ChemBERTa-2 (Ahmad et al., 2022) adds multi-  
 133 task regression on physico-chemical properties. MoMu (Su et al., 2022) trains jointly on molecular  
 134 graphs and natural language descriptions.  
 135

136 **Text-Molecule Joint Modeling** Recent works explore the joint modeling of natural language and  
 137 molecular representations. MolT5 (Edwards et al., 2022) adapts T5 to perform both molecule-to-  
 138 text and text-to-molecule generation tasks. Text2Mol (Edwards et al., 2021) learns cross-modal  
 139 embeddings between molecular graphs and textual descriptions. MoleculeSTM (Liu et al., 2022)  
 140 and CLAMP (Seidl et al., 2023) use contrastive learning between molecules and text. CLAMP  
 141 learns CLIP-style contrastive alignments between molecules and text to improve downstream activi-  
 142 ty prediction from natural language assay descriptions. Instruction-following approaches include  
 143 Galactica (Taylor et al., 2022), ether0 (Narayanan et al., 2025), and MolecularGPT (Liu et al., 2024).  
 144

145 **Task Conditioning and Prompting** In scientific domains, task conditioning appears in protein  
 146 modeling (Ferruz et al., 2022; Liu et al., 2023), drug design (Bagal et al., 2021; Born & Manica,  
 147 2023) and optimization (Wu et al., 2024). However, most molecular models use fixed task identifiers  
 148 or classification heads rather than natural language descriptions.  
 149

150 In summary, prior molecular pretraining has been largely optimized for token- or sequence-level  
 151 objectives on SMILES, often requiring massive corpora before substructure knowledge emerges.  
 152 We instead weakly supervise *on chemistry* via task-conditioned targets, derived via inexpensive  
 153 calculations described in natural language, and we couple this with a dual-masking objective that  
 154 ties text semantics to molecular structure. Empirically, this yields competitive accuracy with far  
 155 fewer pretraining molecules, strong few-shot transfer and high interpretability; theoretically, task-  
 156 similarity and motif-sparsity analyses explain when and why these gains appear.  
 157

### 158 4 CHEMICALLY INFORMED TASK CONDITIONING 159

#### 160 4.1 PROBLEM SETUP 161

162 We pre-train a single 150M-parameter transformer on hundreds of molecular motifs expressed as  
 163 natural language descriptors. Each task  $t$  has a programmatic supervision function  $g_t$  that extracts  
 164 chemical properties from molecules: substructure indicators (“contains halogen group”), counts  
 165 (“number of aromatic rings”), or simple properties (“molecular mass”).  
 166

162 We unify the tasks and molecules by encoding them into text and jointly passing them throughout  
 163 our network in the following form  
 164

$$\begin{array}{ccccccc} d & & [\text{SEP}] & y_t & [\text{SEP}] & x \\ \text{task description} & & & \text{value tokens} & & \text{SMILES} \end{array}$$

168 This format enables conditional training, the model learns to predict masked SMILES tokens given  
 169 properties and masked property values given SMILES, enabling a seamless switch between property  
 170 prediction and generation as well as addition of new tasks.  
 171

## 172 4.2 TRAINING OBJECTIVE

174 We train with two alternating masked language modeling objectives. The SMILES objective (Equation  
 175 (1)) teaches the model to generate molecules conditioned on task descriptions and target prop-  
 176 erty values:

$$\mathcal{L}_{\text{SMILES}}(\theta) = \mathbb{E}_{t,x,M_x} \left[ - \sum_{i \in M_x} \log p_{\theta}(x_i | x_{\setminus i}, y_t, d_t) \right] \quad (1)$$

180 The property value objective (Equation (2)) teaches property prediction conditioned on molecular  
 181 structure and task description:

$$\mathcal{L}_{\text{value}}(\theta) = \mathbb{E}_{t,x,M_y} \left[ - \sum_{j \in M_y} \log p_{\theta}(y_{t,j} | x, d_t) \right] \quad (2)$$

187 This bidirectional training creates a unified architecture for conditional generation, regression, and  
 188 classification driven entirely by natural language prompts.

## 190 4.3 THEORETICAL FOUNDATIONS

192 We provide theoretical justification for two key claims: why semantic similarity between task de-  
 193 scriptions should predict transfer performance, and motif pretraining tasks should improve sample  
 194 efficiency.

### 196 4.3.1 TASK SIMILARITY CONTROLS TRANSFER

197 We first formalize the intuition that semantically similar task descriptions should enable better zero-  
 198 shot transfer. We define this semantic similarity as the cosine similarity between task description  
 199 embeddings:  $s(d, d') = \langle e(d), e(d') \rangle$ .

200 **Theorem 1** (Task-Semantic Adaptation Bound). *Under standard Lipschitz and bounded loss as-  
 201 sumptions, the domain error  $R$  on a target task  $d'$  is bounded by:*

$$R_{d'}(h) \leq \underbrace{\sum_{t=1}^T \alpha_t R_{d_t}(h)}_{\text{weighted source risk}} + \underbrace{L \sum_{t=1}^T \alpha_t \|e(d') - e(d_t)\|}_{\text{task geometry term}} + \underbrace{\mathcal{O}(\sqrt{1/n})}_{\text{few-shot term}} \quad (3)$$

208 for any convex combination of source tasks  $\{\alpha_t\}$  and constant  $L > 0$ .

210 Where  $R_{d_t}(h)$  represents the model's source domain error (on the pre-training tasks) while  $R_{d'}(h)$   
 211 represents model's target domain error (on new tasks). The task embedding is represented as  $e(d)$   
 212 and  $\alpha_t$  is chosen via softmax over distance  $\alpha_t = \text{softmax}(\|e(d') - e(d_t)\|)$ .

213 *Proof.* See section section A.1.

215 The task geometry term shows that transfer performance degrades with the distance between task  
 216 embeddings. For unit-norm embeddings,  $\|e(d') - e(d_t)\|^2 = 2(1 - \cos \angle(e(d'), e(d_t)))$ , higher

216 cosine similarity implies better transfer when the weighted source domain errors are appropriately  
 217 regularized. This provides theoretical backing for our empirical observation that zero-shot performance  
 218 correlates with semantic similarity and indicates the number of shots needed to adapt to a  
 219 re-phrased or related task (see Section 6.2).  
 220

221 **4.3.2 MOTIF PRETRAINING IMPROVES SAMPLE EFFICIENCY**  
 222

223 Next, we establish that when molecular properties depend on sparse combinations of motifs (e.g.,  
 224 functional groups) weak supervision on chemical motifs dramatically reduces sample complexity.  
 225 This is a chemically informed prior based on the realization that chemists have achieved much  
 226 success with so-called group contribution methods (Gani, 2019; Kühne et al., 1995; Constantinou  
 227 & Gani, 1994; Fredenslund, 2012), where a property is predicted based on a linear or higher-order  
 228 combination of group-specific factors (see Background section).

229 Suppose the pre-trained representations are *motif-aligned*, where motifs might correspond to functional  
 230 group features, and suppose downstream molecular properties depend on sparse combinations  
 231 of  $k \ll p$  motifs, as suggested by the group contribution method. Under standard sparse regression  
 232 assumptions:

233 **Theorem 2** (Motif Sample Complexity). *When molecular properties depend on  $k$  motifs out of  $p$  total features, explicit motif supervision reduces sample complexity from  $\tilde{\mathcal{O}}(p/\varepsilon^2)$  to  $\tilde{\mathcal{O}}(k \log p/\varepsilon^2)$  for achieving prediction error  $\varepsilon$ .*  
 234

235 *Proof.* See section section A.2.  
 236

237 **5 METHODS**  
 238

239 **5.1 DATASET CONSTRUCTION**  
 240

241 We construct our pretraining dataset by programmatically generating chemical task-property pairs  
 242 from half a million diverse molecules from ChemPile-MLift (Mirza et al., 2025) using the Chem-  
 243 Caption package, which interfaces with RDKit (Landrum, 2006). Our property set spans atom and  
 244 bond counts, manually curated functional group indicators, ring system features, molecular descrip-  
 245 tors, hydrogen bonding patterns, and substructure motifs. This yields over 300 distinct chemical  
 246 properties per molecule.  
 247

248 Task descriptions are generated using templated natural language patterns. Task descriptions use  
 249 templates like “does the molecule contain `<PROPERTY_NAME>`” or “what is the  
 250 `<PROPERTY_NAME>`”, or “number of `<PROPERTY_NAME>`”. Property values are serialized  
 251 as text tokens: binary values as “1”/“0”, integers directly, and continuous values are first normalized  
 252 and then quantized to four decimal places. This process generates approximately 150 million task-  
 253 molecule pairs.  
 254

255 **5.2 MODEL ARCHITECTURE AND TRAINING**  
 256

257 We employ a 150M-parameter ModernBERT architecture (Warner et al., 2025) with a shared vo-  
 258 cabulary combining SMILES tokens derived using a regular expression-based tokenizer (Schwaller  
 259 et al., 2018), as well as natural language tokens, and numerical value tokens derived from the  
 260 ModernBERT tokenizer. Input sequences follow the format `[task description] [SEP]`  
 261 `[property value] [SEP] [SMILES]` with a maximum sequence length of 1024. Through-  
 262 out all of the experiments no sequence has exceeded this limit.  
 263

264 Training alternates between the SMILES objective (Equation (1)) and the property prediction objec-  
 265 tive (Equation (2)) every 20 batch steps. The property prediction objective masks the entire property  
 266 value and predicts it conditioned on the task description and SMILES sequence. The SMILES  
 267 completion objective randomly masks 25% of the SMILES tokens and predicts them conditioned on the  
 268 description of the task and the value of the property. Both objectives use cross-entropy loss with uni-  
 269 form task sampling across our property collection. We train the model for 3 epochs, for parameter  
 breakdown see Section A.5.

270 5.3 BASELINES  
271

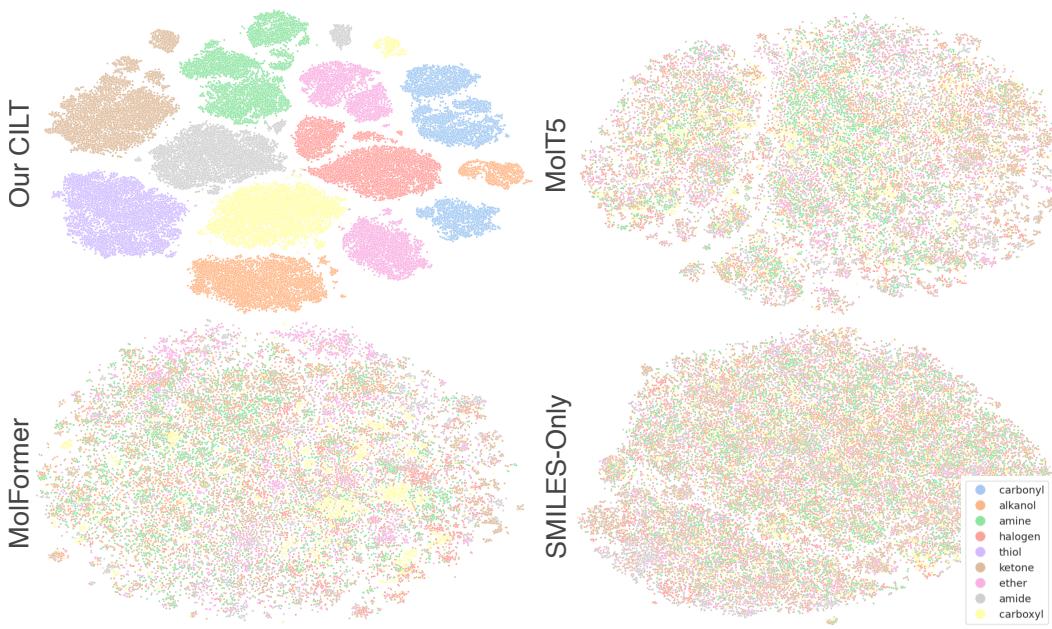
272 For comparison, we consider the following leading large chemical pretrained models: Mol-  
273 CLR (Wang et al., 2022a), ChemBERTa (Chithrananda et al., 2020), MolFormer (Ross et al.,  
274 2022), MolBert (Fabian et al., 2020), Grover (Rong et al., 2020), MolT5 (Edwards et al., 2022)  
275 and MoleculeSTM (Liu et al., 2022). We test all models on the MoleculeNet benchmark (Wu et al.,  
276 2018) and photoswitch dataset (Griffiths et al., 2022) (detailed description can be found in Sec-  
277 tion A.3.1 and Section A.3.2, respectively).

278 In the linear probe experiments, we train linear regression models for the regression tasks and lo-  
279 gistic regression models for the classification tasks. For both, we utilize  $L_1$  regularization (with  
280 optimal parameters see section A.2), additionally, for the logistic regression we employ the liblinear  
281 solver and balanced class weights. For all experiments, we use 4-fold cross-validation with scaffold  
282 splitting.

283

284 6 EXPERIMENTS AND RESULTS  
285

286 To demonstrate the effectiveness of our method, we evaluate CILT on multiple standard benchmarks  
287 in multiple systematic experiments: **a) embedding alignment** assessing the alignment of embed-  
288 dings with chemically relevant features; **b) zero-/few-shot transfer** evaluating the performance of  
289 CILT on unseen tasks and the amount of data needed for adaptation to these tasks; **c) linear probes**  
290 comparing embeddings across different models to evaluate innate learned molecular representations;  
291 **d) ablations** for targeted assessment of our training methodology.



314 Figure 2: **Visualization of learned embeddings represented via t-SNE.** Representations are ex-  
315 tracted from the hold-out test set (scaffold-split) used for the pre-training of CILT. The models used  
316 for comparisons are MolFormer (Ross et al., 2022), MolT5 (Edwards et al., 2022) and a SMILES-  
317 only pre-trained version of CILT.

318

319

320 6.1 REPRESENTATIONS ALIGNMENT WITH FUNCTIONAL GROUPS  
321

322 **Experiment** To show the benefit of pre-training with molecular fingerprints as a weak prior we  
323 compare the embeddings across CILT, SMILES only trained version of CILT, MolFormer (Ross  
et al., 2022) and MolT5 (Edwards et al., 2022) Figure 2.

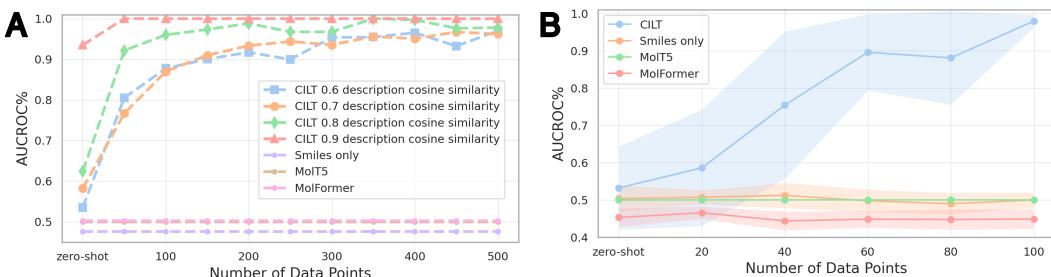
324 **Results** Figure 2 show that CILT’s embeddings cluster align with the presence and absence of  
 325 functional groups, while the classical MLM approaches and multimodal modeling are not able to  
 326 make this distinction. This indicates that the model can capture relevant chemical features that  
 327 are known to chemists and therefore offers a higher level of interpretability. This gives additional  
 328 support to our assumption that we induce motif-aligned coordinates in our representation (see The-  
 329 orem 2). For a full breakdown of per-functional group embeddings see Section A.7.

330 Additionally, we also find attention patterns to show chemically meaningful behaviors (Section A.6).  
 331 Chemically relevant atoms have higher attention scores, and attention patterns link the task to the  
 332 property and then to relevant atoms.  
 333

## 334 6.2 ZERO-SHOT TRANSFER

335

336 **Experiment** Theorem 1 predicts that semantically similar task descriptions should enable better  
 337 zero-shot transfer. To evaluate this, we conducted an experiment on a subset of functional group  
 338 presence tasks. We rephrase the original template 20 times (see Section A.4) and measure the  
 339 cosine similarity between the new and original task description. We then group the tasks by cosine  
 340 similarity and evaluate the model on them. First, we measure the zero-shot performance, and then  
 341 we gradually increase the number of fine-tuning data points until all of the tasks converge. Baseline  
 342 models do not have the ability to adapt to new task descriptions, therefore we report only their  
 343 zero-shot performance.  
 344



354  
 355 **Figure 3: Adaptation to new tasks. A** Adaptation to the new task description for the already seen  
 356 task based on the cosine similarity to the original task description. Baseline models do not have  
 357 the ability to adapt to new task descriptions, their zero-shot performance is reported. **B** Required  
 358 number of data points to adapt to the unseen tasks across 12 methylations.

359

360 **Results** Figure 3 A shows that across all of the datasets the cosine similarity is correlated both  
 361 in the zero-shot performance and the adaptation setting. We see that the datasets with higher co-  
 362 sine similarity between the new task description and the original task description from pretraining  
 363 adapt with fewer data points. This gives support to our assumptions that semantically similar task  
 364 descriptions should enable better zero-shot transfer. Furthermore, none of the baseline models show  
 365 zero-shot performance better than random, indicating that the models are not capable of distinguishing  
 366 between different functional groups.  
 367

## 368 6.3 FEW-SHOT TRANSFER

369

370 **Experiment** Theorem 2 predicts that motif-alignment leads to more data efficient learning. We  
 371 test this by altering the original task. We perform methylations (replacing one H with  $\text{CH}_3$ ) on  
 372 the substructures that CILT has been trained to understand. We gathered 15 of these new tasks to  
 373 evaluate our model. After evaluating the zero-shot, we gradually increase the number of training  
 374 points by 20 (10 positive and 10 negative samples) until our models converge. For the baseline  
 375 models, we freeze the backbone and replace the last layer with the prediction head and fine-tune the  
 376 model as a binary classifier.  
 377

378 **Results** Figure 3 B shows that across all of the methylations, CILT can fine-tune with less than  
 379 100 samples and even perform zero-shot inference in some settings. This gives support to our

assumptions that motif alignment leads to more data-efficient learning. Furthermore, none of the baseline models show performance significantly better than random at any step of the fine-tuning process.

## 6.4 TRANSFERABILITY OF THE EMBEDDINGS

**Experiment** We assess the robustness and transferability of the embeddings of CILT and other baseline encoders using linear probing (Alain & Bengio, 2016). We report the %AUCROC for classification tasks and MAE for regression tasks along with the standard deviations.

**Table 1: Embedding quality estimated using linear probes.** Logistic regression and linear regression trained on embeddings over 4-fold cross-validation scaffold split. For classification we report %AUCROC ( $\uparrow$ ) and for regression MAE ( $\downarrow$ ). The best results in each column are bolded and the second best are underlined. CILT is the best model for classification tasks.

Model	Classification (%AUCROC $\uparrow$ )								
	BACE	BBBP	ClinTox	HIV	SIDER	Tox21	ToxCast	MUV	Avg.
MolCLR	73.4 $\pm$ 3.6	82.42 $\pm$ 2.1	70.5 $\pm$ 3.7	71.2 $\pm$ 0.9	58.9 $\pm$ 4.8	69.7 $\pm$ 7.6	62.5 $\pm$ 10.1	70.54 $\pm$ 13.9	69.9
ChemBERTa	80.0 $\pm$ 3.6	88.0 $\pm$ 2.2	97.2 $\pm$ 1.5	73.9 $\pm$ 1.9	54.1 $\pm$ 6.0	67.8 $\pm$ 6.8	64.0 $\pm$ 10.5	72.8 $\pm$ 11.1	74.7
MolFormer	74.3 $\pm$ 2.1	89.8 $\pm$ 1.0	97.2 $\pm$ 1.5	73.9 $\pm$ 0.9	55.8 $\pm$ 5.1	68.0 $\pm$ 6.2	65.3 $\pm$ 10.2	71.9 $\pm$ 15.7	74.5
Grover	<b>84.2 <math>\pm</math> 3.8</b>	84.1 $\pm$ 0.8	82.8 $\pm$ 3.1	<b>78.5 <math>\pm</math> 2.3</b>	56.7 $\pm$ 6.6	71.3 $\pm$ 6.6	67.0 $\pm$ 10.7	73.8 $\pm$ 12.6	75.0
MolBERT	81.0 $\pm$ 4.2	82.9 $\pm$ 2.2	77.9 $\pm$ 6.3	75.4 $\pm$ 2.2	56.9 $\pm$ 4.6	70.4 $\pm$ 6.9	63.9 $\pm$ 10.4	<b>76.2 <math>\pm</math> 12.8</b>	73.1
MolT5	81.9 $\pm$ 3.5	94.3 $\pm$ 1.6	97.4 $\pm$ 2.7	75.8 $\pm$ 1.6	<b>60.3 <math>\pm</math> 7.8</b>	<b>74.0 <math>\pm</math> 6.7</b>	<b>69.9 <math>\pm</math> 10.4</b>	74.0 $\pm$ 13.9	<b>78.4</b>
MoleculeSTM	73.7 $\pm$ 4.2	87.6 $\pm$ 1.9	<u>98.0 <math>\pm</math> 0.6</u>	71.1 $\pm$ 1.0	56.3 $\pm$ 5.2	69.6 $\pm$ 6.2	64.2 $\pm$ 10.7	67.4 $\pm$ 11.8	73.5
CILT(500k)	80.4 $\pm$ 1.2	92.5 $\pm$ 1.2	97.7 $\pm$ 1.5	73.9 $\pm$ 1.5	55.2 $\pm$ 6.3	66.3 $\pm$ 6.9	64.4 $\pm$ 10.3	71.9 $\pm$ 13.7	75.3
CILT(250k)	81.3 $\pm$ 2.5	<b>94.5 <math>\pm</math> 1.3</b>	<b>98.3 <math>\pm</math> 0.1</b>	75.6 $\pm$ 0.7	58.5 $\pm$ 6.8	<u>72.5 <math>\pm</math> 6.0</u>	<u>68.0 <math>\pm</math> 11.2</u>	<u>75.2 <math>\pm</math> 12.3</u>	78.0
Regression (MAE $\downarrow$ )									
Model	Lipo	ESOL	FreeSolv	CAM	PBE0	En $- \pi^*$	Eπ $- \pi^*$	Zn $- \pi^*$	Rank
MolCLR	1.00 $\pm$ 0.04	1.03 $\pm$ 0.09	1.16 $\pm$ 0.34	36.7 $\pm$ 21.3	<b>37.5 <math>\pm</math> 7.9</b>	25.8 $\pm$ 12.9	50.5 $\pm$ 7.7	<u>13.8 <math>\pm</math> 5.3</u>	3.0
ChemBERTa	0.81 $\pm$ 0.30	0.82 $\pm$ 0.73	0.86 $\pm$ 0.27	34.2 $\pm$ 21.1	43.4 $\pm$ 16.1	26.7 $\pm$ 12.3	<b>47.3 <math>\pm</math> 10.6</b>	<u>13.8 <math>\pm</math> 5.3</u>	2.0
MolFormer	0.81 $\pm$ 0.04	0.83 $\pm$ 0.73	0.88 $\pm$ 0.23	43.1 $\pm$ 12.3	55.2 $\pm$ 14.2	26.9 $\pm$ 12.3	50.9 $\pm$ 9.1	<u>13.8 <math>\pm</math> 5.3</u>	3.8
Grover	0.81 $\pm$ 0.03	0.82 $\pm$ 0.73	<b>0.85 <math>\pm</math> 0.27</b>	39.8 $\pm$ 23.3	44.6 $\pm$ 18.0	<b>23.5 <math>\pm</math> 8.7</b>	67.5 $\pm$ 11.1	<u>16.5 <math>\pm</math> 5.2</u>	3.0
MolBERT	1.00 $\pm$ 0.04	1.03 $\pm$ 0.08	1.64 $\pm$ 0.34	47.0 $\pm$ 25.8	41.5 $\pm$ 21.8	31.0 $\pm$ 11.3	58.6 $\pm$ 10.3	16.6 $\pm$ 5.0	5.4
MolT5	0.81 $\pm$ 0.03	0.82 $\pm$ 0.73	0.86 $\pm$ 0.27	<b>33.3 <math>\pm</math> 17.7</b>	43.7 $\pm$ 15.2	24.7 $\pm$ 13.5	47.4 $\pm$ 12.1	<u>13.8 <math>\pm</math> 5.3</u>	<b>1.9</b>
MoleculeSTM	0.81 $\pm$ 0.03	<u>0.82 <math>\pm</math> 0.73</u>	<u>0.86 <math>\pm</math> 0.27</u>	44.1 $\pm$ 15.3	55.0 $\pm$ 12.1	27.3 $\pm$ 12.0	50.6 $\pm$ 7.8	<u>13.8 <math>\pm</math> 5.3</u>	3.6
CILT(500k)	<b>0.80 <math>\pm</math> 0.02</b>	0.88 $\pm$ 0.18	0.91 $\pm$ 0.30	46.9 $\pm$ 15.5	58.5 $\pm$ 7.6	27.5 $\pm$ 12.0	51.3 $\pm$ 7.3	13.9 $\pm$ 5.2	4.9
CILT(250k)	<u>0.81 <math>\pm</math> 0.02</u>	0.90 $\pm$ 0.18	0.91 $\pm$ 0.30	49.1 $\pm$ 19.1	65.8 $\pm$ 7.0	27.5 $\pm$ 12.0	51.3 $\pm$ 7.3	13.9 $\pm$ 5.2	5.1

**Results** Table 4 shows that CILT demonstrates competitive performance across all the datasets. In the classification setting, it achieves the 2 best and 3 second best scores, while in the regression setting, it shows the second-to-last performance. We theorise that the lackluster performance on regression tasks comes from pre-training being dominated by classification tasks.

## 6.5 ABLATIONS

**Experiment** To isolate the effect of task conditioning, we train a control model using identical architecture and hyperparameters but with standard masked language modeling on SMILES sequences only, without task descriptions or property values. This control methodology represents conventional molecular pretraining approaches like ChemBERTa and MolFormer.

We evaluate both the task-conditioned model and the SMILES-only baseline on the same downstream benchmarks using identical fine-tuning protocols.

**Results** Table 2 shows that task-conditioned pretraining outperforms SMILES-only pretraining on 15 out of 16 tasks across two benchmark datasets. This confirms that our chemically meaningful pretraining tasks provide measurable benefits over standard molecular language modeling.

## 7 DISCUSSION

**Parameter–Performance Frontier** In Figure 4, we plot the average classification performances from the linear probe experiments (Section 6.4) and compare them against the log number of

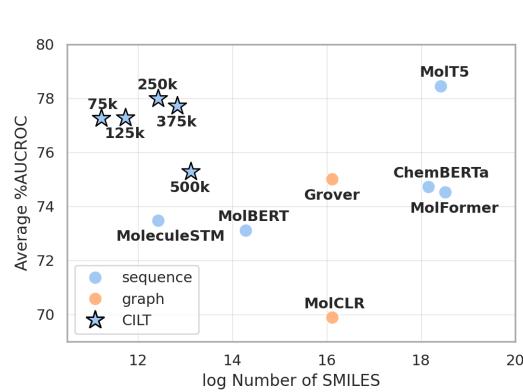
432  
 433 **Table 2: Ablation results.** Logistic regression and linear regression trained on embeddings over a  
 434 4-fold cross-validation scaffold split. For classification we report %AUCROC ( $\uparrow$ ) and for regression  
 435 MAE ( $\downarrow$ ). The best results are bolded. We find that CILT outperforms the SMILES-only model on  
 436 both classification and regression tasks.

Classification (%AUCROC $\uparrow$ )									
Model	BACE	BBBP	ClinTox	HIV	SIDER	tox21	ToxCast	MUV	Mean
SmilesOnly	74.7 $\pm$ 2.3	90.5 $\pm$ 1.1	97.3 $\pm$ 2.0	70.1 $\pm$ 1.2	55.2 $\pm$ 6.1	65.7 $\pm$ 6.6	63.4 $\pm$ 10.1	68.7 $\pm$ 13.7	73.2
CILT(500k)	80.4 $\pm$ 1.2	92.5 $\pm$ 1.2	97.7 $\pm$ 1.5	73.9 $\pm$ 1.5	55.2 $\pm$ 6.3	66.3 $\pm$ 6.9	64.4 $\pm$ 10.3	71.9 $\pm$ 13.7	75.3
CILT(250k)	<b>81.3 <math>\pm</math> 2.5</b>	<b>94.5 <math>\pm</math> 1.3</b>	<b>98.3 <math>\pm</math> 0.1</b>	<b>75.6 <math>\pm</math> 0.7</b>	<b>58.5 <math>\pm</math> 6.8</b>	<b>72.5 <math>\pm</math> 6.0</b>	<b>68.0 <math>\pm</math> 11.2</b>	<b>75.2 <math>\pm</math> 12.3</b>	<b>78.0</b>

Regression (MAE $\downarrow$ )									
Model	Lipo	FreeSolv	ESOL	CAM	PBE0	$E\pi - \pi^*$	$E\pi - \pi^*$	$Zn - \pi^*$	Rank
SmilesOnly	0.81 $\pm$ 0.03	0.89 $\pm$ 0.07	0.91 $\pm$ 0.18	49.2 $\pm$ 16.9	77.4 $\pm$ 15.3	30.0 $\pm$ 11.9	62.8 $\pm$ 6.9	17.1 $\pm$ 4.8	2.6
CILT(500k)	<b>0.80 <math>\pm</math> 0.02</b>	<b>0.88 <math>\pm</math> 0.18</b>	<b>0.91 <math>\pm</math> 0.30</b>	<b>46.9 <math>\pm</math> 15.5</b>	<b>58.5 <math>\pm</math> 7.6</b>	<b>27.5 <math>\pm</math> 12.0</b>	<b>51.3 <math>\pm</math> 7.3</b>	<b>13.9 <math>\pm</math> 5.2</b>	<b>1.1</b>
CILT(250k)	0.81 $\pm$ 0.02	0.90 $\pm$ 0.18	0.91 $\pm$ 0.30	49.1 $\pm$ 19.1	65.8 $\pm$ 7.0	27.5 $\pm$ 12.0	51.3 $\pm$ 7.3	13.9 $\pm$ 5.2	1.6

449 molecules used in pre-training. Our model CILT shows competitive performance across multiple  
 450 versions while only requiring a fraction of molecules. This challenges the assumption that sequence-  
 451 based molecular foundation models need to be trained on a huge number of molecules to work well.



467 **Figure 4: Log number of pretraining molecules**  
 468 **vs. downstream performance.** We show the  
 469 number of molecules used in pretraining of base-  
 470 line models and CILT vs. the average classifi-  
 471 cation performance of linear probes on Molecul-  
 472 eNet. CILT shows the best tradeoff between  
 473 dataset size and performance.

**Meaningful Representations Through Soft Inductive Biases.** Our approach succeeds by implementing soft inductive biases—preferences for certain solutions without hard constraints (Wilson, 2025). Rather than restricting the model architecture, we guide learning through natural language task conditioning. This creates representations that cluster by chemically important features without explicit supervision, while attention mechanisms focus on chemically relevant atoms when processing task descriptions. Our theoretical analysis shows that semantic similarity between task descriptions directly predicts transfer performance (Theorem 2), while Theorem 1 formalizes how motif-based supervision reduces sample complexity from  $\mathcal{O}(p)$  to  $\mathcal{O}(k \lg p)$ . The model learns chemical intuition not as an emergent property by scaling data, but as an explicit objective encoded through structured tasks.

### Task Conditioning as Architectural Innovation

The natural language conditioning framework offers practical advantages beyond efficiency. Unlike approaches that require architectural changes for new properties and downstream applications, our text-based task descriptions enable immediate extensibility. New chemical tasks can be incorporated without re-training by simply providing appropriate natural language descriptions, making the system immediately adaptable to new chemical properties.

483 **Future Directions** The current CILT model is pre-trained on a naive selection of motifs and task  
 484 descriptions; therefore, the next future step would be to improve the selection of pre-training motifs  
 485 and rephrase the task descriptions (Maini et al., 2024; Pieler et al., 2024). The semantic similarity  
 results also suggest principled curriculum learning possibilities.

486 8 CONCLUSIONS  
487488 Foundation models (White, 2023; Ramos et al., 2025; Alampara et al., 2025) for scientific domains  
489 commonly follow the standard approach following the NLP blueprint: scale data and parameters  
490 until patterns emerge (Frey et al., 2023). But scientific domains differ fundamentally from language.  
491 Chemical datasets are small, diverse, and experimental data is expensive. But scientific domains  
492 possess structured theoretical knowledge that language modeling lacks. In chemistry, for instance,  
493 this has been encoded over decades via QSPR relationships and group contribution theory. Rather  
494 than rediscovering them from data, we can use them as a weak supervision signal.495 We demonstrate that chemically-informed pretraining achieves competitive performance with orders  
496 of magnitude less data. By encoding chemical priors as soft inductive biases through natural lan-  
497 guage task conditioning, CILT learns interpretable representations that respect chemical structure  
498 while enabling rapid adaptation to new tasks.499 Our approach of pre-training on a broad basis of weakly supervised tasks in multiple masking ob-  
500 jectives might be a recipe for other domains where there is little data, but one can generate tasks  
501 with some weak-supervision-like techniques.  
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## 724 A APPENDIX

### 725 A.1 PROOF OF THEOREM 1: TASK-SEMANTIC ADAPTATION BOUND

726 We prove that the domain error on a target task can be bounded in terms of source task performance  
 727 plus a term that depends on the semantic similarity between task descriptions. The key insight is to  
 728 use optimal transport theory to relate distributional differences to task embedding distances.

729 Let  $P_d$  denote the joint distribution of motif-task pairs  $(\phi_\theta(X, d), g_d(X))$  for task  $d$ , where  $X \sim \mathcal{D}$ .  
 730 The domain error under task  $d$  is  $R_d(h) = \mathbb{E}_{(Z, Y) \sim P_d} [\ell(h(Z), Y)]$ .

731 **Step 1: Kantorovich-Rubinstein bound.** We want to bound the difference in domain error be-  
 732 tween the target task  $d'$  and a weighted combination of source tasks  $d$ . Since the loss function  $\ell$   
 733 is  $L_f$ -Lipschitz by assumption, we can apply the Kantorovich-Rubinstein duality, which provides a  
 734 connection between differences in expectations and Wasserstein distances (Villani, 2008):

$$735 \left| R_{d'}(h) - \sum_{t=1}^T \alpha_t R_{d_t}(h) \right| = \left| \mathbb{E}_{P_{d'}} [\ell(h(Z), Y)] - \mathbb{E}_{\sum_t \alpha_t P_{d_t}} [\ell(h(Z), Y)] \right| \leq L_f W_1 \left( P_{d'}, \sum_{t=1}^T \alpha_t P_{d_t} \right).$$

736 This converts the problem from bounding differences in domain errors to bounding Wasserstein  
 737 distances between distributions (which is a geometric problem about the learned representations).

738 **Step 2: Pushforward representation.** The joint distributions  $P_d$  arise from our specific model  
 739 architecture. We can represent them as pushforwards of simpler distributions through our learned  
 740 mapping.

741 Define the map  $\Psi : (x, u) \mapsto (\phi_\theta(x, d(u)), g_{d(u)}(x))$  that transforms molecules and task embed-  
 742 dings into motifs and tasks. This map encapsulates both our learned representation function and the  
 743 ground truth property computation.

756 Since each task  $d$  corresponds to a fixed task embedding  $e(d)$ , we can write:  
 757

$$758 \quad P_d = \Psi_{\#}(\mathcal{D} \otimes \delta_{e(d)}),$$

759  
 760 where  $\Psi_{\#}$  denotes the pushforward measure. This means the distribution  $P_d$  is obtained by taking  
 761 the product of the molecular distribution  $\mathcal{D}$  with a point mass at the task embedding  $e(d)$ , then  
 762 applying the transformation  $\Psi$ .

763 For the weighted combination of source distributions:  
 764

$$765 \quad \sum_{t=1}^T \alpha_t P_{d_t} = \Psi_{\#} \left( \mathcal{D} \otimes \sum_{t=1}^T \alpha_t \delta_{e(d_t)} \right).$$

766  
 767  
 768 **Step 3: Wasserstein contraction.** Now we can use the property that the Wasserstein distance  
 769 contracts under Lipschitz maps. By assumption, the map  $\Psi$  is  $L_{\Psi}$ -Lipschitz in the task embedding  
 770 component. This means that if two task embeddings are close, the resulting motif-task distributions  
 771 will also be close.

772 The contraction property gives us:  
 773

$$774 \quad W_1 \left( P_{d'}, \sum_{t=1}^T \alpha_t P_{d_t} \right) \leq L_{\Psi} W_1 \left( \mathcal{D} \otimes \delta_{e(d')}, \mathcal{D} \otimes \sum_{t=1}^T \alpha_t \delta_{e(d_t)} \right)$$

775  
 776 Since the molecular distribution  $\mathcal{D}$  is the same in both cases, the Wasserstein distance only depends  
 777 on the task embedding component:  
 778

$$779 \quad W_1 \left( \delta_{e(d')}, \sum_{t=1}^T \alpha_t \delta_{e(d_t)} \right) = \sum_{t=1}^T \alpha_t \|e(d') - e(d_t)\|$$

780 The distributional distance between tasks thus reduces to the geometric distance between their em-  
 781 beddings. This justifies why semantic similarity should predict transfer performance.  
 782

783  
 784 **Step 4: Finite-sample bound.** Finally, we need to account for the fact that we only have finite  
 785 samples from the target task. The standard approach uses Rademacher complexity to bound the  
 786 gap between empirical and population risk. For bounded loss functions and hypothesis class  $\mathcal{H}$ ,  
 787 concentration inequalities give (Mohri et al., 2018):  
 788

$$789 \quad R_{d'}(h) \leq \hat{R}_{d'}(h) + 2\mathfrak{R}_n(\mathcal{H}) + 3\sqrt{\frac{\ln(2/\delta)}{2n}}$$

790 with probability at least  $1 - \delta$ , where  $\hat{R}_{d'}(h)$  is the empirical domain error on the target task.  
 791

792 Combining all steps and minimizing the empirical term over  $h \in \mathcal{H}$  yields the bound in Theorem 1.  
 793 The interpretation is that target task performance is bounded by a weighted combination of source  
 794 performance, plus a penalty term proportional to the distance between task embeddings, plus a  
 795 finite-sample correction.  $\square$

## 801 A.2 PROOF OF THEOREM 2: MOTIF SAMPLE COMPLEXITY

802 We analyze when explicit motif supervision can reduce sample complexity compared to standard  
 803 dense regression. The key insight is that chemical properties often depend on sparse combinations  
 804 of motifs, making this a sparse regression problem where  $k \ll p$  motifs matter.  
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806  
 807 **Setup and intuition.** Consider a pretrained encoder that produces representations  $\psi_{\theta}(x) \in \mathbb{R}^p$   
 808 that are *motif-aligned*—meaning different coordinates respond to different motifs. If downstream  
 809 molecular properties depend on only  $k$  out of  $p$  possible motifs, then the optimal linear head  $w^*$   
 810 should be  $k$ -sparse.

810 We analyze the LASSO estimator (Tibshirani, 1996), which is designed to recover sparse solutions:  
 811

$$812 \hat{w} \in \arg \min_{w \in \mathbb{R}^p} \frac{1}{2n} \|y - \Psi w\|_2^2 + \lambda \|w\|_1$$

813 where  $\Psi \in \mathbb{R}^{n \times p}$  stacks rows  $\psi_\theta(x_i)^\top$  and  $y_i = w^* \top \psi_\theta(x_i) + \xi_i$ .  
 814

815 **Step 1: Basic inequality.** The proof follows the standard template for LASSO analysis. By optimality of  $\hat{w}$ , it achieves lower objective value than the true parameter  $w^*$ :  
 816

$$817 \frac{1}{2n} \|y - \Psi \hat{w}\|_2^2 + \lambda \|\hat{w}\|_1 \leq \frac{1}{2n} \|y - \Psi w^*\|_2^2 + \lambda \|w^*\|_1$$

818 Since  $y = \Psi w^* + \xi$  where  $\xi$  is noise, we can expand and simplify to get:  
 819

$$820 \frac{1}{2n} \|\Psi \Delta\|_2^2 \leq \frac{1}{n} \xi^\top \Psi \Delta + \lambda (\|w^*\|_1 - \|\hat{w}\|_1),$$

821 where  $\Delta = \hat{w} - w^*$  is the estimation error.  
 822

823 The left side is the prediction error, while the right side has a stochastic term and a regularization term.  
 824

825 **Step 2: Controlling the stochastic term.** The term  $\frac{1}{n} \xi^\top \Psi \Delta$  involves the noise and is the main source of randomness. We can bound it using the dual norm relationship:  
 826

$$827 \frac{1}{n} \xi^\top \Psi \Delta = \left\langle \frac{1}{n} \Psi^\top \xi, \Delta \right\rangle \leq \left\| \frac{1}{n} \Psi^\top \xi \right\|_\infty \|\Delta\|_1$$

828 Since the noise  $\xi$  is sub-Gaussian, concentration inequalities tell us that with high probability:  
 829

$$830 \left\| \frac{1}{n} \Psi^\top \xi \right\|_\infty \leq C\sigma \sqrt{\frac{\log p}{n}}$$

831 We choose the regularization parameter  $\lambda$  to be twice this bound, so that:  
 832

$$833 \frac{1}{n} \xi^\top \Psi \Delta \leq \frac{\lambda}{2} \|\Delta\|_1$$

834 This is a standard technique in high-dimensional statistics: choose  $\lambda$  large enough to dominate the stochastic fluctuations.  
 835

836 **Step 3: Decomposability and cone constraint.** Now we analyze the regularization term  $\|w^*\|_1 - \|w\|_1$ . Since  $w^*$  is  $k$ -sparse with support  $S = \text{supp}(w^*)$ , we can decompose:  
 837

$$838 \|w^*\|_1 - \|\hat{w}\|_1 = \|w_S^*\|_1 - \|\hat{w}_S\|_1 - \|\hat{w}_{S^c}\|_1$$

839 Using the reverse triangle inequality  $\|a\|_1 - \|a + b\|_1 \leq \|b\|_1$ :  
 840

$$841 \|w_S^*\|_1 - \|\hat{w}_S\|_1 = \|w_S^*\|_1 - \|w_S^* + \Delta_S\|_1 \leq \|\Delta_S\|_1$$

842 Therefore:  $\|w^*\|_1 - \|\hat{w}\|_1 \leq \|\Delta_S\|_1 - \|\Delta_{S^c}\|_1$ .  
 843

844 Combining with the previous steps gives:  
 845

$$846 \frac{1}{2n} \|\Psi \Delta\|_2^2 \leq \frac{\lambda}{2} \|\Delta\|_1 + \lambda (\|\Delta_S\|_1 - \|\Delta_{S^c}\|_1) = \frac{3\lambda}{2} \|\Delta_S\|_1 - \frac{\lambda}{2} \|\Delta_{S^c}\|_1$$

847 Rearranging:  $\|\Delta_{S^c}\|_1 \leq 3\|\Delta_S\|_1$  (cone constraint).  
 848

864 **Step 4: Restricted eigenvalue and final bound.** The cone constraint (Hastie et al., 2015) allows  
 865 us to control the estimation error using the restricted eigenvalue (RE) condition (Raskutti et al.,  
 866 2010). This condition requires that the design matrix  $\Psi$  has good properties when restricted to  
 867 sparse vectors:

$$\frac{1}{n} \|\Psi \Delta\|_2^2 \geq \kappa \|\Delta\|_2^2$$

870 for all  $\Delta$  satisfying the cone constraint.

871 The RE condition is natural for motif-aligned representations: it says that different motifs produce  
 872 sufficiently different representation patterns that they can be distinguished statistically.

873 Using the Cauchy-Schwarz inequality  $\|\Delta_S\|_1 \leq \sqrt{k} \|\Delta\|_2$  and combining with our earlier bound:

$$\frac{\kappa}{2} \|\Delta\|_2^2 \leq \frac{1}{2n} \|\Psi \Delta\|_2^2 \leq \frac{3\lambda}{2} \sqrt{k} \|\Delta\|_2$$

874 Solving:  $\|\Delta\|_2 \leq \frac{3\sqrt{k}}{\kappa} \lambda$ .

875 For the prediction error:

$$\frac{1}{n} \|\Psi(\hat{w} - w^*)\|_2^2 \leq \frac{9k}{\kappa} \lambda^2$$

876 **Sample complexity conclusion.** With  $\lambda = C\sigma\sqrt{\frac{\log p}{n}}$ , achieving prediction error at most  $\varepsilon^2$  re-  
 877 quires:

$$\frac{9k}{\kappa} \cdot C^2 \sigma^2 \frac{\log p}{n} \leq \varepsilon^2$$

878 Solving for  $n$ :

$$n \geq \frac{9C^2 \sigma^2 k \log p}{\kappa \varepsilon^2} = \tilde{\mathcal{O}}\left(\frac{\sigma^2}{\kappa} \cdot \frac{k \log p}{\varepsilon^2}\right).$$

879 This improves upon the standard dense regression bound of  $\tilde{\mathcal{O}}(p/\varepsilon^2)$  by a factor of  $p/(k \log p)$ .  
 880 When motifs are sparse ( $k \ll p$ ), this represents an exponential improvement in sample complexity.

□

### 897 A.3 DATA

898 We provide a short overview of the dataset used in this study.

#### 901 A.3.1 MOLECULENET

902 We use MoleculeNet Wu et al. (2018) as one of our benchmarks. All of the benchmarks are used  
 903 with scaffold splitting. The benchmark contains the following datasets:

904 **BACE** BACE contains approximately 1.5k molecules and their bioactivity measurement for in-  
 905 hibition of human  $\beta$ -secretase 1 (BACE-1). The bioactivity values are an aggregate of scientific  
 906 literature and not from a single bioassay.

907 **BBBP** The blood-brain barrier penetration dataset contains approximately 2k molecules, and its  
 908 activity is determined by whether it is able to pass the highly selective membrane and enter the brain  
 909 fluid.

910 **ClinTox** The clinical toxicity (ClinTox) contains two bioactivity prediction tasks: (1) FDA ap-  
 911 proved and (2) failure of clinical trials. The dataset contains approximately 58k molecules.

912 **HIV** The HIV dataset contains approximately 40k of molecules and measures the evidence of  
 913 anti-HIV activity.

**SIDER** The side effect resources (SIDER) dataset contains approximately 1.4k molecules spanning 27 assays measuring the side effects of drugs.

**Tox21** The Tox21 dataset measures the drug-related effects spanning 12 different prediction tasks with over 7.8k molecules.

**ToxCast** The ToxCast dataset provides 617 classification tasks based on in vitro drug screening. The dataset contains 8.5 molecules.

**MUV** The maximum unbiased validation (MUV) dataset spans 17 tasks designed to identify active compounds. The dataset contains approximately 93k molecules.

**Lipo** The lipophilicity dataset contains hydrophobicity measurements of 4.2k molecules.

**ESOL** The Delaney Solubility Dataset contains water solubility measurements for over 1.1k of molecules.

**FreeSolv** The Freesolv dataset contains the measurements for hydration free energy for small molecules and contains 624 molecules.

### A.3.2 PHOTOSWITCH

For additional regression tasks, we use the photoswitch dataset (Griffiths et al., 2022), where we use the datasets that contain more than 100 molecules, and we again scaffold-split the datasets.

**CAM** The CAM-B3LYP benchmark contains 117 molecules and computed electronic transition wavelengths in nm.

**PBE0** The PBE0 dataset contains 114 molecules and computed electronic transition wavelengths.

**E and Z isomer** These datasets contain the wavelengths of transitions between different electronic states ( $n, \pi, \pi^*$ ) that have been observed for the different isomers.

### A.4 TEMPLATE REPHRASES

List of rephrased templates for functional groups used in Section 6.2. The  $\langle \text{GROUP} \rangle$  parameters are replaced with the name of the functional group:

- “is the  $\langle \text{GROUP} \rangle$  group present”
- “does it have a  $\langle \text{GROUP} \rangle$  group”
- “is there a  $\langle \text{GROUP} \rangle$  group in it”
- “does this structure include a  $\langle \text{GROUP} \rangle$  group”
- “is a  $\langle \text{GROUP} \rangle$  group part of the molecule”
- “does the compound contain a  $\langle \text{GROUP} \rangle$  group”
- “can a  $\langle \text{GROUP} \rangle$  group be found here”
- “is the  $\langle \text{GROUP} \rangle$  functional group present”
- “does the molecule feature a  $\langle \text{GROUP} \rangle$  group”
- “is there evidence of a  $\langle \text{GROUP} \rangle$  functional group”
- “does this molecule exhibit a  $\langle \text{GROUP} \rangle$  group”
- “is a  $\langle \text{GROUP} \rangle$  functional group detectable”
- “does the structure show the presence of  $\langle \text{GROUP} \rangle$ ”
- “can a  $\langle \text{GROUP} \rangle$  group be identified here”
- “is  $\langle \text{GROUP} \rangle$  part of the chemical composition”

- 972           • “does the sample possess a  $\langle \text{GROUP} \rangle$  group”  
 973           • “is there a  $\langle \text{GROUP} \rangle$  moiety in this compound”  
 974           • “does this substance carry a  $\langle \text{GROUP} \rangle$  group”  
 975           • “can the molecule be classified as containing a  $\langle \text{GROUP} \rangle$  group”  
 976           • “is the  $\langle \text{GROUP} \rangle$  function observed in this case”  
 977  
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979           A.5 TRAINING PARAMETERS  
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982           Table 3: **Training hyperparameters.** Hyperparameter setting used to train our model.  
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Hyperparameter	Value
Batch size	76
GPUs	6 x NVIDIA H100
GPUh	252h
Alternating loss steps	20
Precision	float16
Hidden size	768
Maximum of positional embeddings	1024
Number of hidden layers	22
Learning rate	0.01
Warmup steps	10000
Optimizer	AdaFactor (Shazeer & Stern, 2018)

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## A.6 ATTENTION

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In Section A.6, we show an example of how attention maps the property value token to the description and the relevant atoms, in this case, that is Fluorine (F). Additionally, we show that the atom itself attends to a phrase “contains halogen” as well as the property value.

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In Section A.6, we show the average attention per SMILES token across all attention heads for the second-to-last layer. The results are averaged over 5000 molecules that contain a halogen group, where we fix the task description as shown in Section A.6.

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Figure 5: **Attention heads in the second to last layer exhibit the ability to correlate the task to prediction and corresponding chemical element.** Top, the source token for correct prediction is attended by the task description and all Fluorine (F) atoms. Bottom, the Fluorine atom receives attention from value tokens as well as the phrase “contains halogen group.” Illustration created using BertViz (Vig, 2019).

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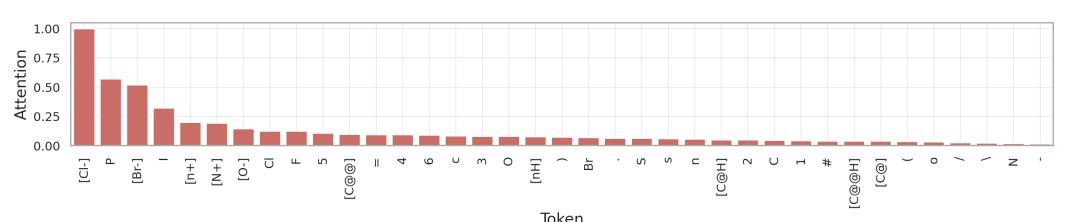
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Figure 6: **Average attention per SMILES token across all attention heads for the second-to-last layer for molecules containing a halogen group.** The task description is fixed as shown in A.6 and the experiment contains 5000 molecules that in turn contain the halogen group.

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## A.7 PER FUNCTIONAL GROUP EMBEDDINGS

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Here we show a full embedding breakdown per functional group. The molecules are from the test set that has been scaffold split against the training set. As shown in the Fig. A.7 CILTs embeddings cluster for each of the groups (except thiol) into clusters based on the functional group.

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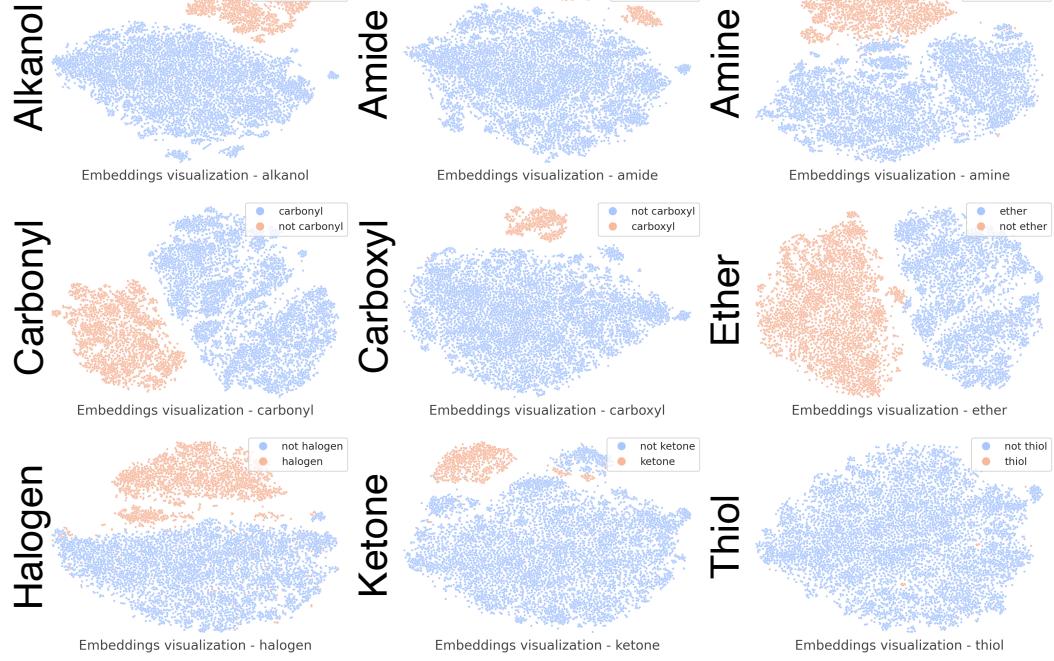


Figure 7: **Functional group embeddings breakdown.** The task description is fixed for each of the functional groups. The model is in prediction mode, where the value of the functional group is masked and the molecule is shown in full. Molecules are from the test set that is scaffold split against the train set.

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## A.8 CILT SCALING RESULTS BREAKDOWN

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Here we expand on the section 6.4 results for all of the version of CILT Models we have trained that are not shown in the main text. We conduct the same linear probe experiments as in the previously mentioned section and give a breakdown based on the number of molecules in the pre-training dataset.

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**Table 4: Linear probe breakdown for different versions of CILT** Logistic regression and linear regression trained on embeddings over 4-fold cross-validation scaffold split. For classification we report %AUCROC ( $\uparrow$ ) and for regression MAE ( $\downarrow$ ).

Model	Classification (%AUCROC $\uparrow$ )								
	BACE	BBBP	ClinTox	HIV	SIDER	Tox21	ToxCast	MUV	Avg.
75k	78.9 $\pm$ 4.0	94.3 $\pm$ 1.3	98.1 $\pm$ 1.6	74.9 $\pm$ 2.0	57.3 $\pm$ 6.4	72.5 $\pm$ 6.3	68.4 $\pm$ 11.0	73.8 $\pm$ 13.0	77.3
125k	79.0 $\pm$ 3.3	94.4 $\pm$ 0.8	98.3 $\pm$ 0.1	75.3 $\pm$ 1.1	58.0 $\pm$ 6.7	72.7 $\pm$ 6.4	68.1 $\pm$ 11.2	72.5 $\pm$ 13.4	77.3
250k	81.3 $\pm$ 2.5	94.5 $\pm$ 1.3	98.3 $\pm$ 0.1	75.6 $\pm$ 0.7	58.5 $\pm$ 6.8	72.5 $\pm$ 6.0	68.0 $\pm$ 11.2	75.2 $\pm$ 12.3	78.0
375k	78.9 $\pm$ 3.8	94.4 $\pm$ 0.7	98.7 $\pm$ 1.1	75.7 $\pm$ 1.0	58.8 $\pm$ 5.8	72.3 $\pm$ 6.0	68.6 $\pm$ 11.0	74.2 $\pm$ 11.6	77.7
500k	80.4 $\pm$ 1.2	92.5 $\pm$ 1.2	97.7 $\pm$ 1.5	73.9 $\pm$ 1.5	55.2 $\pm$ 6.3	66.3 $\pm$ 6.9	64.4 $\pm$ 10.3	71.9 $\pm$ 13.7	75.3

Model	Regression (MAE $\downarrow$ )								
	Lipo	ESOL	FreeSolv	CAM	PBE0	En $- \pi^*$	E $\pi$ $- \pi^*$	Zn $- \pi^*$	Rank
75k	0.81 $\pm$ 0.02	0.91 $\pm$ 0.30	0.90 $\pm$ 0.18	42.0 $\pm$ 12.6	66.5 $\pm$ 7.4	27.6 $\pm$ 11.9	51.3 $\pm$ 8.0	14.0 $\pm$ 5.2	3
125k	0.81 $\pm$ 0.02	0.91 $\pm$ 0.30	0.90 $\pm$ 0.18	38.7 $\pm$ 13.9	67.7 $\pm$ 7.9	27.5 $\pm$ 12.0	51.3 $\pm$ 7.3	14.0 $\pm$ 5.2	3
250k	0.81 $\pm$ 0.02	0.90 $\pm$ 0.18	0.91 $\pm$ 0.30	49.1 $\pm$ 19.1	65.8 $\pm$ 7.0	27.5 $\pm$ 12.0	51.3 $\pm$ 7.3	13.9 $\pm$ 5.2	4
375k	0.81 $\pm$ 0.02	0.90 $\pm$ 0.18	0.91 $\pm$ 0.30	43.6 $\pm$ 11.9	66.4 $\pm$ 6.8	27.6 $\pm$ 12.0	51.3 $\pm$ 7.3	13.9 $\pm$ 5.2	3
500k	0.80 $\pm$ 0.02	0.88 $\pm$ 0.18	0.91 $\pm$ 0.30	46.9 $\pm$ 15.5	58.5 $\pm$ 7.6	27.5 $\pm$ 12.0	51.3 $\pm$ 7.3	13.9 $\pm$ 5.2	2.5

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1188 A.9 USE OF LLMs  
11891190 Large language models were employed as assistive tools for tasks including text rewriting,  
1191 spellchecking, minor stylistic improvements, and the writing of this statement. All content was  
1192 reviewed and verified by the authors, who take full responsibility for the final manuscript. LLMs  
1193 did not contribute to research ideation or substantive writing decisions.

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