How Low Can You Go? Identifying Prototypical In-Distribution Samples for Unsupervised Anomaly Detection

Anonymous authors
Paper under double-blind review

Abstract

Unsupervised anomaly detection (UAD) alleviates large labeling efforts by training exclusively on unlabeled in-distribution data and detecting outliers as anomalies. Generally, the assumption prevails that large training datasets allow the training of higher-performing UAD models. However, in this work, we show that UAD with extremely few training samples can already match – and in some cases even surpass – the performance of training with the whole training dataset. Building upon this finding, we propose an unsupervised method to reliably identify prototypical samples to further boost UAD performance. We demonstrate the utility of our method on seven different established UAD benchmarks from computer vision, industrial defect detection, and medicine. With just 25 selected samples, we even exceed the performance of full training in 25/67 categories in these benchmarks. Additionally, we show that the prototypical in-distribution samples identified by our proposed method generalize well across models and datasets and that observing their sample selection criteria allows for a successful manual selection of small subsets of high-performing samples. Our code is available at https://anonymous.4open.science/r/uad_prototypical_samples/

1 Introduction

Unsupervised anomaly detection (UAD) or out-of-distribution (OOD) detection aims to distinguish samples from an in-distribution (ID) from any sample that stems from another distribution. To address this task, typically, machine-learning models are employed to represent the in-distribution by exclusively using samples from that distribution for training. The converged model detects OOD samples via their distance to the in-distribution. Compared to supervised training, this setup alleviates the need for large labeled datasets, is not susceptible to class imbalance, and is not restricted to anomalies seen during training. Due to these advantages, UAD has several vital applications in computer vision: It is used to detect pathological samples in medical images (Schlegl et al., 2019; Lagogiannis et al., 2023; Bercea et al., 2022; Meissen et al., 2023), to spot defects in industrial manufacturing (Bergmann et al., 2019; Roth et al., 2022; Deng & Li, 2022; Bae et al., 2023), or as safeguards to filter unsuitable input data for supervised downstream models, for example, in autonomous driving.

In deep learning, the prevailing assumption is that more data leads to better models. However, training with only very few samples would, have numerous advantages. Small datasets are cheap, easy to obtain, and also available for a wider variety of tasks. Additionally, it also makes more tasks feasible to solve where data may be very difficult or expensive to acquire. Moreover, small training datasets lead to better explainability since output scores can directly be related to (dis-)similarities to the training data. This improved explainability, in turn, lowers the entry bar for designing performant algorithms, especially for actors with few resources, thus contributing to the democratization of AI. While overfitting with very few training samples diminishes the utility of supervised machine learning algorithms, UAD models are not impacted in the same way. In fact, they rely on overfitting to the ID data and, as a result, not generalizing to OOD data. This makes UAD suitable for training with extremely small datasets.

In this paper, we present findings showing that only very few training samples are required to achieve similar or even better anomaly detection performance compared to training with 100% of the available training data

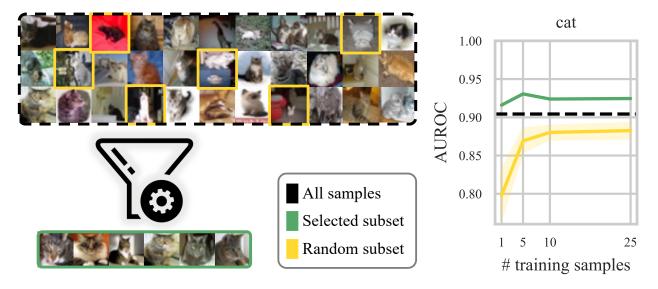


Figure 1: Selecting only a few prototypical in-distribution samples (identified by our method) for training can result in higher anomaly detection performance than training with 100% of the available data. Results for anomaly detection on the *cat* class from CIFAR10. Black dashed: full training. Yellow: randomly selected samples, including standard deviations over different random selections. Green: Best-performing samples identified with our method.

(which we denote as "full training" in the remainder of the manuscript). Through the unification of concepts from anomaly detection and core-set selection, we additionally propose an unsupervised method for selecting a high-performing subset of samples from the initial training dataset, as depicted in Figure 1, and evaluate the effectiveness of this approach on a multitude of different models, datasets, and tasks. We further propose two weakly supervised selection strategies that serve as weak upper bounds for the introduced unsupervised sample selection and provide additional insights into the method's selection process. In summary, the main contributions of this paper are:

- We show for the first time that an exceedingly small number of training samples can suffice for performant, robust, and interpretable UAD, achieving state-of-the-art performance on a multitude of established benchmarks.
- We propose an unsupervised method to reliably find well-performing subsets prototypical indistribution samples and describe their common characteristics.
- We further demonstrate that the prototypical samples identified by our method and their characteristics translate to equally good performance for other models, datasets, and even tasks.
- Lastly, we provide a theoretical justification explaining the increase in performance through training with very few samples.

2 Related Work

Our work combines ideas from both the fields of anomaly detection and core-set selection. Here, we give a brief overview of these concepts and related research work.

2.1 Anomaly Detection

Anomaly detection is deeply rooted in computer vision, with many influential works benchmarking their models on natural-image datasets, such as CIFAR10 and CIFAR100 (Krizhevsky et al., 2009), MNIST

(LeCun et al., 1998), or Fashion-MNIST (Xiao et al., 2017). Early works attempting to solve UAD on these benchmarks were mostly based on (variational) autoencoders (Zhou & Paffenroth, 2017; Kim et al., 2019; Liu et al., 2020; Abati et al., 2019) or GANs (Perera et al., 2019; Deecke et al., 2019; Akcay et al., 2019) trying to restrict the learned manifold of the generative model. The model is expected to faithfully reconstruct in-distribution samples, whereas OOD samples can be detected due to their large reconstruction errors. Also, one-class classification models (Ruff et al., 2018) or ones that learn surrogate tasks (Golan & El-Yaniv, 2018; Bergman & Hoshen, 2020) have been successfully used. More recently, works based on pre-trained neural networks (such as ResNets He et al. (2016)) have become popular and still are the best-performing models for the aforementioned datasets (Bergman et al., 2020).

The release of MVTec-AD (Bergmann et al., 2019) for industrial defect detection sparked several works focusing on this dataset as it was the first to contain a variety of useful, real-world anomaly detection tasks. After early attempts to solve these with various techniques, including autoencoders (Bergmann et al., 2018) and knowledge distillation methods (Bergmann et al., 2020), research converged on self-supervised approaches (Zavrtanik et al., 2021; Li et al., 2021), ResNets pre-trained on ImageNet (Defard et al., 2021; Deng & Li, 2022; Roth et al., 2022), or combinations thereof (Bae et al., 2023).

Anomaly detection was also successfully applied in medical computer vision, where it is used to discriminate samples from healthy subjects (in-distribution) from diseased ones (outliers). Schlegl et al. (2019) have successfully discovered biomarkers in retinal OCT images using a GAN. To detect tumors and lesions in brain MRI, numerous autoencoder-based approaches (You et al., 2019; Baur et al., 2021; Zimmerer et al., 2019) and diffusion models (Wyatt et al., 2022) have been proposed. Furthermore, anomaly detection has been successfully applied in chest X-ray images to detect COVID-19 (Zhang et al., 2020) or other malignancies (Lagogiannis et al., 2023; Mao et al., 2020).

However, without exception, the existing works have followed the established paradigm of using the largest available training dataset, a notion we aim to challenge in this study.

2.2 Core-set Selection

To this end, we utilize methods from the field of core-set selection. Core-set selection aims to create a small informative dataset such that the models trained on it show a similar test performance compared to those trained on the original dataset. Core-set selection techniques for deep learning include minimizing the feature-space distance (Welling, 2009) or the distance of gradients with respect to a neural network (Mirzasoleiman et al., 2020) between the selected subset and the original dataset. In anomaly detection, core-set selection has been used by Roth et al. (2022). Here, however, the selection is done on patch features instead of images. In MemAE, Gong et al. (2019) restricted the latent space of an autoencoder to a set of learned in-distribution feature vectors to perform anomaly detection. While this work also finds prototypical feature vectors, they again cannot be linked back to training samples and, consequently, cannot be used for core-set selection.

3 Surfacing Prototypical In-Distribution Samples Through Core-Set Selection

In unsupervised anomaly detection, the task is to train a model that can determine the binary label $y \in Y$ (ID: y = -1 or OOD: y = 1) of a sample $x \in X$. In this setting, it is commonly assumed that the training dataset X_{train} contains only ID samples ($y = -1, \forall x \in X_{\text{train}}$), alleviating large labeling efforts for anomaly detection models. Without loss of generality, a typical neural network used for anomaly detection can be divided into two parts: The feature-extractor ψ transforms a sample x to its latent representation $z \in Z$, and a predictor ϕ that computes the anomaly score s from s. The anomaly score $s \in \mathbb{R}$ is a potentially unbounded continuous value that represents the "outlierness" of a sample s. Combined, the anomaly detection model s0 computes the anomaly score of a sample:

$$\theta(x) = \phi(\psi(x)) = s. \tag{1}$$

Such anomaly detection models are usually trained on large datasets. When performing core-set selection, we want to determine a subset X_{sub} with $M \in \mathbb{N}$ samples from the original training dataset X_{train} with $N \in \mathbb{N}$ samples denoted as $x_i \in \mathbb{R}^D$ with i = 1, 2, ..., N, such that:

minimize
$$E(X_{\text{sub}}, \theta)$$
 subject to
$$X_{\text{sub}} \subset X_{\text{train}},$$

$$|X_{\text{sub}}| = M,$$
 (2)

where $E(X_{\text{sub}}, \theta)$ is the detection error produced by a model θ trained on X_{sub} .

Previous work by Defard et al. (2021) has shown that the latent space Z_{train} represents a semantically meaningful compression of the training distribution X_{train} that can be modeled as a multivariate Gaussian. Due to the limited representational power of unimodal Gaussian distributions, we extend this idea by instead fitting a Gaussian Mixture Model (GMM) with M components to this latent space. Let $\mathcal{G} = \{\mu_1, \mu_2, \dots, \mu_M\}$ represent the set of means (centroids) of the M components of the GMM, where μ_j denotes the mean of the j-th component. This allows us to choose the samples corresponding to the latent codes that are closest to the centroids as core-set samples.

$$X_{\text{sub}} = \left\{ x_i \mid \forall \mu_j \in \mathcal{G}, \underset{x_i \in X_{\text{train}}}{\text{arg min}} \mid |\psi(x_i) - \mu_j||_2 \right\}. \tag{3}$$

The use of a GMM ensures that multiple different modes of normality are represented in X_{sub} .

3.1 Weakly-Supervised Baselines

In addition to the unsupervised core-set selection, we are interested in finding the optimal training subsets when labeled data is available. Finding these subsets would offer additional insights into "normality" in anomaly detection and enable us to put the performance achieved by our core-set selection strategy into perspective, establishing an upper bound. However, the problem is \mathcal{NP} -hard. For a subset size of M, there exist $C(N,M) = \binom{N}{M} = \frac{N!}{M!(N-M)!}$ possibilities. We therefore propose two approximate solutions to this problem, where we make use of the following simplification: By evaluating the detection error obtained from training with individual samples, we aim to identify small yet high-performing training datasets.

3.1.1 Greedy Selection

Since it is possible to train UAD models with only one sample, we can heuristically estimate the quality of each sample individually as $E(\{x_i\}, \theta, X_{\text{val}})$ for i = 1, 2, ..., N. From this information, we construct X_{sub} as:

$$X_{\text{sub}} = \underset{\{x_j | x_j \in X_{\text{train}}, \ 1 \le j \le M\}}{\arg \min} \sum_{j=1}^{M} E(\{x_j\}, \theta, X_{\text{val}}). \tag{4}$$

In our work, we select the AUROC as the optimization target E. Note that the set of samples that produce the smallest errors is not necessarily equal to the set of samples that together produce the lowest error.

3.1.2 Evolutionary Algorithm

While the greedy approach above is intuitive, fast, and easy to implement, it prefers subsets of visually similar samples, as we will later show. This is desirable in some cases; however, there are also scenarios in which multiple modes of normality should be covered by the selected subset. To get a better coverage of the normal variations in a dataset, we propose a second approach, as described in the following. For each combination of a training sample $x_i \in X_{\text{train}}$ and a validation sample $x_k \in X_{\text{val}}$, we compute an anomaly score $s(x_i, x_k)$ by training the anomaly detection model on x_i only and running inference on x_k . The objective is to find a subset $X_{\text{sub}} = \{x_i | x_i \in X_{\text{train}}, 1 \le i \le M\}$ that maximizes a fitness function f described as:

$$f(X_{\text{sub}}) = \sum_{x_k, y_k \in X_{\text{val}}} \max_{x_i \in X_{\text{sub}}} y_k \cdot s(x_i, x_k).$$
 (5)

Maximizing f allows for finding M training samples x_i that achieve the best performance in classifying validation samples x_k as ID or OOD. Since this problem is also \mathcal{NP} -hard, we approximate the solution using an evolutionary algorithm:

Algorithm 1: Evolutionary Algorithm

Data: Training dataset X_{train} , validation dataset X_{val} , population size P, anomaly scores

 $s(x_i, x_k) \, \forall x_i \in X_{\text{train}}, x_k \in X_{\text{val}}, \text{ fitness function } f, \text{ number of generations } G$

Result: Approximately optimal subset X_{sub}^*

Initialize a random population $\mathcal{P} = \{X_{\sup,p} | 1 \leq p \leq P\};$

for $qen \leftarrow 1$ to G do

Evaluate the fitness function f for each individual $X_{\sup,p} \in \mathcal{P}$;

Remove the least-fit $\frac{P}{2}$ individuals $X_{\text{sub},p} \in \mathcal{P}$ from \mathcal{P} to determine the best subset \mathcal{P}' ; Randomly apply either a crossover (combine two individuals) or mutation (replace one sample)

operation to each individual in \mathcal{P}' to create a modified population \mathcal{P}'' ;

Generate a new population $\mathcal{P} = \mathcal{P}' \cup \mathcal{P}''$;

end

return Best individual found in the final population;

In the crossover operation, random subsets of two individuals $X_{\text{sub},1}$, $X_{\text{sub},2} \in \mathcal{P}'$ (called parents) are merged to produce a new individual X'_{sub} , such that $|X'_{\text{sub}}| = M$. In the mutation operation, one sample $x_1 \in X_{\text{sub}}$ of an individual $X_{\text{sub}} \in \mathcal{P}'$ is randomly replaced with another sample $x_2 \in D \setminus X_{\text{sub}}$ to produce a new individual X'_{sub} :

$$X'_{\text{sub}} = X_{\text{sub}} \setminus \{x_1\} \cup \{x_2\}. \tag{6}$$

In contrast to the greedy selection strategy that favors visually similar samples, the subsets found by the evolutionary algorithm have better coverage of the different notions of normality contained in the training dataset (c.f. Figure 8). However, note that this enhanced coverage could also be harmful when the normal dataset is noisy and contains samples that should be considered abnormal. In such cases, greedy selection is more effective at filtering out these samples.

Experiments

4.1 **Datasets and Models**

To evaluate our methods, we use datasets from the natural- and medical-image domains, showing the applicability of our method in diverse tasks. CIFAR10, CIFAR100 (Krizhevsky et al., 2009), MNIST (LeCun et al., 1998), and Fashion-MNIST (Xiao et al., 2017) are trained in a one-vs-rest setting, where one class is used as the in-distribution, and all other classes are combined as outliers. MVTec-AD (Bergmann et al., 2019) is an industrial defect detection dataset and a frequently used benchmark for UAD models. In the chest X-ray images of the RSNA Pneumonia Detection dataset (Stein et al., 2018), the in-distribution constitutes images of healthy patients, while anomalous samples show signs of pneumonia or other lung opacities. In addition, we use CheXpert (Irvin et al., 2019) to test if the samples found by our method generalize to other datasets. Similarly to RSNA, the in-distribution samples here are images labeled with "No Finding". while OOD samples either display pneumonia or other lung opacities. Lastly, we detect MRI slices with glioma in the BraTS dataset (Menze et al., 2014; Bakas et al., 2017). For each dataset, we chose a respective state-of-the-art model: PANDA (Reiss et al., 2021) is used for CIFAR10, CIFAR100, MNIST, and Fashion-MNIST, PatchCore (Roth et al., 2022) for MVTec-AD, and FAE (Meissen et al., 2023; Lagogiannis et al.,

Table 1: AUROC scores of training with the full dataset and training with 1, 5, 10, and 25 best-performing (with greedy search, the evolutionary algorithm, and core-set selection) or random samples. Since the performance of randomly selected subgroups can vary strongly, we repeated these experiments over ten different subsets. Bold numbers show the best performance per dataset, and underlined numbers are the best per sample size. Numbers marked with * surpass training with 100% of the data. This table shows the averaged results over each class in the respective datasets. Detailed results for all classes can be found in the Appendix.

	Method	CIFAR10	CIFAR100	F-MNIST	MNIST	MVTec-AD	BraTS	RSNA
le	Random	84.96 ±1.2	77.05 ± 1.4	83.51 ± 3.3	76.11 ± 2.2	83.61 ± 1.2	93.85 ± 5.3	61.86 ± 5.2
sample	Greedy	95.02	89.94	94.00	90.15	89.70	95.25	70.64
Sa	Evo	87.68	79.99	86.82	74.82	85.55	94.00	65.62
	Core-set	90.08	79.62	91.47	82.29	82.80	96.25	67.36
les	Random	91.95 ± 0.7	86.40 ±0.8	91.86 ± 1.1	89.36 ±1.1	90.43 ±0.9	97.83 ± 1.1	73.14 ± 3.5
samples	Greedy	96.36	92.49	95.31	93.62	91.52	97.25	74.19
sar	Evo	94.30	90.40	93.42	92.96	94.12	<u>99.06</u> *	76.45
ಸ್ತ	Core-set	93.85	89.74	93.89	92.55	92.13	96.88	75.06
les	Random	93.59 ± 0.4	89.05 ± 0.5	93.24 ± 0.3	93.00 ± 0.6	92.80 ± 0.6	97.96 ± 1.0	75.43 ± 3.0
samples	Greedy	96.37	92.59	95.38	94.91	92.94	97.06	77.80
sar	Evo	95.51	91.78	94.28	95.37	96.37	98.81*	76.61
10	Core-set	94.28	91.03	94.47	$\overline{94.94}$	96.77	98.19	76.78
les	Random	94.68 ± 0.2	91.52 ± 0.2	94.16 ± 0.2	96.24 ± 0.3	95.29 ± 0.3	98.09 ± 0.6	77.08 ± 0.9
samples	Greedy	96.29	92.60	95.52	96.06	93.53	97.94	78.60*
sar	Evo	95.51	91.88	95.07	97.30	98.52^{*}	98.87*	80.59*
25	Core-set	94.85	92.88	95.13	97.33	98.37	99.12*	79.18*
	Full training	96.58	94.92	95.79	98.41	98.48	98.75	77.97

2023) for RSNA, CheXpert and BraTS. We additionally used Reverse Distillation (RD) by Deng & Li (2022) for RSNA to test the generalizability of identified samples across models. Details about the datasets and models can be found in the supplementary material.

4.2 Experimental Setup

We carefully tune all baselines and methods using established protocols where applicable. Details are in Appendix C. We restricted the maximum subset size to M=25 samples in our experiments as we did not experience substantial increases in performance beyond this point. Multiplied by the number of experiments and selection strategies, this decision saved significant amounts of our limited resources and allowed for more extensive experimentation in other dimensions.

5 Results and Discussion

In the following, we first present our main findings in Section 5.1, Section 5.2, and Section 5.3. Then, we provide theoretical justification for these findings (Sections 5.4 and 5.5) and dive deeper into the specific characteristics of the different datasets and how they impact the core-set selection performance in Section 5.6.

5.1 A Few Selected Samples Can Outperform Training With the Whole Dataset

As shown in Table 1, UAD models achieve high performance even when trained with only a few samples.

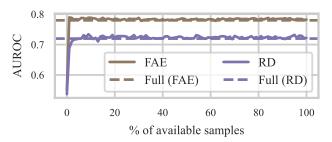


Figure 2: Performance in RSNA does not increase with more training samples.

Surprisingly, even randomly selected subsets can result in a strong model on all considered datasets, confirming our hypothesis that UAD models do not overfit. Figure 2 depicts a typical observation that detection performance saturates already with very few samples, regardless of the UAD model used. However, not all random subsets perform equally well, which can be seen by the large standard deviations (up to 15.1) for many categories (see Table 2 in Appendix D). Our proposed core-set selection strategy substantially outperforms random selection on all datasets and even performs better than full train-

ing on BraTS and RSNA. Notably, for the latter, it uses as little as 0.3% of the available training data. In addition to this, the evolutionary algorithm and especially the greedy selection strategy also demonstrate good performance. Moreover, our core-set selection strategy is not far behind and outperforms the other two with 25 samples on CIFAR100 and MNIST despite not using any labels. Overall, our selection strategies outperform full training in 25/67 categories tested in this study (see Tables 2 to 6 in Appendix D). Furthermore, the gap between random and informed selection becomes even more pronounced in the very-low data regime (1–10 samples).

5.2 What Characterizes Normal Samples?

Our method not only allows training strong UAD models with only a few samples, but it also provides insights into what constitutes a prototypical in-distribution image. Figure 3 shows the best- and worst-performing samples for each class in CIFAR10 on the left. The "best" images display well-lit prototypical objects that are well-centered, have good contrast, and have mostly uniform backgrounds. In contrast, the "worst" indistribution images include drawings (bird and horse), toys (frog, truck), historical objects (plane, car), or images with bad contrast (dog). In noisy, less well-curated datasets like RSNA, our method can effectively detect and filter low-quality samples. Figure 3 reveals severe deformations, foreign objects such as access tubes or implants, low tissue contrast, or dislocations in the worst-performing samples. The best-performing ones, on the other side, are well-centered and detailed, contain male and female samples, and clearly show the lungs, a prerequisite for detecting pneumonia.



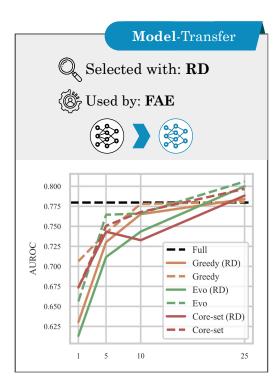
Figure 3: Best- and worst-performing samples in CIFAR10 and RSNA. Identified using our proposed core-set selection strategy.

Motivated by the insights we gained about the characteristics of in-distribution samples, we manually selected a training subset. We chose the RSNA dataset for this experiment because, unlike MVTec-AD, it contains atypical samples in the "normal" data and because the images are large enough to be visually inspected, in contrast to the other natural image datasets. We selected samples that had similar characteristics as displayed in Figure 3 and covered the distribution of ID samples well. We only started the evaluation of the

manually selected samples once their selection was complete. No information other than the characteristics described above was used for the manual selection, and the author selecting the samples is not a trained radiologist or other medical expert. When training with these manually selected samples, we achieved AUROCs of 67.88, 76.61, 79.73, and 81.04 for 1, 5, 10, and 25 samples, respectively. The manual selection strategy, therefore, outperformed all automatic selection strategies and even full training, giving the best results on RSNA in this work.

We repeated a similar experiment for the – arbitrarily selected – "8" class on MNIST. This time, however, we did not look at the best- or worst-performing samples from this class but simply selected samples that are visually close to a prototypical 8. We discarded samples that had non-closed lines, where the curves of the 8 were excessively slim, and those with irregular or wavy lines. With these samples, we achieved AUROCs of **76.69**, **92.42**, **94.59**, and **96.37** for 1, 5, 10, and 25 samples, respectively, outperforming both random selection and the evolutionary strategy, while also only falling 1.45 points below full training performance.

5.3 Prototypical Samples Are Transferrable to Other Models and Datasets



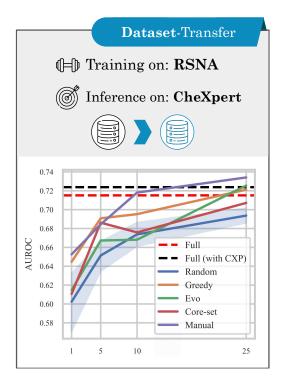


Figure 4: Prototypical samples transfer well to other datasets and models. **Left**: Surfaced samples with the RD model achieve high performance when used with FAE. Test performance of FAE on RSNA when samples are selected using RD (full lines) or FAE (dashed lines). **Right**: Training with 25 carefully selected samples from RSNA can exceed full training performance with CheXpert (8443 samples) when evaluated on the latter. Test performance on CheXpert after training on CheXpert samples (black, dashed line) or RSNA (other lines).

We also trained a second type of UAD model, Reverse Distillation (RD), on RSNA. This model matches encoder and decoder representations at different levels. The left side of Figure 4 shows how the best-performing samples selected with our proposed core-set selection and the two weakly-supervised baselines on RD perform when applied to the FAE model. Although RD generally performs worse than FAE, its best-performing set of samples works well on FAE, performing almost on par with the ones found with FAE itself and even exceeding full performance. We conclude from this result that there are commonalities between the best samples that are independent of the model. This means that samples found using one model can be transferred to another.

Similarly, high performance for sample combinations on RSNA translates well to the CheXpert dataset. As expected, training with RSNA samples gives a slightly lower performance on CheXpert than training on CheXpert itself (red and black dashed lines in Figure 4, right). This gap, however, can be closed by training with only 25 high-performing samples from RSNA. Even more impressive, we reached higher performance on CheXpert when training with the 25 manually selected RSNA samples than when training on the full CheXpert dataset itself.

5.4 Long-Tail In-Distribution Samples Can Degrade Anomaly Detection Performance

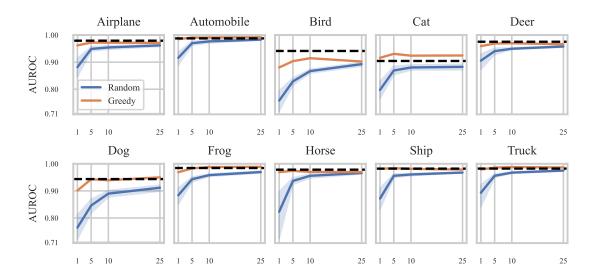
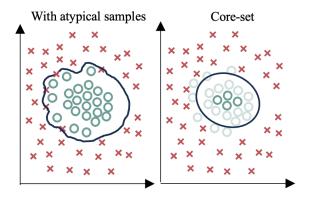


Figure 5: Only 10 representative training samples are needed to surpass the performance of training with the whole dataset on five out of the ten classes in CIFAR10. AUROC for training with 1, 5, 10, and 25 random or best (greedy selection) samples on CIFAR10. For random samples, the experiments were repeated ten times with different samples. The dashed black line represents training with all 4000 ID samples.

In Table 1 and figs. 2 and 4, we have seen that very small datasets can exceed the performance of full training. Figure 5 even shows that peak performance for the "cat" class of CIFAR10 is achieved with only five samples. These results suggest that there exist samples in many datasets whose inclusion degrades performance. We hypothesize that the reason for this is due to the nature of the in-distributions, which often have long tails, as shown by Feldman (2020) and Zhu et al. (2014). The long-tail hypothesis states that the majority of the in-distribution samples have only low inter-sample variance, with the exception of a few rare samples that differ a lot from the rest (while still being part of the in-distribution). While these samples can be actively contrasted to other classes and memorized by supervised machine learning models (Feldman & Zhang, 2020), such mechanisms are not available for UAD models, where the long-tail in-distribution samples are treated as any other training sample. This can shift the decision boundary in an unfortunate way (Figure 6, left). Training with carefully selected samples effectively ignores these data points and can lead to better performance despite using fewer samples (Figure 6, right). Figure 7 reveals that the datasets used in our study also follow a long-tail distribution and that the samples at the tails perform worse. Additionally, the worst-performing samples in Figure 3 are clearly atypical and, thus, likely also lay at the tail of the in-distribution.

5.5 Should Samples From the Long Tails of the In-Distribution Be Considered Outliers?

Our experiments suggest that there are samples in the training datasets that lie at the tails of the indistribution and lower the performance of the UAD model. Our subset-selection strategies are effective at filtering out these data points. Of course, ignoring such long-tail samples during training will declare them as outliers, which, at first sight, is false given the labels. We argue, however, that these data points



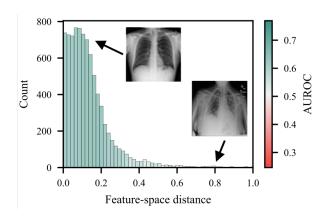


Figure 6: Training with only a few selected samples helps to ignore training samples that lie at the tails of the in-distribution. Illustration of our hypothesis of how long-tail in-distribution samples can skew the decision boundary. **Left**: Some ID samples might be closer to the OOD data and shift the decision boundary. **Right**: Training with a few carefully selected samples creates a better decision boundary.

Figure 7: RSNA follows a long-tail distribution in the FAE feature-space, and the samples at the tails perform worse. Histogram of distances of all training samples from RSNA to the center of an FAE model and two example images from the center and the tail. The bins of the histogram are colored-coded according to the AUROCs of each single sample as described in Section 3.1.1.

should be considered as such because they warrant special consideration in downstream tasks. For example, a subsequent supervised classification algorithm is more likely to misclassify these, and in such a scenario, flagging long tail samples as OOD is desired. Atypical X-ray images, as shown on the bottom right of Figure 3, can pose difficulties (even for manual diagnosis) and should also receive special attention. Further, as we have shown, including these samples during training can lower the classification performance for other samples that are then falsely flagged as ID. The selection methods presented in this study can, therefore, not only be used to identify the most prototypical in-distribution samples but can also be used to automatically filter a dataset from noisy or corrupted images.

5.6 The Distributions of Normality and Abnormality Differ Between Datasets

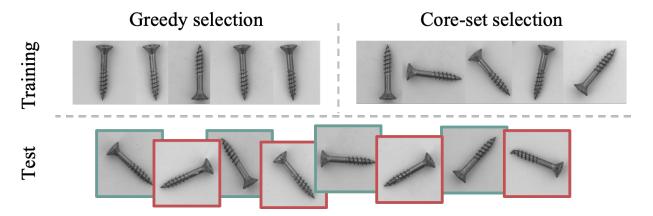


Figure 8: Greedy selection favors visually similar samples, while the core-set selection achieves a better coverage of the normal variations in the training dataset. Best five training samples for the *screw* category in MVTec-AD, found using greedy selection (left) and the unsupervised core-set selection (right). Bottom: normal (green) and defective (red) samples in the test set.

Despite all being used for anomaly detection and showing similar behavior regarding training with few samples, the datasets considered in our study vary greatly with respect to their in and out distribution, as well as the relationships between the two. While for some datasets, the variability between ID samples is comparably low; other datasets contain multiple modes of normality. The objects within each class of the MVTec-AD dataset are very similar and often only oriented differently, and the chest X-ray images all show the same anatomical region. The brain MR images in BraTS are registered to an atlas and, consequently, have even lower anatomical variance. This is in contrast to CIFAR10, CIFAR100, and Fashion-MNIST, where the in-distribution training class can exhibit various shapes, poses, and colors. Similarly, OOD samples can be local and subtle, as in MVTec-AD, BraTS, and RSNA, or global, as in CIFAR10, CIFAR100, MNIST. Fashion-MNIST. A good subset of prototypical samples should cover the different modes of normality but exclude the samples that are atypical and semantically too close to the out-distribution. When looking at examples of the different types of in- and out-distributions described above, we can identify the strengths and weaknesses of the different selection strategies. An example of a dataset with different ID modes and subtle anomalies is the "screw" category of MVTec-AD. Figure 8 shows that greedy selection favors combinations of similarly oriented images and fails to cover the whole space of differently-oriented, normal samples. Our proposed unsupervised core-set selection strategy, on the other hand, (and also the evolutionary algorithm) better covers the different orientations that should all be considered normal (see also Table 5 in Appendix D).

6 Conclusion

In many domains of Deep Learning, the prevailing assumption is that more training data leads to better models. Our work challenges this practice for UAD and highlights the importance of data quality over data quantity. Specifically, we have shown that it is a common phenomenon of UAD models to achieve state-of-the-art performance with extremely few training samples. Our proposed core-set selection strategy is a fast and easy-to-implement, unsupervised approach to automatically extract such high-performing, prototypical training samples. We have further shown that these prototypical in-distribution samples are transferrable across models, datasets, and even modalities and tasks and that the characteristics derived from them allow for manual selection. This understanding of the well-performing training samples can help in designing better UAD models in the future. Our findings further make UAD models much more attractive in practice despite their performance still lacking behind that of their supervised counterparts.

References

- Davide Abati, Angelo Porrello, Simone Calderara, and Rita Cucchiara. Latent space autoregression for novelty detection. In *CVPR*, pp. 481–490, 2019.
- Samet Akcay, Amir Atapour-Abarghouei, and Toby P Breckon. Ganomaly: Semi-supervised anomaly detection via adversarial training. In *ACCV*, pp. 622–637. Springer, 2019.
- Jaehyeok Bae, Jae-Han Lee, and Seyun Kim. Pni: Industrial anomaly detection using position and neighborhood information. In *CVPR*, pp. 6373–6383, 2023.
- Spyridon Bakas, Hamed Akbari, Aristeidis Sotiras, Michel Bilello, Martin Rozycki, Justin S Kirby, John B Freymann, Keyvan Farahani, and Christos Davatzikos. Advancing the cancer genome atlas glioma mri collections with expert segmentation labels and radiomic features. *Scientific data*, 4(1):1–13, 2017.
- Christoph Baur, Stefan Denner, Benedikt Wiestler, Nassir Navab, and Shadi Albarqouni. Autoencoders for unsupervised anomaly segmentation in brain mr images: a comparative study. *Medical Image Analysis*, 69:101952, 2021.
- Cosmin I Bercea, Benedikt Wiestler, Daniel Rueckert, and Shadi Albarqouni. Federated disentangled representation learning for unsupervised brain anomaly detection. *Nature Machine Intelligence*, 4(8):685–695, 2022.
- Liron Bergman and Yedid Hoshen. Classification-based anomaly detection for general data. arXiv preprint arXiv:2005.02359, 2020.

- Liron Bergman, Niv Cohen, and Yedid Hoshen. Deep nearest neighbor anomaly detection. arXiv preprint arXiv:2002.10445, 2020.
- Paul Bergmann, Sindy Löwe, Michael Fauser, David Sattlegger, and Carsten Steger. Improving unsupervised defect segmentation by applying structural similarity to autoencoders. arXiv preprint arXiv:1807.02011, 2018.
- Paul Bergmann, Michael Fauser, David Sattlegger, and Carsten Steger. Mytec ad a comprehensive real-world dataset for unsupervised anomaly detection. In *CVPR*, pp. 9584–9592, 2019. doi: 10.1109/CVPR. 2019.00982.
- Paul Bergmann, Michael Fauser, David Sattlegger, and Carsten Steger. Uninformed students: Student-teacher anomaly detection with discriminative latent embeddings. In CVPR, pp. 4183–4192, 2020.
- Lucas Deecke, Robert Vandermeulen, Lukas Ruff, Stephan Mandt, and Marius Kloft. Image anomaly detection with generative adversarial networks. In *Machine Learning and Knowledge Discovery in Databases: European Conference, ECML PKDD 2018, Dublin, Ireland, September 10–14, 2018, Proceedings, Part I 18*, pp. 3–17. Springer, 2019.
- Thomas Defard, Aleksandr Setkov, Angelique Loesch, and Romaric Audigier. Padim: a patch distribution modeling framework for anomaly detection and localization. In *International Conference on Pattern Recognition*, pp. 475–489. Springer, 2021.
- Hanqiu Deng and Xingyu Li. Anomaly detection via reverse distillation from one-class embedding. In *CVPR*, pp. 9737–9746, 2022.
- Vitaly Feldman. Does learning require memorization? a short tale about a long tail. In *Proceedings of the 52nd Annual ACM SIGACT Symposium on Theory of Computing*, pp. 954–959, 2020.
- Vitaly Feldman and Chiyuan Zhang. What neural networks memorize and why: Discovering the long tail via influence estimation. Advances in Neural Information Processing Systems, 33:2881–2891, 2020.
- Izhak Golan and Ran El-Yaniv. Deep anomaly detection using geometric transformations. Advances in neural information processing systems, 31, 2018.
- Dong Gong, Lingqiao Liu, Vuong Le, Budhaditya Saha, Moussa Reda Mansour, Svetha Venkatesh, and Anton van den Hengel. Memorizing normality to detect anomaly: Memory-augmented deep autoencoder for unsupervised anomaly detection. In *CVPR*, pp. 1705–1714, 2019.
- Kaiming He, Xiangyu Zhang, Shaoqing Ren, and Jian Sun. Deep residual learning for image recognition. In *CVPR*, pp. 770–778, 2016.
- Jeremy Irvin, Pranav Rajpurkar, Michael Ko, Yifan Yu, Silviana Ciurea-Ilcus, Chris Chute, Henrik Marklund, Behzad Haghgoo, Robyn Ball, Katie Shpanskaya, et al. Chexpert: A large chest radiograph dataset with uncertainty labels and expert comparison. In *Proceedings of the AAAI conference on artificial intelligence*, volume 33, pp. 590–597, 2019.
- Ki Hyun Kim, Sangwoo Shim, Yongsub Lim, Jongseob Jeon, Jeongwoo Choi, Byungchan Kim, and Andre S Yoon. Rapp: Novelty detection with reconstruction along projection pathway. In *ICLR*, 2019.
- Alex Krizhevsky, Geoffrey Hinton, et al. Learning multiple layers of features from tiny images. Technical report, Toronto, ON, Canada, 2009.
- Ioannis Lagogiannis, Felix Meissen, Georgios Kaissis, and Daniel Rueckert. Unsupervised pathology detection: A deep dive into the state of the art. *IEEE Transactions on Medical Imaging*, pp. 1–1, 2023. doi: 10.1109/tmi.2023.3298093.
- Yann LeCun, Léon Bottou, Yoshua Bengio, and Patrick Haffner. Gradient-based learning applied to document recognition. *Proceedings of the IEEE*, 86(11):2278–2324, 1998.

- Chun-Liang Li, Kihyuk Sohn, Jinsung Yoon, and Tomas Pfister. Cutpaste: Self-supervised learning for anomaly detection and localization. In CVPR, pp. 9664–9674, 2021.
- Wenqian Liu, Runze Li, Meng Zheng, Srikrishna Karanam, Ziyan Wu, Bir Bhanu, Richard J Radke, and Octavia Camps. Towards visually explaining variational autoencoders. In *CVPR*, pp. 8642–8651, 2020.
- Yifan Mao, Fei-Fei Xue, Ruixuan Wang, Jianguo Zhang, Wei-Shi Zheng, and Hongmei Liu. Abnormality detection in chest x-ray images using uncertainty prediction autoencoders. In *MICCAI*, pp. 529–538. Springer, 2020.
- Felix Meissen, Johannes Paetzold, Georgios Kaissis, and Daniel Rueckert. Unsupervised anomaly localization with structural feature-autoencoders. In *Brainlesion: Glioma, Multiple Sclerosis, Stroke and Traumatic Brain Injuries*, pp. 14–24, Cham, 2023. Springer Nature Switzerland. ISBN 978-3-031-33842-7.
- Bjoern H Menze, Andras Jakab, Stefan Bauer, Jayashree Kalpathy-Cramer, Keyvan Farahani, Justin Kirby, Yuliya Burren, Nicole Porz, Johannes Slotboom, Roland Wiest, et al. The multimodal brain tumor image segmentation benchmark (brats). *IEEE transactions on medical imaging*, 34(10):1993–2024, 2014.
- Baharan Mirzasoleiman, Jeff Bilmes, and Jure Leskovec. Coresets for data-efficient training of machine learning models. In *International Conference on Machine Learning*, pp. 6950–6960. PMLR, 2020.
- Pramuditha Perera, Ramesh Nallapati, and Bing Xiang. Ocgan: One-class novelty detection using gans with constrained latent representations. In CVPR, pp. 2898–2906, 2019.
- Tal Reiss, Niv Cohen, Liron Bergman, and Yedid Hoshen. Panda: Adapting pretrained features for anomaly detection and segmentation. In CVPR, pp. 2806–2814, 2021.
- Karsten Roth, Latha Pemula, Joaquin Zepeda, Bernhard Schölkopf, Thomas Brox, and Peter Gehler. Towards total recall in industrial anomaly detection. In *CVPR*, pp. 14318–14328, 2022.
- Lukas Ruff, Robert Vandermeulen, Nico Goernitz, Lucas Deecke, Shoaib Ahmed Siddiqui, Alexander Binder, Emmanuel Müller, and Marius Kloft. Deep one-class classification. In *International conference on machine learning*, pp. 4393–4402. PMLR, 2018.
- Thomas Schlegl, Philipp Seeböck, Sebastian M Waldstein, Georg Langs, and Ursula Schmidt-Erfurth. f-anogan: Fast unsupervised anomaly detection with generative adversarial networks. *Medical image analysis*, 54:30–44, 2019.
- Anouk Stein, Carol Wu, Chris Carr, George Shih, Jamie Dulkowski, Jayashree Kalpathy-Cramer, Leon Chen, Luciano Prevedello, Marc Kohli, Mark McDonald, et al. Rsna pneumonia detection challenge, 2018. URL https://kaggle.com/competitions/rsna-pneumonia-detection-challenge.
- Max Welling. Herding dynamical weights to learn. In *Proceedings of the 26th Annual International Conference on Machine Learning*, pp. 1121–1128, 2009.
- Julian Wyatt, Adam Leach, Sebastian M Schmon, and Chris G Willcocks. Anoddpm: Anomaly detection with denoising diffusion probabilistic models using simplex noise. In CVPR, pp. 650–656, 2022.
- Han Xiao, Kashif Rasul, and Roland Vollgraf. Fashion-mnist: a novel image dataset for benchmarking machine learning algorithms. arXiv preprint arXiv:1708.07747, 2017.
- Suhang You, Kerem C Tezcan, Xiaoran Chen, and Ender Konukoglu. Unsupervised lesion detection via image restoration with a normative prior. In *International Conference on Medical Imaging with Deep Learning*, pp. 540–556. PMLR, 2019.
- Vitjan Zavrtanik, Matej Kristan, and Danijel Skočaj. Draem-a discriminatively trained reconstruction embedding for surface anomaly detection. In CVPR, pp. 8330–8339, 2021.
- Jianpeng Zhang, Yutong Xie, Yi Li, Chunhua Shen, and Yong Xia. Covid-19 screening on chest x-ray images using deep learning based anomaly detection. arXiv preprint arXiv:2003.12338, 27(10.48550), 2020.

- Chong Zhou and Randy C Paffenroth. Anomaly detection with robust deep autoencoders. In *Proceedings of the 23rd ACM SIGKDD international conference on knowledge discovery and data mining*, pp. 665–674, 2017.
- Xiangxin Zhu, Dragomir Anguelov, and Deva Ramanan. Capturing long-tail distributions of object subcategories. In CVPR, pp. 915–922, 2014.
- David Zimmerer, Fabian Isensee, Jens Petersen, Simon Kohl, and Klaus Maier-Hein. Unsupervised anomaly localization using variational auto-encoders. In *Medical Image Computing and Computer Assisted Intervention–MICCAI 2019: 22nd International Conference, Shenzhen, China, October 13–17, 2019, Proceedings, Part IV 22*, pp. 289–297. Springer, 2019.

A Appendix: Datasets

A.0.1 CIFAR10, CIFAR100, MNIST, and FashionMNIST

We follow the widely used training setup from Reiss et al. (2021) for these datasets. CIFAR10 contains 6000 images per class. For each class $c \in C$, we create a training dataset $X_{\text{train, c}}$ using 4000 samples from said class. The remaining samples are split equally into a validation and a test set and combined with the same amount of samples from every other class as outliers. Training, validation, and test sets for CIFAR100, MNIST, and Fashion-MNIST are created analogously. For CIFAR100, we used the 20 superclasses instead of the 100 detailed classes.

A.0.2 MVTec-AD

MVTec-AD is a dataset for defect detection in industrial production. Here, we used the original splits as outlined by Bergmann et al. (2019).

A.0.3 BraTS

The multimodal brain tumor image segmentation benchmark (BraTS) contains 369 MRI from patients with glioma. We extract 5 slices around the center line and use 101 slices without glioma for training. The remaining 80 normal slices are split 50:50 into a validation and a test set and are complemented with 40 pathological slices each. Following Meissen et al. (2023), we use the T2-weighted sequences, perform histogram equalization on the slices, and resize them to 128×128 .

A.0.4 RSNA and CheXpert

The RSNA Pneumonia Detection dataset (Stein et al., 2018) is a subset of 30 000 frontal view chest radiographs from the National Institutes of Health (NIH) CXR8 dataset that was manually labeled by 18 radiologists for one of the following labels: "Normal", "Lung Opacity", or "No Lung Opacity / Not Normal". The CheXpert database contains 224 316 chest-radiographs of 65 240 patients, acquired at Stanford Hospital with 13 structured diagnostic labels. To make the CheXpert compatible with RSNA, we only considered frontal-view images without support devices and further excluded those where any of the labels were marked as uncertain. In addition to the image data and labels, demographic information about the patients' gender and age was available for both datasets.

For RSNA, we used the "Normal" label as in-distribution images and combined the "pneumonia" and "No Opacity/Not Normal" (has lung opacities, but not suspicious for pneumonia) as OOD. Similarly, for CheXpert, the "No Finding" label was used as in-distribution, and "pneumonia" and "lung opacity" as OOD. For both datasets, we created a validation- and a test set that are balanced w.r.t. gender (male and female), age (young and old), the presence of anomalies, and contain 800 samples each. The remaining in-distribution samples were used for training. As part of preprocessing, all Chest X-ray images were center cropped and resized to 128×128 pixels. Note that we treated both datasets individually and did not combine them for training or evaluation.

B Appendix: Models

B.0.1 PANDA

PANDA (Reiss et al., 2021) is a model built upon DeepSVDD by Ruff et al. (2018). Like the latter, it relies on training a one-class classifier by using the compactness loss. Given a feature-extractor ψ and a center vector c, the compactness loss is defined as:

$$\mathcal{L}_{\text{compact}} = \sum_{x \in D} ||\psi(x) - c||^2.$$
 (7)

The center vector c is computed as the mean feature vector of the training dataset D on the untrained model ψ_0 :

$$c = \frac{1}{|D|} \sum_{x \in D} \psi_0(x) \,. \tag{8}$$

Instead of training a specialized architecture from scratch, PANDA benefits from useful features of pre-trained models. Specifically, it extracts features from the penultimate layer of a ResNet152 (He et al., 2016) and categorizes samples into ID / OOD by the L1-distance to the k nearest neighbors (kNN), with k=2. PANDA only fine-tunes layer3 and layer4 and uses early stopping to determine the optimal distance between ID and OOD samples. Training has no effect when using only one sample as the feature representation of the only sample is always identical to c. We used the original configuration from their paper for our experiments.

B.0.2 PatchCore

PatchCore (Roth et al., 2022) follows a similar concept as PANDA. However, instead of performing a kNN search on pooled, global features, PatchCore achieves localized anomaly detection by using a memory bank of locally-aware patch features \mathcal{M} instead. To make the kNN search computationally feasible, PatchCore performs core-set selection on the memory bank to retain only a subset of representative features \mathcal{M}_{sub} :

$$\mathcal{M}_{\text{sub}}^* = \underset{\mathcal{M}_{\text{sub}} \subset \mathcal{M}}{\min} \max_{m \in \mathcal{M}} \min_{n \in \mathcal{M}_{\text{sub}}} ||m - n||_2.$$
 (9)

Compared to PANDA, PatchCore does not require fine-tuning of the feature extractor. We used the same hyperparameters for PatchCore as in the original publication.

B.0.3 FAE

The Structural Feature-Autoencoder (FAE) (Meissen et al., 2023) extracts spatial feature maps from a pretrained and frozen feature extractor ψ (a ResNet18 He et al. (2016) in practice). The feature maps a resized and concatenated and fed into a convolutional autoencoder f_{θ} that is trained via the structural similarity (SSIM) loss for reconstruction:

$$\mathcal{L} = SSIM(\psi(x), f_{\theta}(\psi(x))). \tag{10}$$

Anomalies are detected using the residual between the feature maps and their reconstruction like in popular image-reconstruction models. It was found to perform best for UAD in Chest X-ray images in a study by Lagogiannis et al. (2023). We use a smaller model in our experiments since it still gave us the same performance on the RSNA full dataset as the larger model and is more resource-efficient. Specifically, we set the fae_hidden_dims parameter to [50, 100]. The other parameters were kept the same as in the original publication.

B.0.4 Reverse Distillation

The Reverse Distillation (RD) model by Deng & Li (2022) utilizes a frozen encoder model and a decoder that mirrors the former, similar to an Autoencoder. Instead of reconstructing the image, however, RD minimizes the cosine distance between feature maps in the encoder and decoder. The same measure is also used during inference to detect anomalies. RD was the second-best performing UAD model for Chest X-ray images in Lagogiannis et al. (2023). We used the same hyperparameters for RD as in Lagogiannis et al. (2023).

C Appendix: Implementation details

As suggested by Reiss et al. (2021), we train PANDA for a constant number of 2355 steps (corresponding to 15 epochs on CIFAR10) using the SGD optimizer with a learning rate of 0.01 and weight decay of 0.00005.

Table 2: Detailed training results for CIFAR10. AUROC scores of full training and training with 1, 5, 10, and 25 best-performing (with greedy search, the evolutionary algorithm, and core-set selection) or random samples. Since the performance of randomly selected subgroups can vary strongly, we repeated these experiments over ten different subsets. Best performances are marked in bold, and underlined numbers are the best per sample size.

	Method	Airplane	Automobile	Bird	Cat	Deer	Dog	Frog	Horse	Ship	Truck	Average
1 sample	Random Greedy Evo Core-set	$\begin{array}{c} 88.14 \pm 6.3 \\ \underline{96.18} \\ 86.15 \\ 94.54 \end{array}$	$\begin{array}{c} 91.59 \pm 4.1 \\ \underline{98.50} \\ 97.08 \\ 96.86 \end{array}$	$75.81 \pm 6.2 \\ \underline{88.04} \\ \underline{88.04} \\ 79.14$	$\begin{array}{c} 79.75 \pm 5.5 \\ \underline{91.59} \\ 78.61 \\ 81.56 \end{array}$	$\begin{array}{c} 90.55 \pm 4.7 \\ \underline{95.92} \\ 93.27 \\ 92.75 \end{array}$	$76.47 \pm 7.6 \\ \underline{90.13} \\ 79.83 \\ 84.63$	$\begin{array}{c} 88.38 \pm \! 5.1 \\ \underline{96.87} \\ 67.29 \\ 92.84 \end{array}$	$82.34 \pm 15.1 \\ \underline{96.94} \\ \overline{94.21} \\ 92.28$	87.19 ± 6.6 $\underline{98.14}$ 96.64 92.09	$\begin{array}{c} 89.32\ \pm 7.6\\ \underline{97.88}\\ 95.73\\ 94.15\end{array}$	$84.96 \pm 1.2 \\ \underline{95.02} \\ 87.68 \\ 90.08$
5 samples	Random Greedy Evo Core-set	$\begin{array}{c} 94.88 \pm\! 1.3 \\ \underline{97.22} \\ 95.85 \\ 94.14 \end{array}$	$\begin{array}{c} 97.00 \pm 1.2 \\ \underline{99.05} \\ 96.60 \\ 97.23 \end{array}$	82.88 ± 2.4 $\underline{90.36}$ 89.19 88.70	86.92 ± 2.7 $\underline{93.06}$ 87.44 87.88	$\begin{array}{c} 94.06 \pm 2.0 \\ \underline{96.86} \\ 96.12 \\ 95.91 \end{array}$	$84.61 \pm 4.5 \\ \underline{94.32} \\ 92.23 \\ 90.36$	$\begin{array}{c} 94.30 \ \pm 1.5 \\ \underline{98.31} \\ 93.89 \\ 95.24 \end{array}$	$93.68 \pm 2.0 \\ \underline{97.39} \\ 97.11 \\ 96.39$	$95.57 \pm 1.1 \underline{98.37} 96.72 96.22$	$\begin{array}{c} 95.58 \pm 1.0 \\ \underline{98.61} \\ 97.88 \\ 96.46 \end{array}$	$\begin{array}{c} 91.95 \pm 0.7 \\ \underline{96.36} \\ 94.30 \\ 93.85 \end{array}$
10 samples	Random Greedy Evo Core-set	$\begin{array}{c} 95.42 \pm \! 1.2 \\ \underline{97.19} \\ 96.47 \\ 94.90 \end{array}$	$\begin{array}{c} 97.65 \pm 0.8 \\ \underline{99.08} \\ 97.98 \\ 97.79 \end{array}$	$\begin{array}{c} 86.70 \pm 1.2 \\ \underline{91.45} \\ 90.64 \\ 88.56 \end{array}$	$\begin{array}{c} 88.02 \pm 1.6 \\ \underline{92.39} \\ 89.32 \\ 88.09 \end{array}$	$\begin{array}{c} 94.94 \pm \! 0.5 \\ \underline{96.97} \\ 96.62 \\ 95.91 \end{array}$	$\begin{array}{c} 89.00 \pm 2.0 \\ \underline{93.94} \\ 93.61 \\ 91.50 \end{array}$	$\begin{array}{c} 95.82 \pm \! 0.8 \\ \underline{98.79} \\ 97.72 \\ 96.31 \end{array}$	$\begin{array}{c} 95.57 \pm 1.3 \\ \underline{97.01} \\ 96.93 \\ 96.46 \end{array}$	$\begin{array}{c} 96.05 \pm \! 0.7 \\ \underline{98.14} \\ 97.62 \\ 96.45 \end{array}$	$96.74 \pm 0.5 \\ \underline{98.79} \\ 98.23 \\ 96.81$	$\begin{array}{c} 93.59 \pm 0.4 \\ \underline{96.37} \\ 95.51 \\ 94.28 \end{array}$
25 samples	Random Greedy Evo Core-set	$\begin{array}{c} 96.19\ \pm0.7\\ \underline{97.11}\\ 96.56\\ 96.88 \end{array}$	$\begin{array}{c} 98.30\ \pm0.3\\ \underline{\textbf{99.17}}\\ 98.18\\ 98.67\end{array}$	$\begin{array}{c} 89.26 \pm 1.2 \\ 90.22 \\ \underline{91.38} \\ 89.06 \end{array}$	$\begin{array}{c} 88.26 \pm 1.7 \\ \underline{92.46} \\ 89.16 \\ 86.26 \end{array}$	$\begin{array}{c} 95.77 \pm 0.5 \\ 96.61 \\ \underline{96.68} \\ 95.98 \end{array}$	$\begin{array}{c} 91.16 \pm 1.9 \\ \underline{\textbf{95.00}} \\ 92.11 \\ 92.98 \end{array}$	$\begin{array}{c} 96.95 \pm\! 0.6 \\ \underline{98.87} \\ 97.87 \\ 97.40 \end{array}$	$\begin{array}{c} 96.59 \pm 0.5 \\ 96.93 \\ \underline{97.42} \\ 96.99 \end{array}$	$\begin{array}{c} 96.77 \pm 0.2 \\ \underline{97.87} \\ 97.64 \\ 96.68 \end{array}$	$\begin{array}{c} 97.50\ \pm0.3\\ \underline{98.65}\\ 98.08\\ 97.56\end{array}$	$\begin{array}{c} 94.68 \pm \! 0.2 \\ \underline{96.29} \\ 95.51 \\ 94.85 \end{array}$
	Full training	97.92	98.76	94.12	90.43	97.53	94.36	98.44	97.83	98.21	98.23	96.58

Table 3: Detailed training results for MNIST. AUROC scores of full training and training with 1, 5, 10, and 25 best-performing (with greedy search, the evolutionary algorithm, and core-set selection) or random samples. Since the performance of randomly selected subgroups can vary strongly, we repeated these experiments over ten different subsets. Best performances are marked in bold, and underlined numbers are the best per sample size.

	Method	0	1	2	3	4	5	6	7	8	9	Average
1 sample	Random Greedy Evo Core-set	$\begin{array}{c} 91.11 \pm\! 2.4 \\ \underline{98.14} \\ 82.31 \\ 94.71 \end{array}$	$\begin{array}{c} 90.34\ \pm 6.0\\ \underline{97.09}\\ 83.30\\ 92.27\end{array}$	$65.18 \pm 10.6 \\ \underline{82.74} \\ 69.87 \\ 71.53$	$77.02 \pm 4.4 \\ \underline{89.92} \\ 75.18 \\ 86.66$	$69.79 \pm 11.2 \\ \underline{92.94} \\ 78.13 \\ 80.33$	$69.69 \pm 9.7 \\ \underline{88.69} \\ 82.36 \\ 78.28$	$69.43 \pm 6.9 \\ \underline{85.01} \\ 78.57 \\ 75.39$	$70.37 \pm 7.6 \\ \underline{87.87} \\ 59.38 \\ 76.65$	$76.91 \pm 3.6 \\ \underline{89.59} \\ 62.56 \\ 82.48$	$81.26 \pm 5.4 \\ \underline{89.53} \\ 76.56 \\ 84.61$	$76.11 \pm 2.2 \\ \underline{90.15} \\ 74.82 \\ 82.29$
5 samples	Random Greedy Evo Core-set	$\begin{array}{c} 98.52\ \pm0.9\\ 99.21\\ 98.76\\ \underline{99.29}\end{array}$	$\begin{array}{c} 97.79\ \pm2.2\\ 98.29\\ 96.96\\ \underline{99.58}\end{array}$	$77.40 \pm 7.3 \\ 85.59 \\ 85.06 \\ \underline{86.83}$	$\begin{array}{c} 89.79\ \pm 1.9\\ 93.13\\ \underline{94.82}\\ 92.32\end{array}$	$\begin{array}{c} 90.25 \pm 1.9 \\ 95.65 \\ \underline{95.78} \\ 89.07 \end{array}$	$\begin{array}{c} 85.58 \pm 4.9 \\ 90.90 \\ \underline{91.44} \\ 91.30 \end{array}$	$\begin{array}{c} 88.90 \pm 4.3 \\ 93.00 \\ \underline{95.57} \\ 94.72 \end{array}$	$\begin{array}{c} 86.97 \pm \! 2.6 \\ \underline{92.52} \\ 89.28 \\ 91.68 \end{array}$	$\begin{array}{c} 87.92 \pm \! 3.5 \\ \underline{94.31} \\ 87.35 \\ 91.88 \end{array}$	$\begin{array}{c} 90.42\ \pm2.1\\ 93.56\\ \underline{94.59}\\ 88.82\end{array}$	$\begin{array}{c} 89.36 \pm 1.1 \\ \underline{93.62} \\ 92.96 \\ 92.55 \end{array}$
10 samples	Random Greedy Evo Core-set	$\begin{array}{c} 98.86 \pm 0.6 \\ \underline{99.43} \\ 99.40 \\ 99.38 \end{array}$	$\begin{array}{c} 99.18 \pm 0.4 \\ 98.83 \\ 99.30 \\ \underline{99.66} \end{array}$	$83.04 \pm 4.0 \\ 89.00 \\ 88.08 \\ \underline{90.60}$	$\begin{array}{c} 92.39 \pm 2.0 \\ 93.99 \\ \underline{94.42} \\ 94.23 \end{array}$	$\begin{array}{c} 93.82 \pm 2.3 \\ 96.89 \\ \underline{97.97} \\ 96.27 \end{array}$	$\begin{array}{c} 90.84 \pm 1.3 \\ 91.39 \\ \underline{92.40} \\ 91.46 \end{array}$	$\begin{array}{c} 92.81 \pm 2.8 \\ 95.62 \\ \underline{97.59} \\ 95.65 \end{array}$	$\begin{array}{c} 93.28 \pm 1.2 \\ 93.97 \\ \underline{97.28} \\ 93.56 \end{array}$	$\begin{array}{c} 92.34 \pm \! 1.8 \\ \underline{96.28} \\ 92.17 \\ 93.45 \end{array}$	$\begin{array}{c} 93.40 \pm 1.1 \\ 93.75 \\ 95.09 \\ \underline{95.16} \end{array}$	$\begin{array}{c} 93.00 \pm 0.6 \\ 94.91 \\ \underline{95.37} \\ 94.94 \end{array}$
25 samples	Random Greedy Evo Core-set	99.38 ±0.2 99.67 99.44 99.76 99.84	$\begin{array}{c} 99.70 \pm 0.1 \\ 99.30 \\ \underline{99.72} \\ 99.71 \\ \hline \\ 99.93 \\ \end{array}$	$\begin{array}{c} 90.26 \pm 2.1 \\ 89.85 \\ 91.65 \\ \underline{92.42} \\ 96.58 \end{array}$	$\begin{array}{c} 95.36 \pm 1.5 \\ 96.63 \\ \underline{97.09} \\ 96.22 \\ \hline 98.02 \end{array}$	$96.66 \pm 1.0 \\ 97.67 \\ \underline{99.01} \\ 97.04$ 98.63	$\begin{array}{c} 93.37 \pm 1.0 \\ 90.84 \\ 95.10 \\ \underline{95.96} \\ \\ 96.45 \end{array}$	97.55 ± 1.3 98.32 99.23 98.89 99.13	97.49 ± 0.5 94.88 98.70 98.66 99.14	95.83 ± 0.6 97.86 95.43 97.37 97.82	96.79 ± 0.5 95.60 97.60 97.23 98.53	$96.24 \pm 0.3 \\ 96.06 \\ 97.30 \\ \underline{97.33} \\ 98.41$

FAE converges very fast on BraTS, RSNA, and CheXpert, so it was trained for 500 steps using the Adam optimizer with a learning rate of 0.0002. In our experiments, the evolutionary algorithm was robust to the population size P and the number of generations G. In all experiments, we therefore empirically set P = 1000 and G = 500. For all models, we used the official PyTorch implementations by the authors.

D Appendix: Detailed results

We show the detailed per-class results for CIFAR10, CIFAR100, MNIST, Fashion-MNIST, and MVTec-AD in Tables 2 to 5, respectively.

Table 4: Detailed training results for Fashion-MNIST. AUROC scores of full training and training with 1, 5, 10, and 25 best-performing (with greedy search, the evolutionary algorithm, and core-set selection) or random samples. Since the performance of randomly selected subgroups can vary strongly, we repeated these experiments over ten different subsets. Best performances are marked in bold, and underlined numbers are the best per sample size.

	Method	0	1	2	3	4	5	6	7	8	9	Average
1 sample	Random Greedy Evo Core-set	$77.24 \pm 11.0 \\ \underline{93.85} \\ 89.28 \\ 90.94$	$\begin{array}{c} 96.45 \pm \! 0.8 \\ \underline{98.12} \\ 83.19 \\ 97.12 \end{array}$	$82.47 \pm 17.5 \\ \underline{93.03} \\ 88.71 \\ 91.63$	$73.80 \pm 7.5 \\ \underline{88.85} \\ 83.60 \\ 83.53$	$75.09 \pm 12.5 \\ \underline{90.46} \\ 79.98 \\ 88.62$	$\begin{array}{c} 91.63 \pm 4.1 \\ \underline{97.14} \\ 94.35 \\ 95.75 \end{array}$	$69.29 \pm 12.4 \\ \underline{84.55} \\ 71.09 \\ 76.01$	$\begin{array}{c} 94.71\ \pm 6.2\\ \underline{98.84}\\ 96.53\\ 98.67\end{array}$	$79.04 \pm 8.4 \\ \underline{95.99} \\ 84.16 \\ 94.42$	$\begin{array}{c} 95.37 \pm\! 4.6 \\ \underline{99.16} \\ 97.30 \\ 98.00 \end{array}$	$\begin{array}{c} 83.51\ \pm 3.3\\ \underline{94.00}\\ 86.82\\ 91.47\end{array}$
5 samples	Random Greedy Evo Core-set	$\begin{array}{c} 90.58 \pm \! 3.7 \\ \underline{94.64} \\ 92.74 \\ 92.05 \end{array}$	$\begin{array}{c} 97.90 \pm 0.5 \\ \underline{98.98} \\ 98.85 \\ 97.95 \end{array}$	$\begin{array}{c} 92.13 \pm\! 1.8 \\ \underline{94.27} \\ 93.94 \\ 92.91 \end{array}$	$\begin{array}{c} 85.24 \pm 5.4 \\ \underline{92.38} \\ 89.42 \\ 91.24 \end{array}$	$\begin{array}{c} 88.77 \pm\! 1.3 \\ \underline{92.95} \\ 88.10 \\ 89.85 \end{array}$	$\begin{array}{c} 95.38 \pm 1.9 \\ \underline{97.98} \\ 94.19 \\ 97.32 \end{array}$	$\begin{array}{c} 80.89\ \pm2.4\\ \underline{86.42}\\ 83.62\\ 84.05\end{array}$	$\begin{array}{c} 98.34\ \pm0.8\\ \underline{99.18}\\ 98.59\\ 98.61\end{array}$	$\begin{array}{c} 91.01\ \pm 6.7\\ \underline{96.84}\\ 96.30\\ 96.10\end{array}$	$\begin{array}{c} 98.41 \pm\! 1.2 \\ \underline{99.42} \\ 98.43 \\ 98.87 \end{array}$	$\begin{array}{c} 91.86 \pm 1.1 \\ \underline{95.31} \\ 93.42 \\ 93.89 \end{array}$
10 samples	Random Greedy Evo Core-set	$\begin{array}{c} 91.78 \pm 2.1 \\ \underline{94.79} \\ 91.41 \\ 93.77 \end{array}$	$\begin{array}{c} 98.31\ \pm0.5\\ 98.98\\ \underline{99.31}\\ 98.85\end{array}$	$\begin{array}{c} 93.22 \pm 0.9 \\ 94.53 \\ \underline{94.57} \\ 94.14 \end{array}$	$\begin{array}{c} 89.62 \pm 2.4 \\ 93.38 \\ \underline{94.72} \\ 92.82 \end{array}$	$\begin{array}{c} 89.87 \pm \! 0.7 \\ \underline{92.00} \\ 90.11 \\ 91.40 \end{array}$	$\begin{array}{c} 96.24 \pm 1.3 \\ \underline{98.55} \\ 95.62 \\ 96.13 \end{array}$	$\begin{array}{c} 81.83 \pm 1.9 \\ \underline{86.74} \\ 82.86 \\ 84.14 \end{array}$	$\begin{array}{c} 98.57 \pm \! 0.7 \\ \underline{99.27} \\ 98.93 \\ 98.72 \end{array}$	$\begin{array}{c} 94.31 \pm\! 1.6 \\ 96.02 \\ \underline{96.90} \\ 95.98 \end{array}$	$\begin{array}{c} 98.62 \pm \! 0.7 \\ \underline{99.50} \\ 98.37 \\ 98.70 \end{array}$	$\begin{array}{c} 93.24 \pm 0.3 \\ \underline{95.38} \\ 94.28 \\ 94.47 \end{array}$
25 samples	Random Greedy Evo Core-set	93.28 ±1.2 95.06 94.10 94.40	98.62 ±0.3 99.03 99.24 99.18	93.74 ±0.7 94.64 94.12 94.16	93.37 ±1.6 94.49 95.15 94.70	91.48 ±0.9 92.97 92.97 91.64	$\begin{array}{c} 95.86 \pm 1.0 \\ \underline{98.54} \\ 95.70 \\ 97.07 \end{array}$	82.45 ±1.4 <u>86.76</u> 84.11 84.80	98.89 ±0.2 <u>99.26</u> 98.94 99.13	95.31 ±1.0 94.86 97.32 <u>97.37</u>	98.62 ±0.5 <u>99.60</u> 99.00 98.87	94.16 ±0.2 <u>95.52</u> 95.07 95.13
	Full training	95.09	99.52	94.44	96.28	93.66	96.58	85.33	99.28	98.88	98.86	95.79

Table 5: Detailed training results for MVTec-AD. AUROC scores of full training and training with 1, 5, 10, and 25 best-performing (with greedy search and evolutionary algorithm) or random samples. Since the performance of randomly selected subgroups can vary strongly, we repeated these experiments over ten different subsets. Best performances are marked in bold, and underlined numbers are the best per sample size.

	Method	Bottle	Cable	Capsule	Carpet	Grid	Hazelnut	Leather	Metal nut
1 sample	Random Greedy Evo Core-set	$\begin{array}{c} 99.71 \pm 0.1 \\ \underline{99.76} \\ 99.52 \\ 99.52 \end{array}$	$83.74 \pm 4.7 \\ 88.19 \\ 88.92 \\ \underline{89.81}$	$66.80 \pm 5.1 \\ \underline{72.80} \\ 63.14 \\ 61.55$	$\begin{array}{c} 97.72\ \pm0.5\\ 98.60\\ 98.23\\ \underline{99.08} \end{array}$	$60.30 \pm 6.9 \\ \underline{71.19} \\ 66.88 \\ 65.99$	$\begin{array}{c} 90.67 \pm 2.6 \\ \underline{93.93} \\ 93.04 \\ 88.68 \end{array}$	$\begin{array}{c} 99.99 \pm 0.0 \\ \underline{100.00} \\ \underline{100.00} \\ \underline{100.00} \end{array}$	71.99 ± 4.0 72.19 70.97 71.07
5 samples	Random Greedy Evo Core-set	$\begin{array}{c} 99.84\ \pm0.2\\ \underline{\textbf{100.00}}\\ 99.52\\ 99.68 \end{array}$	$\begin{array}{c} 91.40 \pm\! 3.6 \\ \underline{96.27} \\ 93.22 \\ 92.82 \end{array}$	$\begin{array}{c} 84.11\ \pm 7.2\\ 84.96\\ \underline{90.91}\\ 87.36\end{array}$	$\begin{array}{c} 97.98 \pm 0.3 \\ \underline{98.64} \\ 98.19 \\ 98.72 \end{array}$	$72.64 \pm 7.1 \\ 73.52 \\ \underline{86.04} \\ 69.40$	$\begin{array}{c} 96.29 \pm 2.2 \\ 98.29 \\ \underline{99.14} \\ 98.64 \end{array}$	$\begin{array}{c} \textbf{100.00} \; \pm 0.0 \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \end{array}$	$\begin{array}{c} 94.93 \pm \! 3.5 \\ 97.75 \\ \underline{98.34} \\ 96.82 \end{array}$
25 samples 10 samples	Random Greedy Evo Core-set	$\begin{array}{c} 99.90 \pm 0.2 \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \\ 99.60 \end{array}$	$\begin{array}{c} 93.49\ \pm1.8\\ \underline{97.58}\\ 96.42\\ 92.75\end{array}$	$\begin{array}{c} 90.29 \pm 2.3 \\ \underline{93.06} \\ 91.26 \\ 90.75 \end{array}$	$\begin{array}{c} 98.04 \pm 0.3 \\ 98.15 \\ 98.39 \\ \underline{98.52} \end{array}$	$\begin{array}{c} 80.91\ \pm 5.7\\ 78.86\\ \underline{94.40}\\ 76.38\end{array}$	$\begin{array}{c} 98.99 \pm 0.7 \\ 99.64 \\ \underline{\textbf{100.00}} \\ 99.68 \end{array}$	$\begin{array}{c} \textbf{100.00} \pm 0.0 \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \end{array}$	$\begin{array}{c} 97.72 \pm 1.6 \\ 98.92 \\ \underline{99.07} \\ 98.19 \end{array}$
25 samples	Random Greedy Evo Core-set	$\begin{array}{c} \underline{100.00} \pm 0.0 \\ \underline{100.00} \\ \underline{100.00} \\ \underline{99.68} \end{array}$	$\begin{array}{c} 96.59\ \pm1.0\\ 96.74\\ \underline{98.29}\\ 97.49\end{array}$	$\begin{array}{c} 93.61 \pm 1.5 \\ 93.94 \\ \underline{94.34} \\ 91.58 \end{array}$	$\begin{array}{c} 98.14 \pm\! 0.3 \\ 98.27 \\ 98.56 \\ \underline{98.88} \end{array}$	$\begin{array}{c} 90.23 \pm 2.9 \\ 83.88 \\ \underline{\textbf{99.11}} \\ 92.86 \end{array}$	$\begin{array}{c} 99.81 \pm 0.3 \\ 99.71 \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \end{array}$	$\begin{array}{c} \textbf{100.00} \; \pm 0.0 \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \\ \underline{\textbf{100.00}} \end{array}$	$\begin{array}{c} 99.20 \pm 0.4 \\ 99.41 \\ \underline{99.76} \\ 99.46 \end{array}$
	Full training	100.00	99.53	99.20	98.43	99.08	100.00	100.00	99.90
	Method	Pill	Screw	Tile	${\bf Toothbrush}$	Transistor	Wood	Zipper	Average
1 sample	Random Greedy Evo Core-set	Pill 78.71 ±5.0 89.23 72.64 74.58	Screw 46.22 ± 4.8 $\underline{55.87}$ 52.96 38.29	Tile 99.59 ± 0.5 100.00 99.13 98.63	Toothbrush 82.58 ± 3.5 88.61 90.00 85.83	$\begin{array}{c} \textbf{Transistor} \\ 83.31 \pm 4.7 \\ 91.42 \\ \underline{92.12} \\ 82.79 \end{array}$	Wood 98.18 ± 0.7 99.21 99.04 99.21	Zipper 94.58 ± 1.5 99.37 96.61 95.64	Average 83.61 ± 1.2 89.70 85.55 82.80
	Random Greedy Evo	$78.71 \pm 5.0 \\ \underline{89.23} \\ 72.64$	$46.22 \pm 4.8 \\ \underline{55.87} \\ 52.96$	$99.59 \pm 0.5 \\ \underline{100.00} \\ 99.13$	$82.58 \pm 3.5 \\ 88.61 \\ \underline{90.00}$	$83.31 \pm 4.7 \\ 91.42 \\ \underline{92.12}$	$98.18 \pm 0.7 \\ \underline{99.21} \\ 99.04$	$94.58 \pm 1.5 \underline{99.37} 96.61$	$83.61 \pm 1.2 \\ \underline{89.70} \\ 85.55$
$\mid 5 \text{ samples} \mid 1$	Random Greedy Evo Core-set Random Greedy Evo	78.71 ± 5.0 $\underline{89.23}$ 72.64 74.58 89.48 ± 2.0 89.77 $\underline{91.41}$	$46.22 \pm 4.8 \\ \underline{55.87} \\ 52.96 \\ 38.29 \\ 52.86 \pm 5.1 \\ 53.40 \\ \underline{61.47}$	$\begin{array}{c} 99.59\ \pm0.5\\ \underline{\textbf{100.00}}\\ 99.13\\ 98.63\\ \\ 99.87\ \pm0.1\\ \underline{\textbf{100.00}}\\ 99.49\\ \end{array}$	$82.58 \pm 3.5 \\ 88.61 \\ \underline{90.00} \\ 85.83 \\ 87.22 \pm 5.2 \\ 87.22 \\ \underline{98.33}$	$83.31 \pm 4.7 \\ 91.42 \\ \underline{92.12} \\ 82.79 \\ 93.77 \pm 1.7 \\ \underline{94.46} \\ \underline{97.58}$	$\begin{array}{c} 98.18 \pm \! 0.7 \\ \underline{99.21} \\ 99.04 \\ \underline{99.21} \\ \\ 98.57 \pm \! 0.4 \\ \underline{99.30} \\ \underline{99.30} \\ \end{array}$	$\begin{array}{c} 94.58 \pm 1.5 \\ \underline{99.37} \\ 96.61 \\ 95.64 \\ \\ 97.55 \pm 1.5 \\ \underline{99.16} \\ 98.90 \\ \end{array}$	$\begin{array}{c} 83.61 \pm 1.2 \\ \underline{89.70} \\ 85.55 \\ 82.80 \\ \hline 90.43 \pm 0.9 \\ 91.52 \\ \underline{94.12} \\ \end{array}$
samples 1	Random Greedy Evo Core-set Random Greedy Evo Core-set Random Greedy Evo Evo Core-set	78.71 ± 5.0 89.23 72.64 74.58 89.48 ± 2.0 89.77 91.41 85.00 90.95 ± 1.8 93.43 93.40	$46.22 \pm 4.8 \\ \underline{55.87} \\ 52.96 \\ 38.29$ $52.86 \pm 5.1 \\ 53.40 \\ \underline{61.47} \\ 52.82$ $58.42 \pm 4.0 \\ 53.68 \\ \underline{75.90}$	$\begin{array}{c} 99.59 \pm \! 0.5 \\ \underline{\textbf{100.00}} \\ 99.13 \\ 98.63 \\ \\ 99.87 \pm \! 0.1 \\ \underline{\textbf{100.00}} \\ 99.49 \\ 99.57 \\ \\ 99.89 \pm \! 0.1 \\ \underline{\textbf{100.00}} \\ 99.71 \\ \end{array}$	$\begin{array}{c} 82.58 \pm \! 3.5 \\ 88.61 \\ \underline{90.00} \\ 85.83 \\ 87.22 \pm \! 5.2 \\ 87.22 \\ \underline{98.33} \\ \underline{98.89} \\ 90.75 \pm \! 1.1 \\ 85.56 \\ \underline{99.44} \end{array}$	$\begin{array}{c} 83.31 \pm 4.7 \\ 91.42 \\ \underline{92.12} \\ 82.79 \\ \\ 93.77 \pm 1.7 \\ \underline{94.46} \\ \underline{97.58} \\ \underline{96.50} \\ \\ 95.86 \pm 1.8 \\ \underline{96.38} \\ \underline{99.58} \\ \end{array}$	$\begin{array}{c} 98.18 \pm \! 0.7 \\ \underline{99.21} \\ 99.04 \\ \underline{99.21} \\ \\ 98.57 \pm \! 0.4 \\ \underline{99.30} \\ \underline{99.30} \\ \underline{98.77} \\ \\ 98.60 \pm \! 0.2 \\ \underline{99.30} \\ \underline{99.39} \\ \end{array}$	$\begin{array}{c} 94.58 \pm 1.5 \\ \underline{99.37} \\ 96.61 \\ 95.64 \\ \\ 97.55 \pm 1.5 \\ \underline{99.16} \\ 98.90 \\ 95.93 \\ \\ \hline 98.13 \pm 1.0 \\ \underline{99.58} \\ 98.58 \\ \end{array}$	$\begin{array}{c} 83.61 \pm 1.2 \\ \underline{89.70} \\ 85.55 \\ 82.80 \\ \hline 90.43 \pm 0.9 \\ 91.52 \\ \underline{94.12} \\ 92.13 \\ \hline 92.80 \pm 0.6 \\ \underline{92.94} \\ \underline{96.37} \\ \end{array}$

Table 6: Detailed training results for CIFAR100. AUROC scores of full training and training with 1, 5, 10, and 25 best-performing (with greedy search, the evolutionary algorithm, and core-set selection) or random samples. Since the performance of randomly selected subgroups can vary strongly, we repeated these experiments over ten different subsets. Best performances are marked in bold, and underlined numbers are the best per sample size.

	Method	Aquatic mammals	Fish	Flowers	Food containers	Fruit and vegetables	Household electrical devices	Household furniture	Insects	Large carnivores	Large man-made outdoor things
1 sample	Random Greedy Evo Core-set	71.70 ± 14.7 91.93 89.68 64.34	75.23 ±8.1 <u>89.09</u> 78.96 75.71	90.83 ± 3.9 96.78 80.31 90.52	$\begin{array}{c} 82.25 \pm \! 3.7 \\ \underline{89.39} \\ 76.69 \\ 80.05 \end{array}$	$71.29 \pm 12.5 \\ \underline{90.97} \\ 84.76 \\ 85.14$	$71.88 \pm 7.1 \\ \underline{85.73} \\ 80.61 \\ 73.08$	$\begin{array}{c} 84.70 \pm 6.1 \\ \underline{96.17} \\ 92.09 \\ 90.44 \end{array}$	$68.09 \pm 12.3 \\ \underline{84.78} \\ 63.07 \\ 61.80$	$77.95 ^{11.7} \\ \underline{92.46} \\ 85.80 \\ 68.79$	86.72 ± 6.0 $\underline{93.98}$ 90.19 89.97
5 samples	Random Greedy Evo Core-set	86.25 ± 3.8 $\underline{94.13}$ 93.04 86.53	$81.82 \pm 6.1 \underline{92.32} 89.85 88.90$	95.82 ± 1.4 98.13 97.08 96.48	$\begin{array}{r} 89.31\ \pm 2.4\\ \underline{95.59}\\ 85.89\\ 91.79\end{array}$	$\begin{array}{c} 85.26 \pm 5.9 \\ \underline{93.00} \\ 92.47 \\ 91.29 \end{array}$	$78.90 \pm 6.2 \\ 85.05 \\ \underline{89.12} \\ 86.67$	$\begin{array}{c} 92.54 \pm 2.0 \\ \underline{96.78} \\ 95.64 \\ 95.75 \end{array}$	$\begin{array}{c} 79.26 \pm 9.0 \\ \underline{91.05} \\ 86.96 \\ 87.01 \end{array}$	$\begin{array}{c} 87.47\ \pm1.7\\ \underline{93.73}\\ 90.11\\ 91.77\end{array}$	$\begin{array}{c} 92.15 \pm 1.5 \\ \underline{95.55} \\ 93.78 \\ 92.95 \end{array}$
samples 10 samples	Random Greedy Evo Core-set	88.50 ± 1.9 $\underline{93.84}$ 93.19 91.31	85.56 ± 4.8 91.92 89.98 90.85	96.97 ±0.7 <u>98.60</u> 98.16 97.41	91.31 ± 1.7 95.97 91.44 92.08	$\begin{array}{c} 90.59\ \pm 3.1\\ 92.88\\ \underline{93.24}\\ 92.28 \end{array}$	$\begin{array}{c} 80.97 \pm\! 4.3 \\ 82.56 \\ \underline{88.27} \\ 86.71 \end{array}$	$\begin{array}{c} 94.77 \pm 0.8 \\ 96.81 \\ \underline{96.86} \\ 95.99 \end{array}$	81.96 ±5.0 <u>89.68</u> 88.92 85.73	$\begin{array}{c} 90.22 \pm 1.4 \\ \underline{93.33} \\ 90.95 \\ 92.60 \end{array}$	92.49 ±1.2 <u>95.57</u> 93.79 94.00
25 samples	Random Greedy Evo Core-set	90.04 ± 1.3 92.36 92.88 91.72	$90.78 \pm 1.2 91.27 \underline{92.50} 91.87$	97.87 ±0.5 <u>98.50</u> 98.39 98.23	$\begin{array}{c} 93.19\ \pm 1.5\\ \underline{95.79}\\ 92.87\\ 94.68\end{array}$	$\begin{array}{c} 93.63 \pm 0.6 \\ 92.53 \\ \underline{95.00} \\ 94.94 \end{array}$	$\begin{array}{c} 84.21 \pm 4.1 \\ 80.51 \\ 90.37 \\ \underline{90.80} \end{array}$	96.28 ± 0.5 97.26 96.84 96.21	$\begin{array}{c} 86.34 \pm 1.7 \\ \underline{90.89} \\ 90.74 \\ 90.31 \end{array}$	$\begin{array}{c} 91.82 \pm 0.8 \\ \underline{93.21} \\ 91.84 \\ 93.09 \end{array}$	93.50 ± 0.6 $\underline{95.45}$ 94.29 93.92
	Full training	93.68	94.81	98.46	95.86	96.59	94.59	97.26	93.00	95.34	94.98
_											
	Method	Large natural outdoor scenes	Large omnivores and herbivores	Medium-sized mammals	Non-insect invertebrates	People	Reptiles	Small mammals	Trees	Vehicles 1	Vehicles 2
1 sample	Method Random Greedy Evo Core-set		omnivores			People 88.73 ±6.4 <u>96.20</u> 83.52 91.10	Reptiles 66.99 ± 7.5 79.39 52.97 72.36		Trees 92.30 ±3.6 95.92 95.58 94.28	Vehicles 1 78.59 ±8.2 93.20 68.73 82.56	Vehicles 2 73.74 ±6.6 89.81 72.15 80.49
	Random Greedy Evo	0utdoor scenes 86.87 ±6.3 94.13 88.88	omnivores and herbivores $66.79 \pm 14.8 \\ 86.01 \\ 78.12$	73.91 ±7.6 84.57 73.68		88.73 ±6.4 96.20 83.52	$66.99 \pm 7.5 $	72.46 ±9.3 88.86 86.70	$\begin{array}{c} 92.30 \pm 3.6 \\ \underline{95.92} \\ 95.58 \end{array}$	$78.59 \pm 8.2 \\ \underline{93.20} \\ 68.73$	73.74 ± 6.6 $\underline{89.81}$ 72.15
5 samples 1	Random Greedy Evo Core-set Random Greedy Evo	0utdoor scenes 86.87 ±6.3 94.13 88.88 93.57 92.30 ±2.0 94.98 95.11	omnivores and herbivores 66.79 ± 14.8 $86.01 \over 78.12$ 74.96 82.54 ± 3.3 90.29 87.96	$\begin{array}{c} \textbf{mammals} \\ \hline 73.91 \pm 7.6 \\ \underline{84.57} \\ 73.68 \\ 79.55 \\ \hline 84.66 \pm 2.1 \\ \underline{89.08} \\ 83.67 \\ \hline \end{array}$		$\begin{array}{c} 88.73 \pm 6.4 \\ \underline{96.20} \\ 83.52 \\ 91.10 \\ \\ 95.66 \pm 1.2 \\ \underline{97.90} \\ 97.33 \end{array}$	66.99 ±7.5 79.39 52.97 72.36 79.22 ±3.2 84.47 81.79	$\begin{array}{c} \textbf{mammals} \\ \hline 72.46 \pm 9.3 \\ \underline{88.86} \\ 86.70 \\ 87.23 \\ \hline 85.18 \pm 4.6 \\ \underline{92.49} \\ 90.32 \\ \\ \end{array}$	$\begin{array}{c} 92.30 \pm 3.6 \\ \underline{95.92} \\ 95.58 \\ 94.28 \\ \\ 95.67 \pm 0.9 \\ \underline{96.89} \\ 96.83 \\ \end{array}$	78.59 ± 8.2 $\underline{93.20}$ 68.73 82.56 88.96 ± 3.2 $\underline{95.61}$ 92.12	73.74 ± 6.6 $\underline{89.81}$ 72.15 80.49 83.75 ± 3.8 $\underline{90.10}$ 87.22
samples 1	Random Greedy Evo Core-set Random Greedy Evo Core-set Random Greedy Evo	$\begin{array}{c} \textbf{outdoor scenes} \\ 86.87 \pm 6.3 \\ \underline{94.13} \\ 88.88 \\ 93.57 \\ 92.30 \pm 2.0 \\ 94.98 \\ \underline{95.11} \\ 93.92 \\ \\ 93.76 \pm 1.2 \\ 95.49 \\ 94.95 \end{array}$	$\begin{array}{c} \textbf{omnivores} \\ \textbf{and herbivores} \\ \hline 66.79 \pm 144.8 \\ \underline{86.01} \\ 78.12 \\ 74.96 \\ 82.54 \pm 3.3 \\ \underline{90.29} \\ 87.96 \\ 86.92 \\ 85.27 \pm 2.7 \\ 89.41 \\ \underline{89.90} \\ \end{array}$	$\begin{array}{c} \textbf{mammals} \\ \hline 73.91 \pm 7.6 \\ \underline{84.57} \\ 73.68 \\ 79.55 \\ \hline 84.66 \pm 2.1 \\ \underline{89.08} \\ 83.67 \\ 87.56 \\ \hline 87.79 \pm 1.3 \\ \underline{90.43} \\ 88.72 \\ \hline \end{array}$	$\begin{array}{c} \textbf{invertebrates} \\ 60.08 \pm 8.3 \\ \hline 79.51 \\ 77.34 \\ 56.37 \\ \hline 71.37 \pm 5.1 \\ \underline{82.67} \\ 81.76 \\ 75.77 \\ \hline 78.13 \pm 2.9 \\ \underline{85.01} \\ 83.32 \\ \end{array}$	$\begin{array}{c} 88.73 \pm 6.4 \\ \underline{96.20} \\ 83.52 \\ 91.10 \\ \hline 95.66 \pm 1.2 \\ \underline{97.90} \\ 97.33 \\ 95.14 \\ \hline 96.39 \pm 0.8 \\ \underline{98.18} \\ \underline{97.69} \end{array}$	$\begin{array}{c} 6.99 \pm 7.5 \\ \hline 79.39 \\ 52.97 \\ 72.36 \\ \hline 79.22 \pm 3.2 \\ 84.47 \\ 81.79 \\ \hline 85.19 \\ \hline 83.14 \pm 3.0 \\ \hline 87.55 \\ 85.42 \\ \end{array}$	72.46 ±9.3 88.86 86.70 87.23 85.18 ±4.6 92.49 90.32 86.94 87.63 ±2.7 91.98 90.41	$\begin{array}{c} 92.30 \pm 3.6 \\ \underline{95.92} \\ 95.58 \\ 94.28 \\ \\ 95.67 \pm 0.9 \\ \underline{96.89} \\ 96.83 \\ 95.48 \\ \\ 96.44 \pm 0.5 \\ \underline{97.67} \\ 97.48 \\ \end{array}$	$\begin{array}{c} 78.59 \pm 8.2 \\ \underline{93.20} \\ 68.73 \\ 82.56 \\ 88.96 \pm 3.2 \\ \underline{95.61} \\ 92.12 \\ 92.46 \\ \\ 92.30 \pm 1.8 \\ \underline{95.92} \\ 94.83 \end{array}$	$\begin{array}{c} 73.74 \pm \! 6.6 \\ \underline{89.81} \\ 72.15 \\ 80.49 \\ 83.75 \pm \! 3.8 \\ \underline{90.10} \\ 87.22 \\ 86.36 \\ 86.80 \pm \! 1.5 \\ 88.95 \\ 88.14 \end{array}$