
Attentive Multi-Channel Molecular Representation in Drug-Target Affinity Prediction

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Abstract

1 Accurate drug-target affinity (DTA) prediction is fundamental to computational
2 drug discovery. Drug-target binding affinity is influenced by multiple factors,
3 including structural conformations, functional groups, and molecular flexibility.
4 However, existing graph neural network (GNN)-based approaches often fail to
5 explicitly capture these fine-grained features, leading to suboptimal performance
6 and limited interpretability. To address these issues, we propose a framework
7 that integrates structure-aware protein embeddings with multi-channel molecular
8 representations across global, scaffold, and local levels. The key innovation lies in
9 a Cross-Channel Attention (CCA) mechanism, which dynamically aligns protein
10 features with molecular channels and assigns adaptive weights, thereby selectively
11 emphasizing binding-relevant information while reducing redundancy. Experi-
12 ments on the Davis dataset demonstrate that our model consistently outperforms
13 strong baseline methods. Beyond performance improvements, the cross-channel
14 attention mechanism also enhances interpretability by highlighting structural and
15 chemical determinants of binding. Overall, this work establishes cross-channel
16 attention as an effective and interpretable paradigm for advancing DTA prediction.

17 1 Introduction

18 Drug-target affinity (DTA) is a crucial task in drug design and development, playing a vital role in
19 drug discovery, candidate screening, and dosage optimization. Traditional experimental approaches
20 such as high-throughput screening (HTS) and surface plasmon resonance (SPR) provide accurate
21 measurements but suffer from high cost, low throughput, and long processing time, making them
22 insufficient for modern large-scale drug development [8]. In recent years, deep learning has emerged
23 as a powerful tool, showing strong ability to model large-scale biomedical data and significantly
24 accelerating progress in DTA prediction.

25 In DTA modeling, the representation of compounds and proteins is a crucial factor affecting model
26 performance. Generally, three mainstream strategies are adopted: one-dimensional sequence repre-
27 sentation (1D), two-dimensional structural graphs (2D), and three-dimensional spatial structures (3D)
28 (Figure 1). At the 1D level, compounds are usually represented by SMILES strings or molecular
29 fingerprints, while proteins are expressed as amino acid sequences encoded through descriptors such
30 as k-mer or PSSM [3]. At the 2D level, compounds are modeled as molecular graphs and proteins as
31 contact maps, capturing topological and spatial dependencies[7]. At the 3D level, with advances such
32 as AlphaFold2, structural conformations and drug-target complexes can be modeled as point clouds,
33 voxel grids, or atom-level graphs to better capture realistic molecular interactions [16].

34 Previous studies have proposed a variety of deep learning-based methods for DTA prediction, which
35 can be broadly divided into three categories: sequence-based models, structure-based models, and
36 hybrid-based models [22]. Sequence-driven models usually rely on SMILES strings for compounds

Representation method	Drug	Target				
1D Sequence-based	SMILES: <chem>O=C(O)CNC(=O)NC1CCCCC1</chem>	Amino acid Sequences: MEMEKEFEQIDKS GSWAAIYQ...SHD				
	Fingerprint: <table><tr><td>0</td><td>1</td><td>1</td><td>1</td><td>...</td><td>0</td></tr></table>			0	1	1
0	1	1	1	...	0	
Structure-based	2D Graph	2D Graph	Contact Map:			
	3D Structure	3D Point Cloud:	3D voxel grid:			

Figure 1: Illustration of compound and protein representations in DTA modeling.

and amino acid sequences for proteins, and employ CNNs, RNNs, or Transformer-based architectures for feature extraction. Representative examples include DeepDTA [24], DeepCDA [1], and AttentionDTA [23], which achieve strong baseline performance but fail to capture spatial structural information. Structure-aware models leverage molecular graphs, protein contact maps, or 3D complex structures to incorporate spatial dependencies. Notable approaches include GSAML-DTA [10], HGRL-DTA [4], and MSGNN-DTA [18]. Although these models offer better interpretability and physical consistency, they are heavily dependent on high-quality structural inputs, which are not always available in real-world scenarios [22]. Moreover, structure-aware models typically impose higher computational costs due to the complexity of processing 3D spatial information and large-scale graph structures. Hybrid models combine the strengths of both paradigms by jointly leveraging sequence and structural information. Representative methods include GraphDTA [14], MGraphDTA [20], ColdDTA [6], and MutualDTA [21], obtaining the balance between performance and efficiency.

Despite recent advances, existing DTA prediction methods still face several challenges as follows:

- **Insufficient feature representation:** Drug–target affinity depends on multiple factors such as conformations, functional groups, and molecular flexibility. However, many GNN-based models fail to capture these fine-grained structural signals, which limits prediction accuracy and generalization.
- **Limitations in feature fusion:** Multi-channel molecular features are often combined via simple concatenation, static weighting, or globally learnable weights. These strategies cannot adaptively assign channel importance for each specific drug–protein pair, and may encounter unstable weight convergence or even yield inferior performance compared to single-channel representations.
- **Lack of interpretability:** Many existing models operate as black-box predictors, offering little biological insight into the determinants of molecular binding, which restricts their utility in practical drug discovery scenarios.

Therefore, to address these challenges, we propose a novel framework for drug–target affinity prediction that integrates GNN-based multi-channel molecular representation with structure-aware protein embedding (Figure 2). On the molecular side, three complementary channels—global topology, scaffold backbone, and local functional groups—are extracted using a pretrained GNN, providing a hierarchical description of chemical features. On the protein side, we employ the pretrained ESM-2 language model to generate structure-aware embeddings that capture contextual and conformational information. To enable effective interaction between drug and protein features, we introduce a Cross-Channel Attention mechanism that adaptively balances the contributions of different molecular channels, thereby enhancing both predictive accuracy and interpretability.

In summary, our main contributions are as follows:

- We propose a drug–target affinity prediction model that incorporates **multi-channel molecular representations**, enabling a more comprehensive capture of global, scaffold, and local chemical features.
- We design a **Cross-Channel Attention (CCA) mechanism** that adaptively weights molecular channels against protein embeddings, effectively leveraging complementary information while mitigating redundancy.
- We conduct extensive experiments, including ablation studies and case analyses, which demonstrate that the proposed model not only outperforms mainstream baselines but also yields biologically meaningful interpretability under different binding modes.

2 Methodology

In this section, we introduce the proposed framework for DTA prediction. We first present the overall architecture of our proposed framework. We then describe the molecular and protein representation modules in detail, followed by the cross-channel attention mechanism, which enables adaptive integration of multi-channel molecular features.

2.1 Model Architecture

The proposed framework consists of three main components: a molecular feature extraction module, a protein feature extraction module, and a cross-channel attention fusion module. Concretely, a pretrained protein language model is employed to encode protein sequences into structure-aware embeddings. On the molecular side, we adopt the multi-channel representation scheme introduced in MolMCL [17], where graph neural networks generate three complementary embeddings capturing global, scaffold, and local features. Building on these representations, our key contribution is the cross-channel attention module, where protein embeddings serve as guiding signals to dynamically adjust the importance of different molecular channels. The fused representation is then passed through a multilayer perceptron to predict drug–target affinity. This design enables effective utilization of structural information and enhances interpretability, as illustrated in Figure 2.

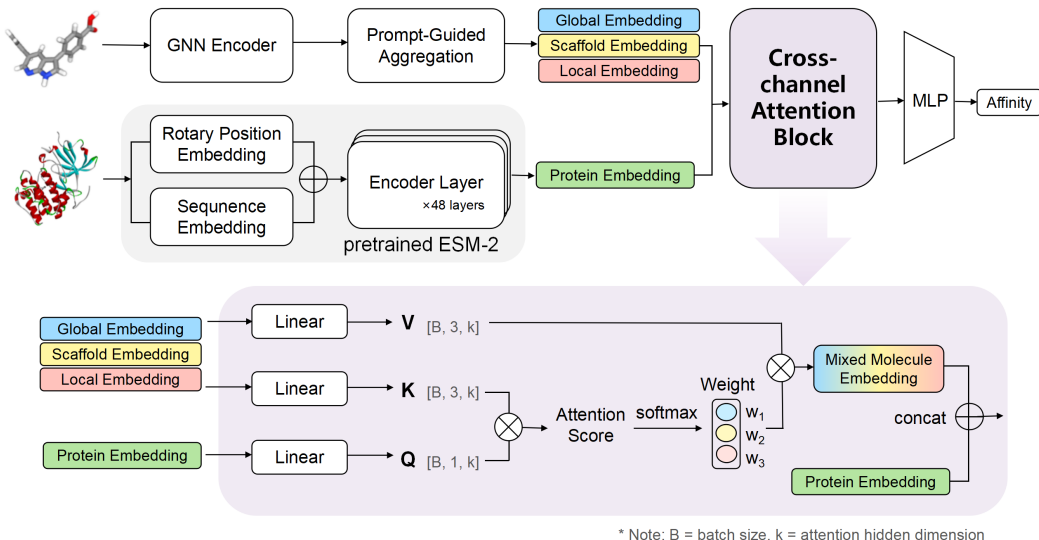


Figure 2: Overall framework. Molecular graphs are encoded by a single GNN and, via prompt-guided aggregation, pooled into three channel embeddings (global, scaffold, local) following MolMCL [17]. Protein sequences are encoded by pretrained ESM-2 [11]. Our key contribution is the **Cross-Channel Attention Block**, which uses the protein embedding to compute softmax weights over the three channels and fuse them for affinity prediction.

98 2.2 Molecular Representation

99 We adopt the MoLMCL framework [17] to construct hierarchical molecular embeddings from three
 100 complementary perspectives: global, scaffold, and local. Concretely, molecular SMILES are first
 101 converted into graphs and encoded by a GNN. Within each channel, a Prompt-Guided Aggregation
 102 module introduces a learnable prompt token to guide attention-based pooling over node features,
 103 producing a channel-specific graph embedding. Through this process, we obtain three embeddings
 104 that correspond to global topology, scaffold backbone, and local functional groups. Each embedding
 105 is channel with a dedicated self-supervised objective to capture multi-level structural information, as
 106 detailed in the following subsections.

107 **Global Channel: Molecule Contrastive Distancing (MCD).** The goal is to learn embeddings
 108 that preserve the overall molecular topology. Training for this channel is conducted through triplet
 109 contrastive learning, where each batch constructs {anchor, positive, negative} triplets. The anchor a
 110 is the original molecule, the positive p is generated by applying subgraph masking to a following the
 111 strategy proposed by MolCLR [19], and the negative n is randomly sampled from other molecules in
 112 the batch. To improve sensitivity to structural similarity, an adaptive-margin triplet loss is employed:

$$\mathcal{L}_{MCD} = \max(0, \alpha_{MCD} + d(a, p) - d(a, n)), \quad (1)$$

113 where $d(\cdot, \cdot)$ denotes the embedding distance between two molecules. The adaptive margin α_{MCD} is
 114 defined as:

$$\alpha_{MCD} = \alpha_{\text{offset}} \cdot \left(1 - \text{sim}_{\text{Tanimoto}}\left(\text{FP}_{\text{mol}}^{(a)}, \text{FP}_{\text{mol}}^{(n)}\right)\right), \quad (2)$$

115 where α_{offset} is a hyper-parameter controlling the margin scale, $\text{sim}_{\text{Tanimoto}}(\cdot, \cdot)$ is the Tanimoto
 116 similarity [2], and $\text{FP}_{\text{mol}}^{(a)}$ and $\text{FP}_{\text{mol}}^{(n)}$ are molecular fingerprints of the anchor and negative molecules,
 117 respectively.

118 **Scaffold Channel: Scaffold Contrastive Distancing (SCD).** This channel focuses on backbone-
 119 level invariance. The anchor a is the original molecule, the positive p is generated by applying
 120 scaffold-invariant perturbations using CReM (an open-source framework for chemically reasonable
 121 mutations), and the negative n is sampled from the batch. A similar adaptive-margin triplet loss is
 122 applied:

$$\mathcal{L}_{SCD} = \max(0, \alpha_{SCD} + d(a, p) - d(a, n)), \quad (3)$$

$$\alpha_{SCD} = \alpha_{\text{offset}} \cdot \left(1 - \text{sim}_{\text{Tanimoto}}\left(\text{FP}_{\text{scaff}}^{(a)}, \text{FP}_{\text{scaff}}^{(n)}\right)\right), \quad (4)$$

124 where $\text{FP}_{\text{scaff}}^{(a)}$ and $\text{FP}_{\text{scaff}}^{(n)}$ represent scaffold-level fingerprints of the anchor and negative molecules.

125 **Local Channel: Context Prediction (CP).** The local channel enhances context understanding by
 126 jointly modeling subgraph-level structures and functional group descriptors through a multi-task
 127 learning scheme with two objectives:

- 128 • *Masked subgraph prediction:* A random atom and its 1-hop neighbors are masked, and
 129 the model predicts the missing features as a multi-label classification task optimized with
 130 cross-entropy loss:

$$\mathcal{L}_{\text{mask}} = - \sum_{i=1}^C y_i \log p_i, \quad (5)$$

131 where C is the number of classes, p_i is the predicted probability for class i , and $y_i \in \{0, 1\}$
 132 is the ground-truth label.

- 133 • *Functional group prediction:* Each molecule is represented by an 86-dimensional normalized
 134 functional group descriptor, and the task is formulated as regression with Smooth L1 loss:

$$\mathcal{L}_{FG} = \frac{1}{d} \sum_{i=1}^d \begin{cases} 0.5(\hat{y}_i - y_i)^2, & \text{if } |\hat{y}_i - y_i| < 1, \\ |\hat{y}_i - y_i| - 0.5, & \text{otherwise,} \end{cases} \quad (6)$$

135 where y_i and $\hat{y}_i$ denote the ground-truth and predicted descriptor values of the i -th functional
 136 group.

137 The total context prediction loss is defined as:

$$\mathcal{L}_{CP} = \mathcal{L}_{mask} + \mathcal{L}_{FG}. \quad (7)$$

138 **Prompt-Guided Aggregation.** In each channel, a learnable prompt token guides a multi-head
 139 attention pooling over atom-level features: the prompt serves as the query, while node embeddings act
 140 as keys and values. This prompt-guided pooling enforces channel-wise structural focus and replaces
 141 uniform pooling with selective, structure-aware aggregation. Prompt-Guided Aggregation operates
 142 during channel encoding and is optimized jointly with the corresponding channel objective in the
 143 pretraining stage, yielding a unified yet channel-aware molecular representation for downstream
 144 prediction.

145 2.3 Protein Representation

146 We employ the ESM-2 model [11], a state-of-the-art protein language model, to encode protein
 147 sequences into structure-aware embeddings. ESM-2 was pretrained on 98M UniRef50 protein
 148 sequences, covering diverse evolutionary and functional contexts, which enables it to capture rich
 149 contextual and structural information from primary amino acid sequences.

150 The model adopts a masked language modeling objective: around 15% of residues are randomly
 151 masked or replaced, and the model is trained to recover the original amino acids. This task drives ESM-
 152 2 to learn rich contextual dependencies across residues, capturing long-range sequence relationships.
 153 In addition, embeddings from upper layers of ESM-2 have been shown to align closely with protein
 154 contact map, demonstrating that the model implicitly encodes structural features despite being trained
 155 without explicit 3D supervision.

156 In our work, we utilize the final-layer embeddings (layer 36) from ESM-2, where residue-level
 157 representations are averaged to form a global protein embedding. These embeddings are particularly
 158 suitable for our multi-channel framework, as they simultaneously encode complementary information
 159 at different levels: global sequence context, structural backbone organization, and local binding-
 160 pocket features. Such hierarchical protein information aligns naturally with the molecule-side
 161 multi-channel design, enabling effective cross-channel attention and more accurate affinity prediction.

162 2.3.1 Cross-Channel Attention Mechanism(CCA)

163 In the preliminary design, the fusion of the three molecular channels relied on fixed weights or static
 164 weighted summation. Such approaches lack sensitivity to protein features and cannot dynamically
 165 adapt to different protein-molecule pairs. To address these limitations, we introduce a Cross-Channel
 166 Attention mechanism that enables more flexible and context-aware integration of representations
 167 (Figure 2).

168 Specifically, the protein embedding is first projected into a query vector $Q \in \mathbb{R}^{B \times 1 \times k}$, while the
 169 three molecular channel embeddings are projected into key and value vectors $K \in \mathbb{R}^{B \times 3 \times k}$ and
 170 $V \in \mathbb{R}^{B \times 3 \times k}$, respectively, where B is the batch size and k represents the hidden dimension of the
 171 attention space. The attention scores are then computed using the scaled dot-product attention:

$$\text{Attention}(Q, K, V) = \text{softmax} \left(\frac{QK^T}{\sqrt{d_k}} \right) V \quad (8)$$

172 where d_k represents the dimensionality of the key vectors, used to scale the dot product and prevent
 173 the attention scores from becoming excessively large. The softmax operation produces dynamic
 174 weights $\{w_1, w_2, w_3\}$ for the three channels, which are used to compute a weighted sum of molecular
 175 embeddings and form the Mixed Molecule Embedding. This fused embedding is subsequently
 176 concatenated with the protein embedding and passed into an MLP predictor for affinity regression.

177 The CCA mechanism offers several advantages compared to other fusion methods:

- 178 • **Adaptive and stable fusion:** Attention weights are computed dynamically for each protein-
 179 molecule pair, enabling the model to adjust the fusion strategy in a context-specific manner.
 180 Unlike approaches that assign a fixed learnable scalar weight to each channel, which may
 181 lead to unstable or biased convergence, this dynamic weighting ensures more reliable and
 182 consistent training behavior.

- **Context-guided alignment:** Protein embeddings guide the computation of fusion weights, enabling the model to capture dependencies between protein context and molecular structural levels.

3 Experiments

In this section, we evaluate the effectiveness of our proposed model on the Davis dataset and compare it with representative baselines. We further conduct ablation studies and case analyses to validate the contribution of each module and the interpretability of our framework.

3.1 Experimental Setup

Objective. The primary goal of our experiments is to assess the predictive performance of the proposed model on DTA prediction tasks and benchmark it against state-of-the-art baselines.

Evaluation Metrics. We adopt three widely used regression metrics. Let y_i denote the ground-truth affinity value, $\hat{y}_i$ the predicted value, and N the number of samples.

(1) Mean Squared Error (MSE). MSE evaluates the absolute prediction accuracy by measuring the average squared difference between predictions and true values:

$$\text{MSE} = \frac{1}{N} \sum_{i=1}^N (y_i - \hat{y}_i)^2. \quad (9)$$

(2) Concordance Index (CI). CI measures the consistency of ranking between predicted and true affinities. For all comparable pairs (i, j) where $y_i > y_j$, CI is defined as:

$$\text{CI} = \frac{1}{Z} \sum_{y_i > y_j} h(\hat{y}_i - \hat{y}_j), \quad (10)$$

where Z is the total number of such pairs and

$$h(x) = \begin{cases} 1 & x > 0, \\ 0.5 & x = 0, \\ 0 & x < 0. \end{cases}$$

(3) Coefficient of Determination (R^2). R^2 assesses the proportion of variance in the ground-truth values explained by the model:

$$R^2 = 1 - \frac{\sum_{i=1}^N (y_i - \hat{y}_i)^2}{\sum_{i=1}^N (y_i - \bar{y})^2}, \quad (11)$$

where $\bar{y}$ is the mean of the ground-truth values.

3.2 Dataset

We conduct experiments on the Davis dataset, which is a widely adopted benchmark for DTA prediction [5]. The dataset consists of 68 kinase inhibitors and 442 protein kinases, resulting in 30,056 interaction pairs with experimentally measured binding affinities (K_d values). These values are provided in log-transformed form, making them suitable for regression-based modeling. The Davis dataset is characterized by its large coverage, containing more than 30k compound-protein pairs, as well as its continuous affinity labels that enable quantitative evaluation.

Split strategy. Following common practice, the dataset is split into 80% training, 10% validation, and 10% testing, controlled by a fixed random seed to ensure reproducibility.

Table 1: Performance comparison on the Davis dataset. The best results are in bold.

Model	MSE ↓	CI ↑	R^2 ↑
KronRLS	0.379	0.871	0.407
DeepDTA	0.261	0.878	0.630
GraphDTA	0.229	0.893	0.670
ColdDTA	0.250	0.884	0.652
NTMFF-DTA	0.237	0.896	0.684
Ours	0.221	0.884	0.732

3.3 Baselines

We compare our proposed model with six representative baseline methods, covering traditional kernel-based approaches, early deep learning models, and the latest multi-scale frameworks:

- **KronRLS** [15]: a classical kernelized regression method that models drug–target pairs via the Kronecker product of drug kernels and protein kernels. Despite its simplicity, KronRLS remains a strong traditional baseline due to its efficiency and effectiveness on small-scale datasets.
- **DeepDTA** [24]: the first end-to-end deep learning framework for DTA prediction. It encodes drug SMILES and protein amino acid sequences separately with convolutional neural networks (CNNs) and learns their joint interactions through fully connected layers.
- **GraphDTA** [14]: extends DeepDTA by representing drugs as molecular graphs instead of SMILES strings, thereby capturing richer structural information with graph neural networks (GNNs). Proteins are still modeled using CNNs over their sequences.
- **ColdDTA** [6]: designed to address the cold-start problem where unseen drugs or proteins appear in the test set. It introduces a cold-start aware loss function that forces the model to generalize better across novel compounds and targets.
- **NTMFF-DTA** [12]: a recent state-of-the-art model that incorporates neural temporal memory modules and multi-scale feature fusion. This design enables the model to capture both local and global dependencies in drug and protein features.

3.4 Implementation Details

Our model is implemented in PyTorch 2.1.2 with CUDA 11.8 and trained on a single NVIDIA RTX 3090 GPU (24GB). We adopt the Adam optimizer with a cosine learning rate scheduler and warm-up strategy, and train for up to 100 epochs with early stopping to prevent overfitting.

3.5 Results

We evaluate our model against five representative baselines on the Davis dataset, and the results are summarized in Table 1. Traditional kernel methods such as KronRLS achieve reasonable performance but are limited by shallow feature representations. DeepDTA improves substantially by introducing CNN encoders for SMILES and protein sequences, while GraphDTA further reduces error by leveraging molecular graph structures. ColdDTA enhances generalization in cold-start scenarios with specialized loss design, and more recent approaches such as NTMFF-DTA demonstrate the benefits of incorporating attention mechanisms and multi-scale modeling. Compared with these baselines, our model achieves the lowest MSE (0.221) and the highest R^2 (0.732), indicating superior regression accuracy and stronger explanatory power. Although NTMFF-DTA attains a slightly higher CI, our approach strikes a better balance between error minimization and predictive stability. These results highlight the effectiveness of combining hierarchical molecular representations, protein language embeddings, and cross-channel attention for prediction.

3.6 Ablation Studies

To investigate the contribution of each component in our proposed model to DTA prediction, we conducted a series of ablation experiments. Specifically, we examined the effect of different molecular

Table 2: Ablation study results on the Davis dataset.

Model Variant	MSE	CI	R ²
Single-channel (CP)	0.238	0.881	0.711
Single-channel (SCD)	0.284	0.868	0.655
Single-channel (MCD)	0.277	0.869	0.664
Mean weight	0.251	0.880	0.695
Full model	0.214	0.886	0.740

channel settings by removing or simplifying the cross-channel fusion mechanism, while keeping the other parts of the architecture unchanged. The experimental settings follow the same data splitting strategy as in the main experiments, and all results were obtained with random seed fixed at the same value for reproducibility.

Regarding the molecular channel representation, we evaluated three reduced configurations: (i) **Single-channel (MCD/SCD/CP)**, where only one type of molecular representation was preserved and the other channels were discarded; (ii) **Mean weight**, where all three molecular channels were averaged with equal weights, discarding the dynamic weighting mechanism; and (iii) **Full model**, where all channels were fused via cross-channel attention (results provided in Table 2).

As shown in Table 2, the full model achieves the best performance across all metrics, demonstrating the effectiveness of integrating complementary information from multiple channels with adaptive attention. In contrast, the single-channel variants exhibited substantial performance degradation, with the CP-only variant performing relatively better than MCD-only and SCD-only settings. This result indicates that the CP channel preserves more fine-grained chemical property features that are directly related to ligand-protein binding, thereby outperforming other single-channel variants. Mean pooling achieved only moderate results, performing worse than the CP-only variant, since although it integrates features from all three channels, it fails to adaptively emphasize the most informative chemical property features and thus cannot fully leverage their complementarity. Overall, these results verify that both multi-channel representation and cross-channel attention are critical to the predictive power of the model.

3.6.1 Case Study and Model Interpretability

To further validate the effectiveness of multi-channel fusion and cross-channel attention in practical tasks, we selected two representative protein-ligand complexes with distinct conformational states of the Abl tyrosine kinase. The chosen complexes are the Abl tyrosine kinase structures with PDB IDs **3UE4** and **4XEY**, which exhibit substantially different ligand binding patterns. The visualization results are shown in Figure 3, where we analyze how distinct binding patterns influence the attention weights allocation across molecular channels, demonstrating the interpretability of the model.

Figure 3(a–c) illustrates the complex structure of 3UE4, originally reported by Levinson and Boxer [9], which depicts the Abl kinase domain bound to the ATP-competitive inhibitor bosutinib. Bosutinib inserts deeply into the conserved ATP-binding pocket, where its rigid backbone forms extensive hydrophobic contacts with residues such as Leu248A, Phe317A, and Met318A, ensuring a stable geometric fit within the cavity. The interaction pattern is dominated by scaffold-driven hydrophobic embedding, while only a limited number of hydrogen bonds (e.g., with Met318A) contribute modestly to the stabilization of the complex. This binding mode is faithfully captured by our model (Figure 3c), which assigns high weight to the *scaffold* channel and minimal weight to the *local* channel. This observation highlights the ability of the model to recognize scaffold-driven binding mechanisms and to adapt its attention toward structural features underpinning ligand affinity.

In contrast, Figure 3(d–f) presents the 4XEY structure, originally reported by Lorenz et al. [13], which reveals the Abl SH2-kinase domain in complex with dasatinib. Dasatinib binds in a mode distinct from bosutinib, where local functional groups rather than the rigid scaffold play the dominant role. The ligand inserts parallel to the protein surface, forming multiple hydrogen bonds with residues such as Met337A, Thr334A, and Met309A, while also engaging in hydrophobic contacts that provide additional stabilization. The interaction pattern is thus driven primarily by fine-grained chemical group recognition, with hydrogen bonds serving as key determinants of binding specificity. This

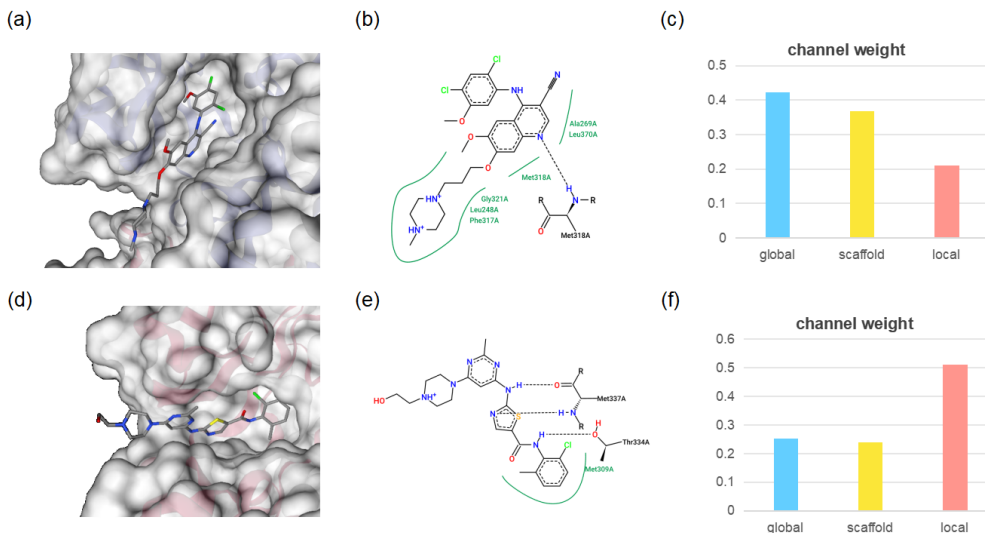


Figure 3: Case study of model interpretability on 3UE4 and 4XEY. (a–c) 3UE4 complex: (a) 3D binding pose, (b) 2D interaction diagram, (c) channel weight distribution. (d–f) 4XEY complex: (d) 3D binding pose, (e) 2D interaction diagram, (f) channel weight distribution.

binding mode is faithfully captured by our model (Figure 3f), which assigns dominant weight to the *local* channel and only minor contributions to the *scaffold* and *global* channels.

Together, these two cases demonstrate the capacity of our model to dynamically adjust attention across channels depending on the binding mode. In 3UE4, binding is scaffold-dominated and attention is focused on the *scaffold* channel, whereas in 4XEY, functional group interactions dominate and attention shifts to the *local* channel. This adaptive allocation not only confirms the rationality of the multi-channel design but also illustrates its structural interpretability. By dynamically allocating attention across multi-channel molecular representations, the framework provides mechanistic insights and enhances trustworthiness, addressing the common criticism of deep learning methods for their limited interpretability.

4 Conclusion

In this work, we presented an attentive multi-channel framework for DTA prediction that integrates structure-aware protein embeddings with multi-channel molecular representations. By employing complementary channels for global topology, scaffold backbone, and local functional groups, and fusing them through a cross-channel attention mechanism, our model effectively captures multi-level structural and chemical information. Extensive experiments on the Davis dataset demonstrated that our method outperforms representative baselines, while ablation studies and case analyses confirmed the importance of both multi-channel representation and adaptive attention for accuracy and interpretability. Beyond predictive performance, the framework provides mechanistic insights into distinct binding modes, enhancing the reliability of computational DTA modeling. These findings suggest that attentive multi-channel learning offers a promising direction for advancing interpretable and generalizable approaches in computational drug discovery.

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