Video Representation Learning of Cardiac MRI for Genetic Discovery

Matt Sooknah Calico Life Sciences mds@calicolabs.com Sivaramakrishnan Sankarapandian Calico Life Sciences sivark@calicolabs.com

Ramprakash Srinivasan Calico Life Sciences rams@calicolabs.com

Johannes Riegler Calico Life Sciences riegler@calicolabs.com

Jun Xu Calico Life Sciences junxu@calicolabs.com

Abstract

In recent years, many studies have utilized cardiac magnetic resonance imaging (cMRI) to define image-derived phenotypes (IDPs) relating to heart structure and function for genome-wide association studies (GWAS). These IDPs are traditionally defined manually from volume, strain, and geometric parameters. Here we introduce an unsupervised learning approach that extracts spatiotemporal representations from cMRI videos in a large human cohort of ∼68,000 subjects from the UK BioBank. The resulting representations can be used to predict age and manually crafted IDPs accurately. We further use these representations to define IDPs to capture both known and potential novel genetic associations. Our work suggests that unsupervised learning can be used to extract rich, unbiased information from medical videos with applications to genetic discovery.

1 Introduction

Time-series cMRI is a commonly used diagnostic tool that captures heart dynamics in high spatial and temporal resolution. A long-standing goal in cardiovascular research is to define features in cMRI scans that capture cardiovascular functions (like muscle contraction, heart rhythm, and blood flow) which influence cardiac health, then combine those features with genetic data in large human cohorts to identify genes implicated in cardiovascular disease.

Prior works have developed methods to extract manually defined IDPs like ejection fraction [\[2\]](#page-9-0) and strain rate [\[39\]](#page-11-0) from cMRI. Multiple studies have conducted GWAS using these traits, identifying key genetic markers of cardiovascular health [\[29,](#page-10-0) [30,](#page-11-1) [1,](#page-9-1) [22\]](#page-10-1). These IDPs are easily defined and interpretable, but only provide a narrow view of the underlying complex structure of the heart. Others have predicted patients' "cardiovascular age" using pre-defined cMRI measurements, and then compared it to chronological age to obtain a "delta-age" phenotype [\[33\]](#page-11-2). Compared to manually defined IDPs, "delta-age" captures many signals into a composite measure of cardiovascular health, but still doesn't capture all the underlying biological variation visible in cMRI.

More recent studies have used unsupervised learning to extract novel phenotypes from imaging in large human cohorts, e.g. brain and cardiac MRI [\[27,](#page-10-2) [6,](#page-9-2) [26\]](#page-10-3). In these cases, a self-supervised model was trained on organ images or 3D meshes, then individual latent feature activations (e.g. from an autoencoder bottleneck) were used as novel IDPs for GWAS. These IDPs can capture subtle signals that supervised methods miss, but no principled ways were proposed to turn the representations into IDPs. In the cardiac MRI case, where we have videos rather than static images, existing

representation-based IDPs also do not consider temporal dynamics or strain maps, which provide useful signals for characterizing cardiovascular disease.

Here, we apply unsupervised representation learning on time-series cardiac MRI from the UK BioBank (N∼68,000 subjects) using video masked auto-encoders [\[42\]](#page-11-3). We train models on multiple types of cMRI videos as well as their deformation fields (similar to optical flow maps), then fuse the resulting representations to maximally capture the spatiotemporal dynamics. We assess the utility of these representations using prediction of age and cardiac indices as a benchmark. We cluster the representations of each subject and identify clusters enriched for cardiovascular disease. Finally, we derive IDPs using delta-age and subject-to-cluster similarity, then perform GWAS, revealing genes associated with cardiovascular health, including both established and potentially novel hits. The overall approach is outlined in Figure [1.](#page-1-0)

2 Spatiotemporal Representation Learning on Cardiac Videos

Figure 1: Overview of spatiotemporal modeling for genetic discovery. (A) We obtain cMRI videos for ∼68,000 subjects and calculate deformation fields corresponding to heart motion. We train a VideoMAE model (encoder E, decoder D), then unmasked videos are passed to the frozen encoder to obtain spatiotemporal patch representations, which are average-pooled to get a single video-level representation *Z*. We can augment the representation by combining latent features from multiple input signals (e.g. images plus deformation fields). (B) We use the representations for multiple downstream tasks - predicting age and cardiac function, clustering subjects (which reveals subpopulations associated with cardiovascular disease risk), and extracting representation-derived phenotypes for GWAS (which could enable the discovery of novel genetic targets).

2.1 Cardiac MRI Videos

We use data from the UK BioBank, a large prospective cohort with genotyping, EHR info, and deep phenotyping of 500k subjects from the UK [\[7\]](#page-9-3). A subset were imaged using multiple modalities including cardiac MRI [\[28\]](#page-10-4); at this time roughly ∼68k subjects have cMRI imaging available. Several types of cMRI imaging were collected; in this study we use the long axis 4-chamber (LA) and short axis (SA) scans. The LA-4ch scans capture a 2D video of a slice of the heart that includes a crosssection of all 4 chambers (left and right ventricles, left and right atria). The SA scans capture a 3D volumetric video that fully covers the left and right ventricles, but excludes the atria. These two scans have orthogonal imaging planes and different trade-offs, with the LA scans covering a larger field of view (FOV) but only at a 2D slice, while the SA scans are pseudo-3D but have a smaller FOV [\[11\]](#page-9-4). In our experiments, we assess the value of representations derived from both scan types. All videos contain a fixed number of frames corresponding to a single heartbeat, with the first frame representing the end-diastole (ED) phase corresponding to maximum filling of the ventricles.

2.2 Temporal Deformation Fields

To identify salient motion features, we generated temporal deformation maps for each video as an additional input to our representation models, inspired by two-stream models that incorporate optical flow signals for video recognition [\[36\]](#page-11-4). The deformation maps are obtained by registering each frame of the video to the first frame (end-diastole), using VoxelMorph [\[3\]](#page-9-5).

Formally, let $v_{i,t} \in \mathcal{R}^{H \times W}$ represent frame *t* of video *i*. We estimate a deformation field that aligns frame 0 to frame *t* via a spatial transformer module [\[14\]](#page-10-5). The deformation field consists of a displacement vector $(\Delta x, \Delta y)$ at each pixel coordinate. We take the L2 norm of the displacement vector at each coordinate to flatten the deformation field to a 2D "magnitude image" $d_{i,t} \in \mathcal{R}^{H \times W}$.

To predict the deformation fields, we first trained a VoxelMorph U-Net model following the general strategy and hyperparameters used in [\[3\]](#page-9-5). As input, we randomly sample pairs of video frames $(v_{i,0}, v_{i,t})$ and train the model to minimize mean-squared-error with a L2 regularization term on the deformation field magnitude. Given the trained model, we run inference on the entire cohort. At inference time, we compute the deformation magnitude $d_{i,t}$ for each video frame t , then concatenate into a single video d_i with the same dimensions as the original video of pixel intensities v_i . This deformation video captures dynamics in heart morphology that complement the original pixel intensities.

2.3 Self-Supervised Video Representation Learning

To extract a low-dimensional representation that captures spatiotemporal dynamics in the videos and deformation fields, we train a self-supervised model based on VideoMAEv2, which has shown excellent performance as a pre-training task for downstream video classification [\[42\]](#page-11-3). The model tokenizes a video of shape $H \times W \times T$ into a sequence of N spatiotemporal patch tokens of shape $H' \times W' \times T'$, embeds them into latent vectors of dimension *D*, then passes them through a sequence of vision transformer layers. During training, some tokens are randomly masked (using tube masking to mask the same patches across time), then a decoder learns to reconstruct the masked video tokens by minimising mean-squared-error loss. During inference, we pass the full (unmasked) video v_i through the encoder to obtain a sequence of spatiotemporal patch embeddings $z_{i,1}, \ldots z_{i,N}$ which are average-pooled to get a single video-level embedding $z_i \in \mathcal{R}^D$.

We train four VideoMAEv2 models, one each for long axis videos, long axis deformation fields, short axis videos, and short axis deformation fields. We then run inference to obtain four sets of embeddings corresponding to each of the four modalities. Since the model is designed for 2D videos, the short axis models are trained on randomly sampled 2D axial slices from underlying 3D volumes, and the inference is run on all z-slices followed by average-pooling so we always get an output embedding of the same dimension *D*. We can further combine information from multiple representations by concatenating their latent dimensions. For example, we can combine the embeddings of videos and deformation fields from the same modality, or combine across modalities (short and long axis). Note that we use the same datasets for training and inference, since the reconstruction objective using in training is not directly connected to any downstream evaluation tasks; this is a common setup in representation-based target discovery [\[6\]](#page-9-2).

2.4 Benchmarking Video Representations

To evaluate the predictive power of the learned embeddings, we construct a benchmark task consisting of predicting age and cardiac function parameters via linear regression. Age is a good benchmark due to its strong association with cardiovascular health [\[23\]](#page-10-6). Cardiac parameters measured from cMRI are also strong predictors of heart function; for example, left ventricular ejection fraction (LVEF) is used in the diagnosis of heart failure with reduced ejection fraction (HFrEF). Critically, we choose cardiac parameters that are calculated from image features across multiple timepoints of the cardiac cycle. For example, ejection fractions (EF) are calculated by comparing the maximum and minimium volumes over time for a particular cardiac chamber. Likewise strain parameters like circumferential strain (CS) and longitudinal strain (LS) assess the contractility of the myocardial wall over time

across different axes. Thus, we hypothesize that models which can better capture spatiotemporal dynamics will do better at predicting these parameters.

3 Deriving Phenotypes from Representations

To test the utility of our embeddings for target discovery, we explored several ways to extract phenotypes from learned representations, which we term RDPs (representation-derived phenotypes).

3.1 Delta Age

First, we compute cardiovascular delta-age in a manner similar to previous work [\[33\]](#page-11-2), but using video embeddings rather than manually-crafted IDPs as the input variables. In brief, we first train a linear model to obtain weights w and bias b that predict subjects' chronological age from their embedding:

$$
PredictedAge_i = \boldsymbol{w}^T \boldsymbol{z_i} + b
$$

We then subtract predicted from chronological age to get a raw delta-age. This raw delta-age value is still correlated with age, so we perform a correction by linear regressing true age from raw delta-age to obtain weight α and bias β , then subtract this prediction to get an unbiased delta-age phenotype:

$$
DeltaAge_i = PredictedAge_i - (\alpha \cdot Chromological Age_i + \beta)
$$

This delta-age phenotype captures the extent to which a subject's predicted age differs from their chronological age, based on features from our spatiotemporal embeddings. Similar formulations of delta-age in the literature have been linked to a variety of health outcomes [\[33,](#page-11-2) [37\]](#page-11-5).

3.2 Clustering

Second, we cluster the subjects based on their embeddings to identify subpopulations with different heart characteristics. We hypothesize that some of these clusters will be enriched (either positively or negatively) for cardiovascular diseases. Instead of using a subject's cluster label as a categorical phenotype (which does not account for the complex hierarchical relationships between clusters and subjects), we derive a continuous phenotype based on subject-to-cluster similarity.

Formally, given an assignment of each subject *i* to a cluster C_k (where $k \in \{1...K\}$), we compute the mean embedding of each cluster:

$$
\boldsymbol{c}_{\boldsymbol{k}}=\frac{1}{|\mathcal{C}_k|}\sum_{i\in\mathcal{C}_k}\boldsymbol{z_i}
$$

then compute the cosine similarity of each subject-level embedding to the cluster means, resulting in *K* phenotypes representing similarity of each subject *i* to each cluster *k*:

$$
Cluster Similarity_{k,i} = CosineSimilarity(c_k, z_i)
$$

This gives us *K* independent phenotypes, each one representing similarity to a prototypical heart embedding of a particular subpopulation inferred from clustering.

4 Experiments

4.1 Dataset Preprocessing

From the raw LA and SA videos, we first segmented the left + right ventricles and atria using a segmentation model described previously [\[2\]](#page-9-0). We then resampled the videos to a consistent spatial

resolution, cropped them to a fixed pixel dimension and normalized the pixel intensities to the range [0, 1]. We then compute temporal deformation maps for both LA and SA scans (as described in Section [2.2\)](#page-2-0), by training a VoxelMorph model with default settings [\[3\]](#page-9-5) on a subset of 512 subjects, then inferring on the whole cohort. Finally, we crop the videos and deformation maps using the bounding box of the segmentation mask, resize to 128x128 pixels, and mask the videos to the foreground pixels to emphasize the signal within the heart substructures. The temporal dimension is always 50 frames representing one full cardiac cycle, and is not altered. After postprocessing, we have 64,293 subjects with LA scans and 63,562 with SA scans, but for benchmarking tasks we only use the intersecting subset to maintain a fixed sample size.

4.2 Predicting Age and Cardiac Parameters

We benchmark and compare embeddings by linear probing on biologically meaningful parameters patient age (which correlates strongly with cardiovascular health) and cardiac function indices such as ejection fractions and strain rates (which relate to cardiac dynamics). We obtain these parameters from published UKBB data fields. For age, use the patient age at the imaging visit, which is available for all 63,562 subjects in our benchmark cohort. For cardiac function parameters, we use published values [\[2\]](#page-9-0), which are only available for 34,929 subjects in our cohort. We split the subjects into 75% training and 25% test. Note that the size of the training and test sets is larger for age prediction than cardiac parameter prediction, due to the differing number of available labels. For each target variable and input embedding, we train a ridge regression model using 5-fold cross validation (trying regularization strengths between $1...10⁴$) to predict the parameter on the train set. The resulting models are run on the test sets and the R^2 values compared across embedding types.

Table 1: Evaluation of age and cardiac parameter prediction $(R^2$ on the test set) across input data modalities using VideoMAEv2. "SA/LA Concat" denotes combining video and deformation fields within short / long axis views respectively; "All Concat" denotes combining all 4 inputs. LVEF = left ventricular ejection fraction, RVEF = right ventricular ejection fraction, LAEF = left atrial ejection fraction, RAEF = right atrial ejection fraction, LVCO = left ventricular cardiac output, LVRS = left ventricular radial strain, LVCS = left ventricular circumferential strain, LVLS = left ventricular longitudinal strain.

Embedding Inputs	Age	LVEF	RVEF	LAEF	RAEF	LVCO	LVRS	LVCS	LVLS
LA Video	0.648	0.574	0.586	0.673	0.691	0.583	0.618	0.704	0.669
LA Deform	0.632	0.557	0.553	0.641	0.659	0.643	0.596	0.683	0.663
SA Video	0.713	0.822	0.795	0.484	0.388	0.845	0.783	0.892	0.497
SA Deform	0.698	0.745	0.738	0.475	0.404	0.810	0.767	0.863	0.487
LA Concat	0.681	0.583	0.593	0.678	0.705	0.666	0.630	0.712	0.711
SA Concat	0.750	0.835	0.820	0.504	0.422	0.868	0.812	0.904	0.515
All Concat	0.773	0.827	0.810	0.692	0.709	0.867	0.810	0.900	0.732

In Table [1,](#page-4-0) we evaluate representations from each modality (long axis, short axis), signal type (image, deformation) as well as hybrid representations from concatenating the latent dimensions. We use a fixed set of hyperparameters based on default settings from the VideoMAEv2 paper: ViT-base model architecture, patch size 8, sampling rate 3, and mask ratio 90%.

We find that the SA-derived representations do better on indices that are traditionally derived from SA scans (LVEF, RVEF, LVCO, LVRS, LVCS) and vice versa for LA (LAEF, RAEF, LVLS). Concatenating raw videos and deformation fields within each modality is always beneficial for performance. Finally, concatenating across views (SA, LA) is only beneficial for some metrics but not others. With this in mind, in our downstream analysis we concatenate the long axis image + deformations into one representation, and likewise for short axis, but do not mix short and long axis views.

4.3 Model Ablations

In Table [2,](#page-5-0) we conduct ablation studies by training multiple VideoMAEv2 models with different hyperparameters, focusing on one input modality (long-axis videos). For comparison, we also

Table 2: Evaluation of age and cardiac parameter prediction $(R^2$ on the test set) across models and hyperparameter settings, using long axis video embeddings as input. I-MAE = Image-MAE, V-MAE = VideoMAE. ResNet50 was trained on ImageNet classification; all other models were trained on cardiac videos in a self-supervised manner. See Table 1 for abbreviations. The row in *italics* denotes the model settings that were used for our other analyses; note that these settings don't correspond to the absolute best performance, but the difference with best-performing settings is marginal.

Model Arch	Model Size	Patch Size	Sample Rate	Mask Ratio	Age	LVEF	LVCO	LVCS	LVLS
ResNet ₅₀	۰				0.352	0.229	0.310	0.346	0.281
I-MAE	base	8	-	75	0.506	0.374	0.494	0.495	0.446
V-MAE _{v2}	small	8	3	90	0.591	0.549	0.498	0.673	0.640
V-MAE _{v2}	base	8	$\overline{2}$	90	0.641	0.571	0.584	0.698	0.669
V-MAE _{v2}	base	8	3	75	0.651	0.575	0.586	0.706	0.679
$V-MAEv2$	base	8	3	90	0.648	0.574	0.583	0.704	0.669
V-MAE _{v2}	base	16	3	90	0.605	0.552	0.484	0.679	0.635
V-MAE _{v2}	base	8	6	90	0.635	0.566	0.572	0.692	0.669
V-MAE _{v2}	base	8	3	95	0.626	0.567	0.576	0.696	0.663
V-MAE _{v2}	large	8	3	90	0.657	0.581	0.600	0.707	0.677

train a conventional ImageMAE model [\[13\]](#page-9-6) on the same inputs, treating each timeframe as an independent image. The ImageMAE uses a comparable architecture to our VideoMAE "base" model (same encoder and decoder dimensions, depth and number of heads). As an additional baseline not trained on cardiac videos, we include ResNet50 [\[12\]](#page-9-7) pretrained on ImageNet classification [\[9\]](#page-9-8), with weights obtained from the *torchvision* package, using the activations of the last layer as an embedding. For these static image models, each 2D frame is encoded separately and the embeddings are average-pooled across time.

We expect the ResNet50 model to extract general texture features, but nothing specific to cardiac MRI (since it was trained on natural images). Conversely, we expect ImageMAE to capture cardiac-specific texture (since it is trained on our dataset), but to lack explicit modeling of temporal dependencies. Accordingly, we see that ResNet50 performs worst, and static MAE performs better but does not meet the performance of any VideoMAE model (even ViT-small) across almost all benchmark tasks.

For VideoMAE models, going from a "small" to "base" ViT architecture yields substantial improvement, while going from "base" to "large" gives marginal improvement at the cost of much longer training time. Likewise increasing patch size from 8 to 16 leads to a performance drop. Sample rate denotes how many video frames are skipped when sampling; reducing from 6 to 3 improves performance slightly, but further reducing to 2 does not. Finally, increasing the mask ratio to 95% slightly harms performance and reducing it to 75% slightly improves performance.

4.4 Clustering Analysis

In Figure [2,](#page-6-0) we visualize both short and long axis embeddings from the whole cohort using UMAP [\[17\]](#page-10-7), overlaying the labels from Leiden clustering [\[40\]](#page-11-6).

Prior to clustering, we first remove confounding effects from the embeddings by regressing out the effects of age, sex and BMI using linear regression, as implemented in the *scanpy* package [\[45\]](#page-11-7). Then we run nearest neighbor connectivity (using cosine distance metric) and Leiden clustering. The resulting number of clusters *K* is determined automatically.

We find that there are some clear outlier clusters in the long axis embeddings, while the short axis clusters are less separated. We hypothesize that the long axis embeddings (which derive from a single cross-sectional slice of both atria and ventricles) may encode more population-level variation than the short axis embeddings (which are averaged over many slices, some of which only cover a small portion of ventricular morphology, and don't cover atria at all). We therefore focus on long axis embeddings in the subsequent analysis.

Figure 2: UMAP and clustering of long axis and short axis embeddings. Long axis shows more separation of outlier clusters, which we subsequently interrogate with PheWAS and GWAS.

4.5 Phenome-Wide Association Studies

We conduct phenome-wide association studies (PheWAS) on the derived RDPs to better understand their associations with traditional phenotypes. For simplicity, we restrict the set of traditional phenotypes to ICD-10 codes reported in the UK BioBank, which roughly correspond to diagnoses of medical conditions. Following the general approach of the *PHESANT* package [\[20\]](#page-10-8), we first preprocess each RDP by regressing out the effect of sex, age, BMI and imaging center, then applying a rank-inverse normal transform so that the RDP follows a normal distribution. Then, for each combination of RDP and ICD-10 code, we perform logistic regression to obtain an effect size and p-value indicating the strength of the association between the RDP and ICD-10 code.

Figure 3: Example volcano plots from ICD-10 PheWAS to identify diseases associated with embedding similarity RDPs for two clusters identified as outliers in the long-axis embeddings (see Figure [2\)](#page-6-0). The RDPs show strong association with cardiovascular disease, suggesting that the representations extract biologically meaningful signal. Dashed line corresponds to Bonferroni-corrected significance threshold.

Figure [3](#page-6-1) shows example PheWAS results from long-axis cluster RDPs (Section [3.2\)](#page-3-0) corresponding to two outlier clusters seen in Figure [2.](#page-6-0) Several clusters are highly enriched for phenotypes related to

cardiometabolic disease, including heart failure, myocardial infarction, atrial fibrillation, coronary atherosclerosis, mitral valve disorders, high cholesterol, high blood pressure, left bundle branch block, angina and type 2 diabetes. Moreover there are distinct enrichments in multiple clusters, suggesting multiple subtypes for diseases like heart failure. Full results are reported in the appendix.

4.6 Genome-Wide Association Studies

We run genome-wide association studies (GWAS) on each RDP to identify genetic associations with latent spatiotemporal features in cMRI videos, using the UKBB genotyping data described in [\[7\]](#page-9-3). We follow the genotype imputation, filtering and QC procedure described in [\[15\]](#page-10-9). We use the GWAS implementation in *regenie* [\[16\]](#page-10-10), adjusting for sex, age, age², imaging center, genotyping array and top 10 genetic principal components. After applying subject-level filtering from [\[15\]](#page-10-9), we are left with *N*=62,605 subjects. We filter variants using simple peak detection with a minimum genomic distance of 30 between neighboring peaks. As this is a proof-of-concept analysis, we do not perform LD-score regression or fine-mapping, but acknowledge their importance for further validation of the genetic associations. We then use variant-to-gene annotations from OpenTargets [\[25\]](#page-10-11) to identify the most likely gene associated with each variant. We report all hits with a p-value below the base genome-wide significance threshold for the number of SNPs being tested. Since we are testing multiple phenotypes, we also report a Bonferroni-adjusted p-value threshold, but note that this may be overly conservative given potential correlations between phenotypes.

Figure 4: Manhattan plots of delta-age and cluster-mean similarities for 3 example clusters derived from long axis embeddings. For clarity, we only label peak hits for each gene where $-log_{10}(p) >= 9$. Dashed line corresponds to genome-wide significance threshold.

Here, we focus on results from the long-axis RDPs (based on combined video + deformation embeddings). A larger list of hits from both long and short axis phenotypes are listed in the appendix. Some example Manhattan plots are shown in Figure [4.](#page-7-0) In total we find 146 significant genes using the base genome-wide significance threshold of $p \le 5.0 \times 10^{-8}$. However, note that we are testing 18 phenotypes (1 delta-age + 17 clustering); with the Bonferroni-adjusted value of $p \le 2.8 \times 10^{-9}$, there are 74 significant genes. We examine some of the more significant and interesting hits below.

4.6.1 GWAS Results for Delta Age

Several top delta-age hits are related to TTN, which controls myocardial contractility and is strongly associated with dilated cardiomyopathy [\[43\]](#page-11-8). It has also been associated with atrial fibrillation [\[32\]](#page-11-9) and left ventricular parameters [\[29\]](#page-10-0). Nearby hits that are assigned to FKBP7 and PRKRA by OpenTargets, but also linked to TTN, have been linked to musculoskeletal wound healing phenotypes [\[5\]](#page-9-9) and hypertrophic cardiomyopathy [\[19\]](#page-10-12). GOSR2, CDKN1A, ADPRHL1 and LSM3 have all been linked to phenotypes like QRS duration [\[31\]](#page-11-10), PR interval [\[24\]](#page-10-13) and electrocardiogram morphology [\[41\]](#page-11-11). MECOM is associated with hypertension and blood pressure phenotypes [\[4\]](#page-9-10). HTR7 has no reported association with cardiovascular disease, but encodes a 5-HT serotonin receptor which is able to control cardiac contraction and may have a role in arrhythmias [\[21\]](#page-10-14). We also reproduce several of the top genes reported in prior studies of delta-age [\[33\]](#page-11-2), including TTN and PI15. We also find a variant assigned to TMEM270 which is proximal to ELN (another gene reported in that study).

4.6.2 GWAS Results Derived by Clustering

Our clustering hits capture established associations with cardiovascular disease, like PITX2 (atrial fibrillation [\[32\]](#page-11-9)) and SYNPO2L (atrial fibrillation [\[8\]](#page-9-11) and heart failure [\[34\]](#page-11-12)). Likewise variants in FADS1 and TMEM258 have been linked to high-density lipoprotein, red cell distribution width, and blood lipid levels in general [\[4\]](#page-9-10). CCDC141 is proximal to TTN and has been associated with pulse pressure [\[38\]](#page-11-13). ADAMTS6 has been reported in studies of cardiac conduction [\[31\]](#page-11-10) and is an emerging gene of interest [\[18\]](#page-10-15). Variants assigned to CWC27 and CENPK have less cardiovascular evidence but are proximal to ADAMTS6. ADAMTS18 may play a role in maintaining haemostatic balance and could be connected to atrial fibrillation and thrombus [\[44\]](#page-11-14). EFEMP1 has been associated with maximum left atrial volume, a potential marker of diastolic dysfunction [\[39\]](#page-11-0), and the encoded protein is also linked to adverse outcomes in heart failure [\[10\]](#page-9-12). Finally, GMDS and RARRES1 do not have much evidence linking to cardiovascular disease, suggesting a need for further exploration.

5 Discussion

Representation learning on large biomedical imaging datasets enables the discovery of new biologically relevant features in an unsupervised and unbiased manner. When combined with genetic data, this approach shows great promise for target identification. Timeseries cMRI captures spatiotemporal dynamics, which are essential for characterizing cardiovascular diseases due to their inherently temporal nature. Though previous work used representations from static short axis heart meshes to identify targets [\[6\]](#page-9-2), and other works have trained a VideoMAE on long axis videos for cardiac function prediction [\[35\]](#page-11-15), we are the first to our knowledge to use VideoMAE representations for target discovery, as well as first to incorporate complete cMRI videos combined with their deformation fields, from multiple cardiac views, to better encode the underlying dynamics of heart function.

Our benchmarking experiments suggest that video representations can capture age and cardiac parameters with high coefficient of determination. The ability to predict measures of cardiac dynamics (like ejection fraction) suggests that the representations encode spatiotemporal information. Deformation field magnitude is overall almost as good as pixel-space information when used as an input to the representation models, and there is a small but consistent benefit to concatenating the representations. Further, VideoMAE performs better than regular MAE or a pretrained ResNet, suggesting that self-supervised video modeling outperforms static image modeling. Future work could explore more sophisticated methods for fusing pixel-space and deformation fields, as well as improved modeling of temporal dynamics in the representations (rather than averaging over time frames).

Clustering the representations reveals enrichment of cardiovascular phenotypes in specific clusters. Our analysis is limited to ICD-10 codes, which are based on billing codes and generally under-estimate the true disease prevalence, so exploration of other phenotypes would be valuable. Clustering based on image-derived representations could enable subtyping of diseases and discovering at-risk patients.

Our GWAS on RDPs (delta-age and subject-to-cluster similarity) captures genes with known associations to cardiomyopathy (TTN), atrial fibrillation (PITX2, SYNPO2L), cardiac conduction (e.g. GOSR2, CDKN1A) and other cardiovascular phenotypes. We also find genes with less established connection to cardiovascular disease (e.g. HTR7, RARRES1, GMDS) which suggest further study.

While prediction benchmarks, PheWAS and GWAS are a useful starting point for validating our representations, further work is needed to define proper benchmarks for evaluating the expressiveness of representation models trained on large human cohorts.

Acknowledgements

This research was conducted using data from UK BioBank, application number 18448.

References

- [1] N. Aung et al. Genome-Wide Analysis of Left Ventricular Image-Derived Phenotypes Identifies Fourteen Loci Associated With Cardiac Morphogenesis and Heart Failure Development. *Circulation*, 140(16):1318– 1330, Oct. 2019. doi: 10.1161/CIRCULATIONAHA.119.041161. URL [https://www.ahajournals.](https://www.ahajournals.org/doi/10.1161/CIRCULATIONAHA.119.041161) [org/doi/10.1161/CIRCULATIONAHA.119.041161](https://www.ahajournals.org/doi/10.1161/CIRCULATIONAHA.119.041161). Publisher: American Heart Association.
- [2] W. Bai et al. A population-based phenome-wide association study of cardiac and aortic structure and function. *Nature Medicine*, 26(10):1654–1662, Oct. 2020. ISSN 1546-170X. doi: 10.1038/s41591-020-1009-y. URL <https://www.nature.com/articles/s41591-020-1009-y>. Publisher: Nature Publishing Group.
- [3] G. Balakrishnan et al. An Unsupervised Learning Model for Deformable Medical Image Registration. In *2018 IEEE/CVF Conference on Computer Vision and Pattern Recognition*, pages 9252–9260, June 2018. doi: 10.1109/CVPR.2018.00964. URL <https://ieeexplore.ieee.org/document/8579062>. ISSN: 2575-7075.
- [4] A. R. Barton et al. Whole-exome imputation within UK Biobank powers rare coding variant association and fine-mapping analyses. *Nature genetics*, 53(8):1260–1269, Aug. 2021. ISSN 1546-1718. doi: 10.1038/s41588-021-00892-1. URL <https://europepmc.org/articles/PMC8349845>.
- [5] P. Baumert et al. Polygenic mechanisms underpinning the response to exercise-induced muscle damage in humans: In vivo and in vitro evidence. *Journal of Cellular Physiology*, 237(7):2862–2876, 2022. ISSN 1097-4652. doi: 10.1002/jcp.30723. URL [https://onlinelibrary.wiley.com/doi/abs/10.1002/](https://onlinelibrary.wiley.com/doi/abs/10.1002/jcp.30723) [jcp.30723](https://onlinelibrary.wiley.com/doi/abs/10.1002/jcp.30723).
- [6] R. Bonazzola et al. Unsupervised ensemble-based phenotyping enhances discoverability of genes related to left-ventricular morphology. *Nature Machine Intelligence*, 6(3):291–306, Mar. 2024. ISSN 2522-5839. doi: 10.1038/s42256-024-00801-1. URL <https://www.nature.com/articles/s42256-024-00801-1>. Publisher: Nature Publishing Group.
- [7] C. Bycroft et al. The UK Biobank resource with deep phenotyping and genomic data. *Nature*, 562(7726): 203–209, Oct. 2018. ISSN 1476-4687. doi: 10.1038/s41586-018-0579-z. URL [https://www.nature.](https://www.nature.com/articles/s41586-018-0579-z) [com/articles/s41586-018-0579-z](https://www.nature.com/articles/s41586-018-0579-z). Publisher: Nature Publishing Group.
- [8] A. G. Clausen et al. Loss-of-Function Variants in the SYNPO2L Gene Are Associated With Atrial Fibrillation. *Frontiers in Cardiovascular Medicine*, 8:650667, Mar. 2021. ISSN 2297-055X. doi: 10.3389/fcvm.2021.650667. URL <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7985167/>.
- [9] J. Deng et al. ImageNet: A large-scale hierarchical image database. In *2009 IEEE Conference on Computer Vision and Pattern Recognition*, pages 248–255, June 2009. doi: 10.1109/CVPR.2009.5206848. URL <https://ieeexplore.ieee.org/document/5206848>. ISSN: 1063-6919.
- [10] M. Dib et al. Proteomic Associations of Adverse Outcomes in Human Heart Failure. *Journal of the American Heart Association*, 13(5):e031154, Mar. 2024. doi: 10.1161/JAHA.123.031154. URL [https:](https://www.ahajournals.org/doi/full/10.1161/JAHA.123.031154) [//www.ahajournals.org/doi/full/10.1161/JAHA.123.031154](https://www.ahajournals.org/doi/full/10.1161/JAHA.123.031154). Publisher: Wiley.
- [11] D. T. Ginat et al. Cardiac Imaging: Part 1, MR Pulse Sequences, Imaging Planes, and Basic Anatomy. *American Journal of Roentgenology*, 197(4):808–815, 2011. doi: 10.2214/AJR.10.7231. URL [https:](https://doi.org/10.2214/AJR.10.7231) [//doi.org/10.2214/AJR.10.7231](https://doi.org/10.2214/AJR.10.7231).
- [12] K. He et al. Deep Residual Learning for Image Recognition. In *2016 IEEE Conference on Computer Vision and Pattern Recognition (CVPR)*, pages 770–778, June 2016. doi: 10.1109/CVPR.2016.90. URL <https://ieeexplore.ieee.org/document/7780459>. ISSN: 1063-6919.
- [13] K. He et al. Masked Autoencoders Are Scalable Vision Learners. pages 16000–16009, 2022. URL [https://openaccess.thecvf.com/content/CVPR2022/html/He_Masked_Autoencoders_](https://openaccess.thecvf.com/content/CVPR2022/html/He_Masked_Autoencoders_Are_Scalable_Vision_Learners_CVPR_2022_paper) [Are_Scalable_Vision_Learners_CVPR_2022_paper](https://openaccess.thecvf.com/content/CVPR2022/html/He_Masked_Autoencoders_Are_Scalable_Vision_Learners_CVPR_2022_paper).
- [14] M. Jaderberg et al. Spatial Transformer Networks. In *Advances in Neural Information Processing Systems*, volume 28. Curran Associates, Inc., 2015. URL [https://proceedings.neurips.cc/paper_files/](https://proceedings.neurips.cc/paper_files/paper/2015/hash/33ceb07bf4eeb3da587e268d663aba1a-Abstract.html) [paper/2015/hash/33ceb07bf4eeb3da587e268d663aba1a-Abstract.html](https://proceedings.neurips.cc/paper_files/paper/2015/hash/33ceb07bf4eeb3da587e268d663aba1a-Abstract.html).
- [15] Y. Liu et al. Genetic architecture of 11 organ traits derived from abdominal MRI using deep learning. *eLife*, 10:e65554, June 2021. ISSN 2050-084X. doi: 10.7554/eLife.65554. URL [https://doi.org/10.7554/](https://doi.org/10.7554/eLife.65554) [eLife.65554](https://doi.org/10.7554/eLife.65554). Publisher: eLife Sciences Publications, Ltd.
- [16] J. Mbatchou et al. Computationally efficient whole-genome regression for quantitative and binary traits. *Nature Genetics*, 53(7):1097–1103, July 2021. ISSN 1546-1718. doi: 10.1038/s41588-021-00870-7. URL <https://www.nature.com/articles/s41588-021-00870-7>. Publisher: Nature Publishing Group.
- [17] L. McInnes et al. UMAP: Uniform Manifold Approximation and Projection for Dimension Reduction, Sept. 2020. URL <http://arxiv.org/abs/1802.03426>. arXiv:1802.03426 [cs, stat].
- [18] T. J. Mead. ADAMTS6: Emerging roles in cardiovascular, musculoskeletal and cancer biology. *Frontiers in Molecular Biosciences*, 9, Oct. 2022. ISSN 2296-889X. doi: 10.3389/fmolb.2022. 1023511. URL [https://www.frontiersin.org/journals/molecular-biosciences/articles/](https://www.frontiersin.org/journals/molecular-biosciences/articles/10.3389/fmolb.2022.1023511/full) [10.3389/fmolb.2022.1023511/full](https://www.frontiersin.org/journals/molecular-biosciences/articles/10.3389/fmolb.2022.1023511/full). Publisher: Frontiers.
- [19] R. Mendes de Almeida et al. Whole gene sequencing identifies deep-intronic variants with potential functional impact in patients with hypertrophic cardiomyopathy. *PLoS ONE*, 12(8):e0182946, Aug. 2017. ISSN 1932-6203. doi: 10.1371/journal.pone.0182946. URL [https://www.ncbi.nlm.nih.gov/pmc/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5552324/) [articles/PMC5552324/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5552324/).
- [20] L. A. Millard et al. Software Application Profile: PHESANT: a tool for performing automated phenome scans in UK Biobank. *International Journal of Epidemiology*, 47(1):29–35, Oct. 2017. ISSN 0300-5771. doi: 10.1093/ije/dyx204. URL <https://doi.org/10.1093/ije/dyx204>.
- [21] J. Neumann et al. Cardiac Roles of Serotonin (5-HT) and 5-HT-Receptors in Health and Disease. *International Journal of Molecular Sciences*, 24(5):4765, Jan. 2023. ISSN 1422-0067. doi: 10.3390/ijms24054765. URL <https://www.mdpi.com/1422-0067/24/5/4765>. Number: 5 Publisher: Multidisciplinary Digital Publishing Institute.
- [22] C. Ning et al. Genome-wide association analysis of left ventricular imaging-derived phenotypes identifies 72 risk loci and yields genetic insights into hypertrophic cardiomyopathy. *Nature Communications*, 14(1): 7900, Nov. 2023. ISSN 2041-1723. doi: 10.1038/s41467-023-43771-5. URL [https://www.nature.](https://www.nature.com/articles/s41467-023-43771-5) [com/articles/s41467-023-43771-5](https://www.nature.com/articles/s41467-023-43771-5). Publisher: Nature Publishing Group.
- [23] B. J. North et al. The Intersection Between Aging and Cardiovascular Disease. *Circulation Research*, 110(8):1097–1108, 2012. doi: 10.1161/CIRCRESAHA.111.246876. URL [https://www.ahajournals.](https://www.ahajournals.org/doi/abs/10.1161/CIRCRESAHA.111.246876) [org/doi/abs/10.1161/CIRCRESAHA.111.246876](https://www.ahajournals.org/doi/abs/10.1161/CIRCRESAHA.111.246876).
- [24] I. Ntalla et al. Multi-ancestry GWAS of the electrocardiographic PR interval identifies 202 loci underlying cardiac conduction. *Nature communications*, 11(1):2542, May 2020. ISSN 2041-1723. doi: 10.1038/ s41467-020-15706-x. URL <https://europepmc.org/articles/PMC7242331>.
- [25] D. Ochoa et al. The next-generation Open Targets Platform: reimagined, redesigned, rebuilt. *Nucleic Acids Research*, 51(D1):D1353–D1359, Nov. 2022. ISSN 0305-1048. doi: 10.1093/nar/gkac1046. URL <https://doi.org/10.1093/nar/gkac1046>.
- [26] S. Ometto et al. Unsupervised cardiac MRI phenotyping with 3D diffusion autoencoders reveals novel genetic insights, Nov. 2024. URL [https://www.medrxiv.org/content/10.1101/2024.11.](https://www.medrxiv.org/content/10.1101/2024.11.04.24316700v1) [04.24316700v1](https://www.medrxiv.org/content/10.1101/2024.11.04.24316700v1). Pages: 2024.11.04.24316700.
- [27] K. Patel et al. Unsupervised deep representation learning enables phenotype discovery for genetic association studies of brain imaging. *Communications Biology*, 7(1):414, Apr. 2024. ISSN 2399-3642. doi: 10.1038/s42003-024-06096-7. URL <https://doi.org/10.1038/s42003-024-06096-7>.
- [28] S. E. Petersen et al. Imaging in population science: cardiovascular magnetic resonance in 100,000 participants of UK Biobank - rationale, challenges and approaches. *Journal of Cardiovascular Magnetic Resonance*, 15(1):46, May 2013. ISSN 1532-429X. doi: 10.1186/1532-429X-15-46. URL [https:](https://doi.org/10.1186/1532-429X-15-46) [//doi.org/10.1186/1532-429X-15-46](https://doi.org/10.1186/1532-429X-15-46).
- [29] J. P. Pirruccello et al. Analysis of cardiac magnetic resonance imaging in 36,000 individuals yields genetic insights into dilated cardiomyopathy. *Nature Communications*, 11(1):2254, May 2020. ISSN 2041-1723. doi: 10.1038/s41467-020-15823-7. URL [https://www.nature.com/articles/](https://www.nature.com/articles/s41467-020-15823-7) [s41467-020-15823-7](https://www.nature.com/articles/s41467-020-15823-7). Publisher: Nature Publishing Group.
- [30] J. P. Pirruccello et al. Genetic analysis of right heart structure and function in 40,000 people. *Nature Genetics*, 54(6):792–803, June 2022. ISSN 1546-1718. doi: 10.1038/s41588-022-01090-3. URL <https://www.nature.com/articles/s41588-022-01090-3>. Publisher: Nature Publishing Group.
- [31] B. P. Prins et al. Exome-chip meta-analysis identifies novel loci associated with cardiac conduction, including ADAMTS6. *Genome biology*, 19(1):87, July 2018. ISSN 1474-760X. doi: 10.1186/s13059-018-1457-6. URL <https://europepmc.org/articles/PMC6048820>.
- [32] C. Roselli et al. Multi-ethnic genome-wide association study for atrial fibrillation. *Nature genetics*, 50(9): 1225–1233, June 2018. ISSN 1546-1718. doi: 10.1038/s41588-018-0133-9. URL [https://europepmc.](https://europepmc.org/articles/PMC6136836) [org/articles/PMC6136836](https://europepmc.org/articles/PMC6136836).
- [33] M. Shah et al. Environmental and genetic predictors of human cardiovascular ageing. *Nature Communications*, 14(1):4941, Aug. 2023. ISSN 2041-1723. doi: 10.1038/s41467-023-40566-6. URL <https://www.nature.com/articles/s41467-023-40566-6>. Publisher: Nature Publishing Group.
- [34] S. Shah et al. Genome-wide association and Mendelian randomisation analysis provide insights into the pathogenesis of heart failure. *Nature Communications*, 11(1):163, Jan. 2020. ISSN 2041-1723. doi: 10.1038/s41467-019-13690-5. URL <https://www.nature.com/articles/s41467-019-13690-5>. Publisher: Nature Publishing Group.
- [35] C. Shen et al. Spatiotemporal Representation Learning for Short and Long Medical Image Time Series. In M. G. Linguraru, Q. Dou, A. Feragen, S. Giannarou, B. Glocker, K. Lekadir, and J. A. Schnabel, editors, *Medical Image Computing and Computer Assisted Intervention – MICCAI 2024*, pages 656–666, Cham, 2024. Springer Nature Switzerland. ISBN 978-3-031-72120-5. doi: 10.1007/978-3-031-72120-5_61.
- [36] K. Simonyan et al. Two-Stream Convolutional Networks for Action Recognition in Videos. In *Advances in Neural Information Processing Systems*, volume 27. Curran Associates, Inc., 2014. URL [https://proceedings.neurips.cc/paper/2014/hash/](https://proceedings.neurips.cc/paper/2014/hash/00ec53c4682d36f5c4359f4ae7bd7ba1-Abstract.html) [00ec53c4682d36f5c4359f4ae7bd7ba1-Abstract.html](https://proceedings.neurips.cc/paper/2014/hash/00ec53c4682d36f5c4359f4ae7bd7ba1-Abstract.html).
- [37] S. M. Smith et al. Estimation of brain age delta from brain imaging. *NeuroImage*, 200:528–539, 2019. ISSN 1053-8119. doi: https://doi.org/10.1016/j.neuroimage.2019.06.017. URL [https://www.sciencedirect.](https://www.sciencedirect.com/science/article/pii/S1053811919305026) [com/science/article/pii/S1053811919305026](https://www.sciencedirect.com/science/article/pii/S1053811919305026).
- [38] P. Surendran et al. Discovery of rare variants associated with blood pressure regulation through metaanalysis of 1.3 million individuals. *Nature Genetics*, 52(12):1314–1332, Dec. 2020. ISSN 1546-1718. doi: 10.1038/s41588-020-00713-x. URL <https://www.nature.com/articles/s41588-020-00713-x>. Publisher: Nature Publishing Group.
- [39] M. Thanaj et al. Genetic and environmental determinants of diastolic heart function. *Nature Cardiovascular Research*, 1(4):361–371, Apr. 2022. ISSN 2731-0590. doi: 10.1038/s44161-022-00048-2. URL [https:](https://www.nature.com/articles/s44161-022-00048-2) [//www.nature.com/articles/s44161-022-00048-2](https://www.nature.com/articles/s44161-022-00048-2). Publisher: Nature Publishing Group.
- [40] V. A. Traag et al. From Louvain to Leiden: guaranteeing well-connected communities. *Scientific Reports*, 9(1):5233, Mar. 2019. ISSN 2045-2322. doi: 10.1038/s41598-019-41695-z. URL [https:](https://www.nature.com/articles/s41598-019-41695-z) [//www.nature.com/articles/s41598-019-41695-z](https://www.nature.com/articles/s41598-019-41695-z). Publisher: Nature Publishing Group.
- [41] N. Verweij et al. The Genetic Makeup of the Electrocardiogram. *Cell systems*, 11(3):229–238.e5, Sept. 2020. ISSN 2405-4720. doi: 10.1016/j.cels.2020.08.005. URL [https://europepmc.org/articles/](https://europepmc.org/articles/PMC7530085) [PMC7530085](https://europepmc.org/articles/PMC7530085).
- [42] L. Wang et al. VideoMAE V2: Scaling Video Masked Autoencoders with Dual Masking. In *2023 IEEE/CVF Conference on Computer Vision and Pattern Recognition (CVPR)*, pages 14549–14560, June 2023. doi: 10.1109/CVPR52729.2023.01398. URL [https://ieeexplore.ieee.org/document/](https://ieeexplore.ieee.org/document/10203656) [10203656](https://ieeexplore.ieee.org/document/10203656). ISSN: 2575-7075.
- [43] J. S. Ware et al. Role of titin in cardiomyopathy: from DNA variants to patient stratification. *Nature Reviews Cardiology*, 15(4):241–252, Apr. 2018. ISSN 1759-5010. doi: 10.1038/nrcardio.2017.190. URL <https://www.nature.com/articles/nrcardio.2017.190>. Publisher: Nature Publishing Group.
- [44] J. Wei et al. ADAMTS-18: A metalloproteinase with multiple functions. *Frontiers in bioscience (Landmark edition)*, 19:1456–1467, June 2014. ISSN 1093-4715. URL [https://www.ncbi.nlm.nih.gov/pmc/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4410865/) [articles/PMC4410865/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4410865/).
- [45] F. A. Wolf et al. SCANPY: large-scale single-cell gene expression data analysis. *Genome Biology*, 19(1): 15, Feb. 2018. ISSN 1474-760X. doi: 10.1186/s13059-017-1382-0. URL [https://doi.org/10.1186/](https://doi.org/10.1186/s13059-017-1382-0) [s13059-017-1382-0](https://doi.org/10.1186/s13059-017-1382-0).

A Appendix / supplemental material

A.1 Comparison to Prior Work on Age Prediction

As another benchmark, we compare our age prediction model to prior work that predicted cardiovascular age from manually defined cMRI phenotypes [\[33\]](#page-11-2). Their analysis is performed on a subset of 5063 "healthy" participants from the UK BioBank imaging cohort. They define "healthy" as the absence of cardiovascular, metabolic or respiratory disease (as defined by presence of disease codes or self-reported illness) with BMI under 30. They then split the data into 80% train / 20% test and trained a gradient boosting model from a series of manually extracted IDPs. Although we do not have access to the list of subjects they used for the train/test split, we followed their data filtering procedure to define a comparable set of 5063 "healthy" participants split into 80% train / 20% test, then trained a ridge regression model from our combined long + short axis embeddings (as described in Section [4.2\)](#page-4-1). Results are reported in Table [S1;](#page-4-0) although the test sets are not exactly comparable, our model shows significantly better performance, further validating our approach.

Table S1: Comparison of age prediction from cMRI features on a healthy subcohort. MAE = mean absolute error.

A.2 PheWAS Hits

Tables [S2](#page-5-0) and [S3](#page-14-0) list the top ICD-10 disease code hits for RDPs derived from long axis and short axis (respectively). For each ICD-10 code, we count the number of RDPs (delta age $+ K$ cluster similarities) with a significance of $p \le 0.05/635$ (applying a Bonferroni correction for the number of ICD-10 codes are being tested). We report the most significant RDP association and the total number of significant associations.

A.3 GWAS Hits

Tables [S4](#page-15-0) and [S5](#page-16-0) list the top gene hits for RDPs derived from long axis and short axis (respectively). Similiar to the PheWAS results, for each gene we count the number of RDPs (delta age $+ K$ cluster similarities) with a significance of $p \le 5.0 \times 10^{-8}$ (applying a Bonferroni correction based on the number of variants being tested). We report the lead variant, most significant RDP association, and the total number of significant associations.

Table S2: Top 50 aggregated PheWAS hits for representation-derived phenotypes from long-axis scans, in descending order of significance.

Table S3: Top 50 aggregated PheWAS hits for representation-derived phenotypes from short-axis scans, in descending order of significance.

Gene Name	Lead Variant	Max $-log10(p)$	# Significant RDPs	Most Significant RDP
TTN	rs2042995	47.328	4	delta_age
FKBP7	rs1001238	45.229	3	delta_age
PRKRA	rs2253324	24.008	1	delta_age
PITX2	rs4611994	21.857	4	cluster11
GOSR2	rs17608766	17.731	1	delta_age
EFEMP1	rs59985551	16.749	3	cluster13
MECOM	rs9850919	16.685	1	delta_age
CWC ₂₇	rs2278353	16.201	6	cluster0
CCDC141	rs17362588	15.792	4	cluster4
CENPK	rs1309553	15.781	4	cluster0
GMDS	rs767102318	15.7078	5	cluster1
RARRES1	rs12637678	15.358	4	cluster13
CDKN1A	rs113578873	15.1916	1	delta_age
ADAMTS6	rs4700662	15.0322	4	cluster1
HTR7	rs10748555	15.0027	\overline{c}	delta_age
TMEM270	rs370616120	14.8768	1	delta_age
FADS1	rs174566	14.4253	$\mathbf{1}$	cluster3
RPRML	rs145153053	14.2427	$\mathbf{1}$	delta_age
TMEM258	rs174533	13.9727	1	cluster3
SYNPO2L	rs60632610	13.7004	1	cluster11
PLEC	rs11786896	13.1577	$\overline{4}$	cluster11
PI15	rs2732010	13.0876	1	delta_age
FADS ₂	rs97384	12.917	1	cluster3
CRIP1	rs55633823	12.7888	\overline{c}	cluster0
WNT3	rs75230966	12.7247	1	delta_age
ADAMTS18	rs17689197	12.6626	3	cluster13
CCDC91	rs143823594	12.6286	4	cluster12
FAM241A	rs11376680	12.5597	3	cluster0
NUDT7	rs62043885	12.3228	3	cluster13
CPED ₁	rs3801387	12.2871	4	cluster1
ADPRHL1	rs76382172	11.8723	1	delta_age
TIFA	rs326847	11.7791	3	cluster0
RSRC1	rs73164066	11.6457	3	cluster13
CASQ2	rs10157905	11.6026	$\mathbf{1}$	cluster11
HHIP	rs1512288	11.5656	\overline{c}	cluster3
FUT11	rs11000771	11.2002	$\mathbf{1}$	cluster11
LSM3	rs13061705	11.1621	1	delta_age
ZFPM1	rs12595858	11.0858	\overline{c}	cluster4
NDST ₂	rs4746151	11.0666	$\mathbf 1$	cluster11
COL8A1	rs114796243	10.8133	3	cluster5
LMCD1	rs165177	10.6812	3	cluster0
TBX3	rs5801092	10.6752	3	cluster1
NPR3	rs13154066	10.6182	$\mathbf{1}$	delta_age
CMSS1	rs12488245	10.5618	\overline{c}	cluster6
CRIM1	rs4670549	10.5224	$\mathbf{1}$	delta_age
	rs62236745		$\mathbf{1}$	
TRIOBP C7orf25	rs13234512	10.4527 10.3517	\overline{c}	cluster16 cluster1
LRRC37A2			$\mathbf{1}$	
	rs3874943	10.3187		delta_age delta age
SLC6A6 KCNJ2	rs10865722 rs2285569	10.2497 10.235	1 1	cluster6

Table S4: Top 50 aggregated GWAS hits for representation-derived phenotypes from long-axis scans, in descending order of significance.

Gene Name	Lead Variant	Max -log10(p)	Num Significant RDPs	Most Significant RDP
TTN	rs2042995	31.6562	1	delta_age
FKBP7	rs2042996	31.3298	1	delta_age
LIMK1	rs113395463	23.1973	1	delta_age
PI15	rs6472877	19.593	1	delta_age
NUDT7	rs62043885	16.1109	3	cluster8
ADAMTS18	rs17689197	15.9459	3	cluster8
RARRES1	rs1714518	14.5289	1	cluster8
GOSR ₂	rs17608766	14.0096	1	delta_age
WRNIP1	rs4355649	13.1031	1	cluster3
ADPRHL1	rs76382172	12.9088	1	delta_age
PITX2	rs6843082	12.4039	$\mathbf{1}$	cluster9
TMEM270	rs79344387	12.1143	$\mathbf{1}$	delta_age
NPR ₃	rs13154066	12.0771	$\mathbf{1}$	delta_age
RPRML	rs117953218	11.9735	$\mathbf{1}$	delta_age
MECOM	rs34585560	10.8959	1	delta_age
FBXO32	rs7006122	10.6356	1	delta_age
LRRC37A	rs2532351	10.621	$\mathbf{1}$	delta_age
WNT3	rs8069437	10.5871	$\mathbf{1}$	delta_age
FAM241A	rs1903403	10.5165	$\overline{4}$	cluster8
SMG ₆	rs903160	10.4007	1	delta_age
SESTD1	rs10930844	10.3843	1	delta_age
RSRC1	rs73164066	10.3262	1	cluster8
TRIB ₂	rs6727552	10.2328	4	cluster3
HTR7	rs10748555	9.9881	1	delta_age
LSM3	rs11715111	9.87709	1	delta_age
KANSL1	rs7225002	9.8478	1	delta_age
CWC27	rs540834152	9.76052	5	cluster ₅
MAPT	rs5820605	9.73052	$\mathbf{1}$	delta_age
TIFA	rs326847	9.59493	\overline{c}	cluster8
CENPK	rs1309558	9.4822	3	cluster ₅
HDGFL1	rs13211463	9.4656	1	
CCDC141	rs373251994	9.37475	1	delta_age delta_age
CDCA ₂	rs11985475	9.02453	$\mathbf{1}$	delta_age
TRIOBP	rs62236745	8.85557	$\mathbf{1}$	cluster10
GMDS	rs2761236	8.83456	5	cluster ₈
GORAB	rs59452262	8.79816	1	cluster1
SRR	rs11371517	8.71519	1	delta_age
MON1B	rs62046461	8.66593	1	cluster8
NMB	rs8033343	8.62444	$\mathbf 1$	cluster6
SLC6A6	rs10865722	8.60522	1	delta_age
CPS1	rs1047891	8.59287	$\mathbf{1}$	delta_age
CFTR	rs10235008	8.58544	1	cluster10
WNT ₂	rs73211959	8.49328	1	cluster10
CBR4	rs12644874	8.42726	\overline{c}	cluster0
WT1	rs10835891	8.41244	1	cluster ₅
PNPT1	rs77045491	8.29785	1	cluster4
EFEMP1	rs1346786	8.29432	$\overline{2}$	cluster2
SOX5	rs137913153	8.28827	1	cluster9
MLF1	rs10154978	8.26661	1	cluster8
ZEB ₂	rs12996668	8.2329	1	cluster10

Table S5: Top 50 aggregated GWAS hits for representation-derived phenotypes from short-axis scans, in descending order of significance.

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