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# Improving Protein Subcellular Localization Prediction with Structural Prediction & Graph Neural Networks

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## Abstract

1 We present a method that improves subcellular localization prediction for proteins  
2 based on their sequence by leveraging structure prediction and Graph Neural  
3 Networks. We demonstrate that Language Models, trained on protein sequences,  
4 and Graph Neural Nets, trained on protein's 3D structures, are both efficient  
5 approaches. They both learn meaningful, yet different representations of proteins;  
6 hence, ensembling them outperforms the reigning state of the art method.

## 7 1 Introduction

8 The elucidation of protein function is a fundamental challenge in biology. A significant aspect of  
9 protein function is the determination of its cellular localization. Many processes, such as disease  
10 mechanisms, drug performance, regulation of metabolic processes, and signaling cascades all depend  
11 on the protein's localization, hence significant experimental efforts are dedicated to this end. Never-  
12 theless, the exponential growth in the availability of proteomics information from newly sequenced  
13 organisms and metagenomes creates a huge gap in the experimental elucidation of protein sub-cellular  
14 localization for gene and genome annotation. This creates a need to *predict* protein localization using  
15 sequence data.

16 Frequently, the location of a protein in the cell is determined by a "localization signal" – a short  
17 segment of the protein that is recognized by receptors located in the target cellular compartment.  
18 For example, proteins that include a *nuclear localization signal* (NLS) are recognized by nuclear  
19 receptors and, once synthesized, will be shuttled into the nucleus. Similarly, proteins that include a  
20 *signal peptide* will be exported to the extracellular space. Many computational methods are thus based  
21 on the identification of short motifs in the protein sequence that determine its cellular localization.  
22 Still, many proteins are shuttled to their target compartments using chaperones or other mechanisms,  
23 leading to inaccurate prediction by motif-based approaches. More recent predictive models utilize  
24 the entire sequence of a protein to predict its localization using protein sequence language models  
25 such as ESM [1]. The prediction power of these methods was shown to be higher than motif-based  
26 approaches [2]. The recent advance in protein structure prediction [3][4] enables another avenue of  
27 development: utilizing protein structure for the prediction of protein localization.

28 In this paper, we propose a Graph Neural Network (GNN) that utilizes protein structure information  
29 for protein sub-cellular localization prediction. We ensemble this model with a transformer-based  
30 language model that takes the protein sequence into account. We demonstrate that this combined  
31 method yields higher prediction performances than previous methods.

## 32 **2 Background work**

### 33 **2.1 Language Model Representation**

34 Meta AI’s Evolutionary Scale Modeling (ESM) [1] is an unsupervised transformer protein language  
35 model. In this paper, we employ ESM-1b, in which the model is trained to predict amino acids from  
36 the surrounding sequence context, as the core model to extract abstract features from the protein  
37 sequences. Although a competing model – Hugging Face’s ProtT5XL [5] – is available, we focused  
38 on ESM-1b for its better ability to scale <sup>1</sup>.

### 39 **2.2 Subcellular localization**

40 Alongside their cellular localization dataset, the authors of Deeploc 2 [2] published a successful  
41 Language Model (LM)-based classifier. It feeds the ESM-1b representation to a multi-class clas-  
42 sifier involving an attention-pooling layer and a multi-layer perception (MLP). Its performance is  
43 summarized in Table 1.

## 44 **3 Method**

45 In recent years, the use Language Models for protein representation has proven to be surprisingly  
46 successful for downstream prediction models; accordingly, their use has become increasingly preva-  
47 lent in the field [1] [6] [5] [7]. Often, LMs are trained in a self-supervised fashion on hundreds of  
48 millions of proteins before being fine-tuned or used as backbone models for downstream tasks [8].  
49 In parallel, the proteomics community underwent a major breakthrough with the inventions of very  
50 successful in-silico protein folders [3] [4]. The goal of our research is to combine those two new  
51 sources of information to outperform models that only utilize one such source.

### 52 **3.1 Data**

53 In this paper we are benchmarking our method on the subcellular localization task made possible  
54 by the Deeploc 2 [2] dataset. This dataset is comprised of a train/validation set of 24,674 protein  
55 sequences mapped to 10 localization classes (*e.g.* Cytoplasm, Nucleus, etc.), and a test set from a  
56 different source – the Human Protein Atlas (HPA)[9] – of 1,532 proteins associated with 8 localization  
57 classes <sup>2</sup>.

58 For each protein entry, we retrieved the AlphaFold model from the AlphaFold Protein Structure  
59 Database [10] in the form of an atom coordinates file (PDB format). For each structure, we generated  
60 a graph representation based on the amino-acid adjacency matrix, inspired by the LM-GVP method  
61 [11]. The dihedral angle between amino-acids is used as a node feature in the model to capture  
62 the geometrical relationships between the adjacent amino acids. We introduced two novel node  
63 attributes to the LM-GVP graph: (1) **positional encoding (PE)**: Adjacency matrices do not encode  
64 protein’s primary sequence, therefore (a) will have low chances of recognizing primary-sequence  
65 motifs, and (b) will be less sensitive to the distinction between short and long range amino acid  
66 interactions, known to have different contributions to protein folding, structure and function [12],  
67 and (2) **AlphaFold’s Local Distance Difference Test (pLDDT)**: pLDDT is a metric of AlphaFold’s  
68 confidence for atom coordinate assignment. It has been observed that regions of the protein structure  
69 that have a higher pLDDT are generally associated to "less rigid" parts of the structure [13]. We  
70 hypothesized that pLDDT increases the predictive power for many downstream tasks, such as cellular  
71 localization.

### 72 **3.2 Baseline**

73 In order to compare the performance of our contribution, we have trained different simple NN  
74 architectures (CNNs, MLPs, shallow models) on the ESM representation. We kept the best performing  
75 model: a 3 layers convolutional neural network.

<sup>1</sup>ProtT5XL has 3B parameters and ESM-1b has 650M parameters. ProtT5XL was shown to be marginally better than ESM-1b for localization prediction. [2]

<sup>2</sup>2 of the training classes are not present in the test set. Furthermore, we also removed 2 supplementary classes from the test results since they were not significantly represented with less than 7 occurrences.

76 **3.3 Model Architecture**

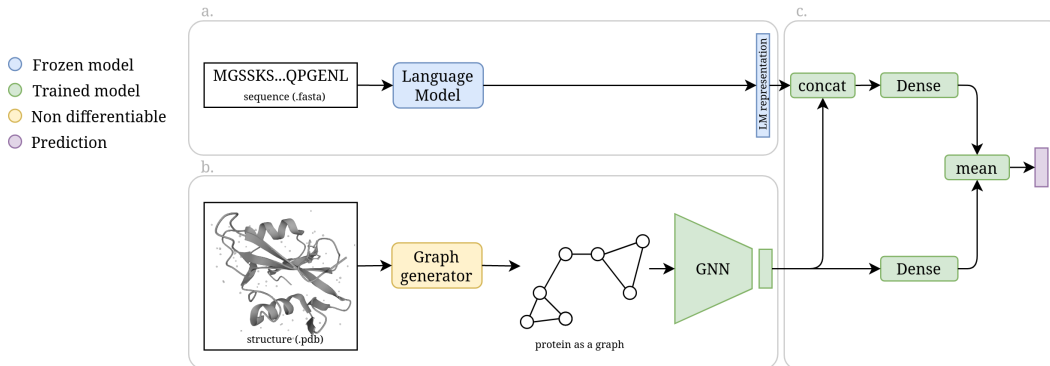


Figure 1: First (a.), a protein sequence is passed to an LM, which outputs a protein representation. In parallel (b.) a PDB structure is processed to generate a graph where nodes represent amino acids and edges are drawn between adjacent amino acids. This graph is used as an input for a GNN (with GVP layers [14]) which outputs a graph representation. Finally (c.), both LM and Graph representations are ensembled through a novel architecture that yields the class probabilities.

77 Similar architectures, like LM-GVP and DeepFRI are using LM representation at the node/amino  
 78 acid-level. Whilst this makes intuitive sense, we realized that it leads to scalability issues. Given that  
 79 the average protein length is 556.90 for the dataset, using one LM representations per amino acid  
 80 can quickly exceed the average memory of single GPU or lead to excessive training times. Instead,  
 81 we are working with a fixed-size representation vector per protein, using the mean to aggregate the  
 82 representations like suggested by the ESM team.

83 Our first hypothesis is that the representations from LM and GVP are complementary. Therefore, our  
 84 first approach was to train two separate classifiers and to ensemble them by computing the mean of  
 85 the output probabilities. The resulting F1-score were 0.55 for the LM model, 0.46 for the GVP model  
 86 and 0.58 for the ensemble. Whilst this ensembling approach already outperformed each classifier  
 87 separately, we realized that we could improve performance if we trained both classifiers together with  
 88 a differentiable mean. We believe that this method allows the GVP layers to "learn away" from what  
 89 the LM already represents. After experimenting with various architectures to combine GVP and LM  
 90 representations, we found the best performing to be a dense-residual type head (Figure 1.c).

91 **4 Results**

Model	Precision	Recall	F1-score
1. GVP alone	0.34	0.69	0.46
2. LM baseline	0.67	0.47	0.55
3. LM attention pooling (Deeploc 2)	-	-	0.57
4. Proposed architecture (Figure 1)	0.50	0.72	0.59
5. Ensemble of 2. and 4.	0.59	0.60	<b>0.60</b>

Table 1: Summary of the classification metrics (micro average) on the HPA test set for different approaches. (1) GVP alone as a classifier (Figure 1.b), (2) LM baseline model (3 layers CNN), (3) Architecture proposed by Deeploc 2, (4) Our dense-residual ensembling (Figure 1), (5) A supplementary (non-differentiable) ensemble by the mean of 2. and 4. slightly outperforms 4. alone.

92 Similar to other researchers[15], we observed that LM-powered models clearly outperform GVP  
 93 alone. However, our proposed ensembling architecture outperforms the best LM and structure-aware  
 94 approaches respectively. Note that the proposed architecture (4) using representations at the protein  
 95 level outperforms the competing model (3) that use representations at the amino acid level; thus  
 96 provides both faster training and better performance.

Model	Cytoplasm	Nucleus	Cell memb.	Mitochondrion	Endo. ret.	Golgi
LM baseline	0.53	0.66	0.33	0.62	0.13	0.19
Ours	0.56	0.75	0.43	0.51	0.20	0.31

Table 2: Micro F1-score per class for both LM baseline and our proposed ensembling architecture (Figure 1) on the HPA test set.

97 We observe that in every class (except Mitochondrion), our proposed architecture outperforms the  
98 LM baseline.

99 In order to show the impact of adding positional encoding as a node feature, we performed an ablation  
100 experiment. We trained with and without PE for 60 epochs. Without PE, the test F1-score reached  
101 0.548, with PE it reached 0.569<sup>3</sup>. This validates the hypothesis previously formulated: helping the  
102 model extract patterns at specific positions increased the classification performance.

## 103 5 Discussion

- 104 • Given that for mitochondrion, the LM baseline outperforms the proposed architecture,  
105 we propose the following explanation. Shuttling proteins into mitochondria is unique:  
106 Mitochondrial entry requires a protein to bind to a chaperone while it is in an unfolded  
107 form. Upon entry to this organelle, the protein will adopt a folded state. [16]. Hence, we  
108 could explain the lower accuracy of GVM by the known dependence on sequence rather  
109 than structure features for mitochondrial localization.
- 110 • We observe that the prediction of nuclear localization by our ensembling method outperforms  
111 DeepLoc’s prediction. As mentioned above, Nuclear localization is frequently mediated  
112 by an NLS. However, other routes for nuclear entry exist: a protein without an NLS may  
113 interact with other proteins that include an NLS, or other chaperones that mediate entry.  
114 These cases are notably more challenging to predict, as they are not mediated directly by  
115 a conserved sequence motif such as the NLS. To demonstrate the benefit of incorporating  
116 structure information for localization prediction, we focused on cases in which the LM  
117 baseline on its own fails in nuclear prediction, but our ensembling method succeeds. We  
118 further distilled the list of these cases to include only the cases where DeepLoc provided  
119 inaccurate prediction. None of these proteins included an NLS (According to SwissProt  
120 annotation [17]), and some were literature-documented cases of proteins that enter the  
121 nucleus via a non-NLS routes, like General transcription factor IIH subunit 1 (UniProt  
122 accession P32780), which is a component of RNA polymerase II. This protein interacts with  
123 other proteins to form a complex before entering the nucleus. The complex nuclear entry is  
124 then mediated by an NLS motif of other components of the complex [18]. This stands as a  
125 promising finding in favor of using structure-aware protein representations.

## 126 6 Conclusion

127 In this research, we successfully combined signals from protein structures with Language Model  
128 representations to outperform the state of the art of Subcellular localization prediction. The principle  
129 takeaways are the following: predicted structures appear not to be the sole answer to better protein  
130 representations; however, once combined with LM representations and fed into a specific model  
131 architecture, such as the one we propose, the quality of representations improves.

132 Contrasting competing models, we focused on making a scalable/lightweight architecture allowing  
133 faster training for more downstream tasks on consumer GPUs, in an effort to democratize structure-  
134 aware protein representations models, whilst increasing the model performance.

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<sup>3</sup>Furthermore, we observed a better performance during the whole duration of training.

## References

- 135
- 136 [1] Alexander Rives, Joshua Meier, Tom Sercu, Siddharth Goyal, Zeming Lin, Jason Liu, Demi Guo,  
137 Myle Ott, C. Lawrence Zitnick, Jerry Ma, and Rob Fergus. Biological structure and function  
138 emerge from scaling unsupervised learning to 250 million protein sequences. *Proceedings of*  
139 *the National Academy of Sciences*, 118(15):e2016239118, 2021.
- 140 [2] Vineet Thumulari, José Juan Almagro Armenteros, Alexander Rosenberg Johansen, Henrik  
141 Nielsen, and Ole Winther. DeepLoc 2.0: multi-label subcellular localization prediction using  
142 protein language models. *Nucleic Acids Res.*, 50(W1):W228–W234, April 2022.
- 143 [3] Candida S. Punla, <https://orcid.org/0000-0002-1094-0018>, [cspunla@bpsu.edu.ph](mailto:cspunla@bpsu.edu.ph), Rosemarie  
144 C. Farro, <https://orcid.org/0000-0002-3571-2716>, [rcfarro@bpsu.edu.ph](mailto:rcfarro@bpsu.edu.ph), and Bataan Peninsula  
145 State University Dinalupihan, Bataan, Philippines. Are we there yet?: An analysis of the  
146 competencies of BEED graduates of BPSU-DC. *International Multidisciplinary Research*  
147 *Journal*, 4(3):50–59, September 2022.
- 148 [4] Minkyung Baek, Frank DiMaio, Ivan Anishchenko, Justas Dauparas, Sergey Ovchinnikov,  
149 Gyu Rie Lee, Jue Wang, Qian Cong, Lisa N. Kinch, R. Dustin Schaeffer, Claudia Millán,  
150 Hahnbeom Park, Carson Adams, Caleb R. Glassman, Andy DeGiovanni, Jose H. Pereira,  
151 Andria V. Rodrigues, Alberdina A. van Dijk, Ana C. Ebrecht, Diederik J. Opperman, Theo  
152 Sagmeister, Christoph Buhllheller, Tea Pavkov-Keller, Manoj K. Rathinaswamy, Udit Dalwadi,  
153 Calvin K. Yip, John E. Burke, K. Christopher Garcia, Nick V. Grishin, Paul D. Adams, Randy J.  
154 Read, and David Baker. Accurate prediction of protein structures and interactions using a  
155 three-track neural network. *Science*, 373(6557):871–876, 2021.
- 156 [5] Ahmed Elnaggar, Michael Heinzinger, Christian Dallago, Ghalia Rihawi, Yu Wang, Llion Jones,  
157 Tom Gibbs, Tamas Feher, Christoph Angerer, Martin Steinegger, Debsindhu Bhowmik, and  
158 Burkhard Rost. Prottrans: Towards cracking the language of life’s code through self-supervised  
159 deep learning and high performance computing, 2020.
- 160 [6] Nadav Brandes, Dan Ofer, Yam Peleg, Nadav Rappoport, and Michal Linial. ProteinBERT: A  
161 universal deep-learning model of protein sequence and function. May 2021.
- 162 [7] Roshan Rao, Nicholas Bhattacharya, Neil Thomas, Yan Duan, Xi Chen, John F. Canny,  
163 Pieter Abbeel, and Yun S. Song. Evaluating protein transfer learning with TAPE. *CoRR*,  
164 abs/1906.08230, 2019.
- 165 [8] Serbulent Unsal, Heval Atas, Muammer Albayrak, Kemal Turhan, Aybar C. Acar, and Tunca  
166 Doğan. Learning functional properties of proteins with language models. *Nature Machine*  
167 *Intelligence*, 4(3):227–245, March 2022.
- 168 [9] Mathias Uhlén, Linn Fagerberg, Björn M. Hallström, Cecilia Lindskog, Per Oksvold, Adil  
169 Mardinoglu, Åsa Sivertsson, Caroline Kampf, Evelina Sjöstedt, Anna Asplund, IngMarie  
170 Olsson, Karolina Edlund, Emma Lundberg, Sanjay Navani, Cristina Al-Khalili Szigyarto,  
171 Jacob Odeberg, Dijana Djureinovic, Jenny Ottosson Takanen, Sophia Hober, Tove Alm, Per-  
172 Henrik Edqvist, Holger Berling, Hanna Tegel, Jan Mulder, Johan Rockberg, Peter Nilsson,  
173 Jochen M. Schwenk, Marica Hamsten, Kalle von Feilitzen, Mattias Forsberg, Lukas Persson,  
174 Fredric Johansson, Martin Zwahlen, Gunnar von Heijne, Jens Nielsen, and Fredrik Pontén.  
175 Tissue-based map of the human proteome. *Science*, 347(6220), January 2015.
- 176 [10] Mihaly Varadi, Stephen Anyango, Mandar Deshpande, Sreenath Nair, Cindy Natassia, Galabina  
177 Yordanova, David Yuan, Oana Stroe, Gemma Wood, Agata Laydon, Augustin Židek, Tim  
178 Green, Kathryn Tunyasuvunakool, Stig Petersen, John Jumper, Ellen Clancy, Richard Green,  
179 Ankur Vora, Mira Lutfi, Michael Figurnov, Andrew Cowie, Nicole Hobbs, Pushmeet Kohli,  
180 Gerard Kleywegt, Ewan Birney, Demis Hassabis, and Sameer Velankar. AlphaFold Protein  
181 Structure Database: massively expanding the structural coverage of protein-sequence space  
182 with high-accuracy models. *Nucleic Acids Research*, 50(D1):D439–D444, 11 2021.
- 183 [11] Zichen Wang, Steven A. Combs, Ryan Brand, Miguel Romero Calvo, Panpan Xu, George Price,  
184 Nataliya Golovach, Emmanuel O. Salawu, Colby J. Wise, Sri Priya Ponnappalli, and Peter M.  
185 Clark. Lm-gvp: A generalizable deep learning framework for protein property prediction from  
186 sequence and structure. *bioRxiv*, 2021.

- 187 [12] Adesh Kumar, Anupaul Baruah, and Parbati Biswas. Role of local and nonlocal interactions in  
188 folding and misfolding of globular proteins. *The Journal of Chemical Physics*, 146(6):065102,  
189 February 2017.
- 190 [13] Kiersten M. Ruff and Rohit V. Pappu. AlphaFold and implications for intrinsically disordered  
191 proteins. *Journal of Molecular Biology*, 433(20):167208, October 2021.
- 192 [14] Bowen Jing, Stephan Eismann, Patricia Suriana, Raphael J. L. Townshend, and Ron O. Dror.  
193 Learning from protein structure with geometric vector perceptrons. *ArXiv*, abs/2009.01411,  
194 2021.
- 195 [15] Vladimir Gligorijević, P. Douglas Renfrew, Tomasz Kosciolk, Julia Koehler Leman, Daniel  
196 Berenberg, Tommi Vatanen, Chris Chandler, Bryn C. Taylor, Ian M. Fisk, Hera Vlamakis,  
197 Ramnik J. Xavier, Rob Knight, Kyunghyun Cho, and Richard Bonneau. Structure-based protein  
198 function prediction using graph convolutional networks. *Nature Communications*, 12(1), May  
199 2021.
- 200 [16] Diana Stojanovski, Maria Bohnert, Nikolaus Pfanner, and Martin van der Laan. Mechanisms of  
201 protein sorting in mitochondria. *Cold Spring Harb. Perspect. Biol.*, 4(10):a011320–a011320,  
202 October 2012.
- 203 [17] The UniProt Consortium. UniProt: the universal protein knowledgebase in 2021. *Nucleic Acids  
204 Research*, 49(D1):D480–D489, 11 2020.
- 205 [18] Luciano Di Croce. Regulating the shuttling of eukaryotic rna polymerase ii. *Molecular and  
206 Cellular Biology*, 31(19):3918–3920, 2011.