IN VIVO CELL-TYPE AND BRAIN REGION CLASSIFICATION VIA MULTIMODAL CONTRASTIVE LEARNING

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Paper under double-blind review

ABSTRACT

Current electrophysiological approaches can track the activity of many neurons, yet it is usually unknown which cell-types or brain areas are being recorded without further molecular or histological analysis. Developing accurate and scalable algorithms for identifying the cell-type and brain region of recorded neurons is thus crucial for improving our understanding of neural computation. In this work, we develop a multimodal contrastive learning approach for neural data that can be fine-tuned for different downstream tasks, including inference of cell-type and brain location. We utilize this approach to jointly embed the activity autocorrelations and extracellular waveforms of individual neurons. We demonstrate that our embedding approach, **N**euronal **E**mbeddings via **M**ultim**O**dal contrastive learning (NEMO), paired with supervised fine-tuning, achieves state-of-the-art cell-type classification for an opto-tagged visual cortex dataset and brain region classification for the public International Brain Laboratory brain-wide map dataset. Our method represents a promising step towards accurate cell-type and brain region classification from electrophysiological recordings.

1 INTRODUCTION

High-density electrode arrays now allow for simultaneous extracellular recording from hundreds to thousands of neurons across interconnected brain regions (Jun et al., 2017; Steinmetz et al., 2021; Ye et al., 2023b; Trautmann et al., 2023). While much progress has been made on developing algorithms for tracking neural activity (Buccino et al., 2020; Magland et al., 2020; Boussard et al., 2023; Pachitariu et al., 2024), identifying the cell-types and brain regions being recorded is still an open problem. Brain region classification is particularly challenging given the diversity of cell types in each region (Yao et al., 2023) and how the locations of regions differ between animals. The development of an accurate method for extracting cell-type and brain region labels will provide insights into brain development, microcircuit function, and brain disorders (Zeng & Sanes, 2017).

Traditional approaches for electrophysiological cell-type classification utilize simple features of the extracellular action potential (EAP) such as its width or peak-to-trough amplitude (Mountcastle et al., 1969; Matthews & Lee, 1991; Nowak et al., 2003; Barthó et al., 2004; Vigneswaran et al., 2011) or features of neural activity, such as the inter-spike interval distribution (Latuske et al., 2015; Jouty et al., 2018). These simple features are interpretable and easy to visualize but lack discriminative power and robustness across different datasets (Weir et al., 2015; Gouwens et al., 2019). Current automated featurization methods for EAPs (Lee et al., 2021; Vishnubhotla et al., 2024) and neural activity (Schneider et al., 2023a) improve upon manual features but are limited to a single modality.

There has been a recent push to develop multimodal methods that can integrate information from both recorded EAPs and spiking activity. PhysMAP (Lee et al., 2024) is a UMAP-based (McInnes et al., 2018a) approach that can predict cell-types using multiple physiological modalities through a weighted nearest neighbor graph. Another recently introduced method utilizes variational autoencoders (VAEs) to embed each physiological modality separately and then combines these embeddings before classification (Beau et al., 2024). Although both methods show promising results, PhysMAP cannot be fine-tuned for downstream tasks as it is nondifferentiable, and the VAE-based method captures features that are important for reconstruction, not discrimination, impairing downstream performance (Guo et al., 2017). Neither approach has been used to classify brain regions.

In this work, we introduce a multimodal contrastive learning method for neurophysiological data, Neuronal Embeddings via MultimOdal Contrastive Learning (NEMO), which utilizes large amounts of unlabeled paired data for pre-training and can be fine-tuned for different downstream tasks including cell-type and brain region classification. We utilize a recently developed contrastive learning framework (Radford et al., 2021) to jointly embed individual neurons' activity autocorrelations and average extracellular waveforms. The key assumption of our method is that information shared between different modalities captures intrinsic properties of a neuron that are predictive of its cell-type and corresponding brain region. Jointly embedding different modalities into a shared latent space, will capture this shared information while discarding modality-specific noise (Huang et al., 2024). We evaluate NEMO on cell-type classification using an optotagged Neuropixels Ultra (NP Ultra) recording from mouse visual cortex (Ye et al., 2023b) and on brain region classification using the International Brain Laboratory (IBL) brain-wide map dataset (IBL et al., 2023). Across both datasets and tasks, NEMO significantly outperforms current state-of-the-art unsupervised (PhysMAP and VAEs) and supervised methods. These results highlight NEMO's ability to extract informative representations despite high variability in the data, and demonstrate that NEMO is a significant advance towards accurate cell-type and brain region classification from electrophysiological recordings.

2 Related work

2.1 Contrastive learning for neuronal datasets

The goal of contrastive learning is to find an embedding space where similar examples are close together while dissimilar ones are well-separated (Le-Khac et al., 2020). Contrastive learning has found success across a number of domains including language (Reimers & Gurevych, 2019), vision (Chen et al., 2020), audio (Saeed et al., 2021), and multimodal learning (Radford et al., 2021; Tian et al., 2020). There has been a surge in the development of contrastive learning methods for neural data. Contrastive learning has been applied to neuronal morphological data (Chen et al., 2022), connectomics data (Dorkenwald et al., 2023) and preprocessed spiking activity (Azabou et al., 2021; Urzay et al., 2023; Schneider et al., 2023b; Antoniades et al., 2023). In each of these applications, associated downstream tasks such as 3D neuron reconstruction, cellular sub-compartment classification, or behavior prediction have shown improvement using this contrastive paradigm. One contrastive method, CEED, has been applied to raw extracellular recordings to perform spike sorting and cell-type classification. In contrast to NEMO, CEED is unimodal (it ignores neural activity) and was never applied to optotagged data or brain region classification (Vishnubhotla et al., 2024).

2.2 Cell-type classification

The goal of cell-type classification is to assign neurons to distinct classes based on their morphology, function, electrophysiological properties, and molecular markers (Masland, 2004). Current transcriptomic (Tasic et al., 2018; Yao et al., 2021; 2023) and optical methods (Cardin et al., 2010; Kravitz et al., 2013; Lee et al., 2020) are effective but require extensive sample preparation or specialized equipment, limiting their scalability and applicability for in-vivo studies (Lee et al., 2024). Recently, calcium imaging has been utilized in conjunction with molecular cell-typing to identify cell-types (Bugeon et al., 2022; Mi et al., 2023). This approach suffers from low temporal resolution and requires significant post hoc effort to collect the molecular imaging data and align it to the calcium data (and therefore this approach can not be used in closed loop in vivo experiments).

A promising alternative is to use the electrophysiological properties of recorded neurons as they capture some of the variability of the transcriptomic profile (Bomkamp et al., 2019). This approach obviates the need for genetic manipulations to identify such types. Simple electrophysiological features extracted from a neuron's EAPs and spiking activity are typically utilized to identify putative cell-types (Gouwens et al., 2019). Automated methods including EAP-specific methods (Lee et al., 2021) and spiking activity-based methods (Schneider et al., 2023a) are an improvement in comparison to manual features. Most recently, multi-modal cell-type classification methods including PhysMAP (Lee et al., 2024) and a VAE-based algorithm (Beau et al., 2024) have been introduced which make use of multiple physiological modalities such as the EAP, activity, or peri-stimulus time histogram (PSTH) of a recorded neuron.

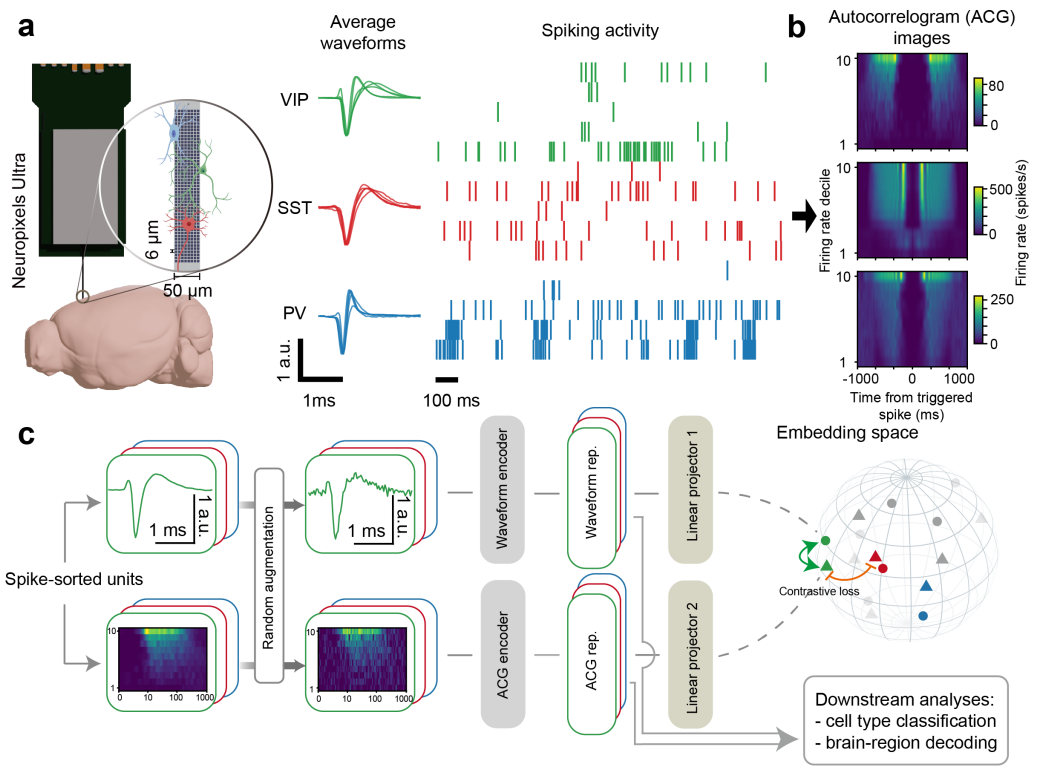

Figure 1: **Schematic illustration of NEMO.** (a) Neuropixels Ultra (Ye et al., 2023b) recordings capture activity from many different cell-types which have distinct extracellular action potentials (EAPs) and spiking activity. We present waveform and spiking activity snippets from five example neurons for each cell-type. (b) We transform the spiking activity of each neuron into a compact autocorrelogram (ACG) image from (Beau et al., 2024) that accounts for variations in the firing rate (see Section 4.1) (c) NEMO utilizes a CLIP-based learning strategy where an EAP encoder and a ACG image encoder are trained to learn an embedding which pushes together randomly augmented EAPs and ACG image from the same neuron while keeping different neurons separate. The learned representations can be utilized for downstream tasks (with or without fine-tuning) such as cell-type and brain-region classification. Schematic created using BioRender.com.

2.3 Brain region classification

Brain region classification consists of predicting the location of a neuron or electrode based on the recorded physiological features (Steinmetz et al., 2018). Rather than predicting a 3D location, the task is to classify the brain region a neuron or electrode occupies, which can be estimated using post-insertion localization via histology (Sunkin et al., 2012). Brain region classification is an important task for understanding fundamental differences in physiology between brain areas as well as for targeting regions that are hard to hit via insertion. Most importantly, brain region classification can provide a real-time estimate of the probe's location in the brain during experiments, significantly increasing the success rate of hitting the target location. Additionally, insertions in primates and human subjects often lack histological information, instead relying on the experimental heuristics that lack standardization between laboratories. As this task is relatively new, only simple features of the EAPs have been utilized for classification (Jia et al., 2019; Tolossa et al., 2024). To our knowledge, no automated feature extraction methods have been utilized for brain region classification.

3 Datasets

For cell-type classification, we utilize an opto-tagged dataset gathered with Neuropixels Ultra probes (NP Ultra; Ye et al., 2023b). For brain region classification, we use a dataset gathered by the IBL: a brain-wide map of neural activity of mice performing a decision-making task (IBL et al., 2023).

NP Ultra opto-tagged mouse data. This dataset consists of NP Ultra recordings of spontaneous activity from the visual cortex of mice. Opto-tagging is utilized to label 477 ground-truth neurons with three distinct cell-types. The ground-truth neurons are composed of 243 parvalbumin (PV), 116 somatostatin (SST), and 118 vasoactive intestinal peptide cells (VIP). There are also 8699 unlabelled neurons that we can utilize for pretraining.

IBL Brain-wide Map. This dataset consists of Neuropixels recordings from animals performing a decision-making task. Each neuron is annotated with the brain region where it is located. We utilize 675 insertions from 444 animals (37017 units). Each brain is parcellated with 10 brain atlas annotations, dividing the atlas into 10 broad areas: isocortex, olfactory areas (OLF), cortical subplate (CTXsp), cerebral nuclei (CNU), thalamus (TH), hypothalamus (HY), midbrain (MB), hindbrain (HB), cerebellum (CB) and hippocampal formation (HPF). We divide this dataset into a training, validation, and testing set by insertion such that we can evaluate each model on heldout insertions.

4 NEMO

We introduce **N**euronal **E**mbeddings via **M**ultim**O**dal contrastive learning (NEMO) which learns a multimodal embedding of neurophysiological data. To extract representations from multiple modalities, we utilize Contrastive Language-Image Pretraining (CLIP; Radford et al., 2021). CLIP uses a contrastive objective to learn a joint representation of images and captions. For NEMO, we utilize the same objective but with modality-specific data augmentations and encoders (see Figure 1c).

4.1 PREPREPROCESSING

We construct a paired dataset of spiking activity and EAPs for all recorded neurons. Using the open-source Python package NeuroPyxels (Beau et al., 2021), we computed an autocorrelogram (ACG) image for each neuron by smoothing the spiking activity with a 250-ms width boxcar filter, dividing the firing rate distribution into 10 deciles, and then building ACGs for each decile (see Figure 1b). This ACG image is a useful representation because the activity autocorrelations of a neuron can change as a function of its firing rate. By computing ACGs for each firing rate decile, the ACG image will account for firing rate dependent variations in the autocorrelations, allowing for comparisons between different areas of the brain, behavioral contexts, and animals (Beau et al., 2024). For the EAPs, we construct a 'template' waveform which is the mean of approximately 500 recorded waveforms for that neuron. For all experiments in the main text, we restrict the template to one channel with maximal amplitude. For multi-channel template results, see Supplement F.

4.2 DATA AUGMENTATIONS

Previous work on contrastive learning for spiking activity utilizes data augmentations including sparse multiplicative noise (pepper noise), Gaussian noise, and temporal jitter (Azabou et al., 2021). As it is computationally expensive to construct ACG images for each batch during training, we instead design augmentations directly for the ACG images rather than the original spiking data. Our augmentations include temporal Gaussian smoothing, temporal jitter, amplitude scaling, additive Gaussian noise, and multiplicative pepper noise (see Supplemental B for more details and Supplementary Figure 14 for an ablation). For our templates, we use additive Gaussian noise as our only augmentation. For a description of multi-channel template augmentations, see Supplement F.

4.3 ENCODERS

We employ separate encoders for each electrophysiological modality. For the ACG image encoder, we use a version of the convolutional architecture introduced in (Beau et al., 2024) with 2 layers and Gaussian Error Linear Units (GeLU) (Hendrycks & Gimpel, 2016). For the waveform encoder, we use a 2 layer multilayer perceptron (MLP) with GeLU units. The representation sizes are 200 dimensional and 300 dimensional for the ACG image encoder and the waveform encoder, respectively. We set the projection size to be 512. For details about hyperparameters, see Supplement B.

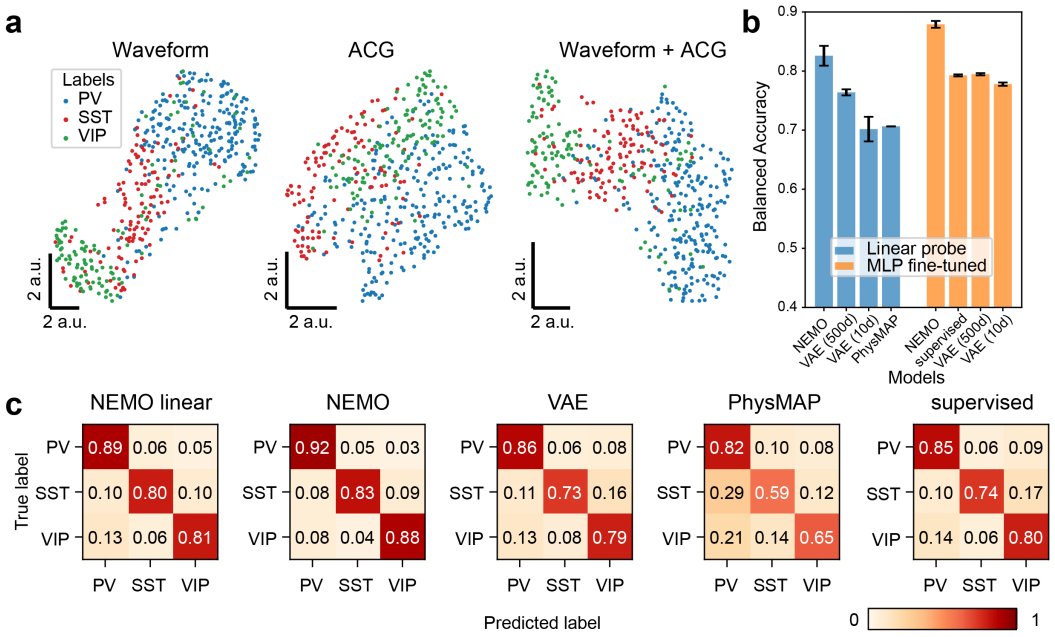

Figure 2: **Comparing NEMO to baseline models on the NP Ultra opto-tagged dataset.** (a) UMAP visualization of the pretrained NEMO representations of unseen opto-tagged visual cortex units, colored by different cell-types. Neurons of the same class form clusters, particularly when combined modalities are used. (b) Balanced accuracy and (c) Confusion matrices for the NP Ultra classification results, normalized by ground truth label and averaged across 5 random seeds. NEMO outperforms the other embedding methods by a significant margin across all cell-types. Surprisingly, NEMO outperforms all other methods even when just using a linear classifier on the embeddings.

4.4 Contrastive Objective

We utilize the contrastive objective defined in CLIP. Let z_{acg} and z_{wf} be the L2 normalized projections of each modality. For a batch B, the objective is as follows,

$$\mathcal{L} = -\frac{1}{2|B|}\sum_{i=1}^{|B|}\left[\log\frac{\exp(z_{\text{acg}_i}\cdot z_{\text{wf}_i}/\tau)}{\sum_{j=1}^{|B|}\exp(z_{\text{acg}_i}\cdot z_{\text{wf}_j}/\tau)} + \log\frac{\exp(z_{\text{acg}_i}\cdot z_{\text{wf}_i}/\tau)}{\sum_{j=1}^{|B|}\exp(z_{\text{acg}_j}\cdot z_{\text{wf}_i}/\tau)}\right] \tag{1}$$

where τ is a temperature parameter which we fix during training. The objective function encourages the model to correctly match z_{acg_i} with its corresponding z_{wf_i}, and vice versa, over all other possible pairs in the batch. This loss can easily be extended to more than two modalities including PSTHs.

4.5 Single-neuron and multi-neuron brain region classification

Brain region classification is a new problem for electrophysiological datasets that requires novel classification schemes. We develop two classification schemes for our evaluation: a single-neuron and multi-neuron classifier. For our single-neuron classifier, we predict the brain area for each neuron independently using its embedding. For our multi-neuron classifier, we predict the brain region for each 20 micron bin along the depth of the probe by ensembling the predictions of nearby neurons within a 60-micron radius (i.e., averaging the logits of the single-neuron model) as shown in Figure 3a (ii). When more than five neurons fall within this range, only the nearest five are selected.

5 Experimental Setup

5.1 Baselines

For our baselines, we compare against current state-of-the-art multimodal cell-type embedding methods and a fully supervised method. For the multimodal cell-type embedding methods, we

Table 1: **Cell-type classification for the NP-Ultra dataset.** The accuracy and F1-scores are reported for three conditions: (i) a linear layer and (ii) MLP on top of the frozen pretrained representations (for VAE and NEMO), and (iii) after MLP finetuning.

Model	**Linear**		**MLP**		**MLP fine-tuned**	
	Acc	**F1**	**Acc**	**F1**	**Acc**	**F1**
Supervised	N/A	N/A	N/A	N/A	0.79 ± 0.00	0.79 ± 0.00
PhysMAP	0.71 ± 0.00[1]	0.69 ± 0.00 [1]	N/A	N/A	N/A	N/A
VAE (10d latent)	0.70 ± 0.02	0.70 ± 0.02	0.68 ± 0.01	0.68 ± 0.01	0.78 ± 0.00	0.77 ± 0.00
VAE (500d rep)	0.76 ± 0.01	0.76 ± 0.01	0.77 ± 0.00	0.76 ± 0.00	0.79 ± 0.00	0.79 ± 0.00
NEMO (500d rep)	$\mathbf{0.83 \pm 0.01}$	$\mathbf{0.83 \pm 0.01}$	$\mathbf{0.84 \pm 0.01}$	$\mathbf{0.84 \pm 0.01}$	$\mathbf{0.88 \pm 0.01}$	$\mathbf{0.88 \pm 0.01}$

compare to PhysMAP (Lee et al., 2024) and a VAE-based method (Beau et al., 2024). For fair comparison, we utilize the same encoder architectures for NEMO and the VAE-based method. We include two versions of the VAE baseline: (1) the latent space (10D) is used to predict cell-type or brain region (from Beau et al. (2024)), or (2) the layer before the latent space (500D) is used to predict cell-type or brain region. For the supervised baseline, we again use the same encoder architectures as NEMO. For training NEMO, we use an early stopping strategy which utilizes validation data. For the VAE-based method, we use the training scheme introduced in (Beau et al., 2024). We fix the hyperparameters for all methods across both datasets. For more details about baselines, training, and hyperparameters, see Supplements B and E.

5.2 Evaluation

For both NEMO and the VAE-based method, the representations from the ACG image and EAPs are concatenated together before classification or fine-tuning. We apply three classification schemes for evaluation including (1) freezing the model and training a linear classifier on the final layer, (2) freezing the model and training a MLP-based classifier on the final layer, (3) fine-tuning both the original model and a MLP-based classifier on the final layer. To ensure balanced training data, we implement dataset resampling prior to fitting the linear classifier. The only method that cannot be fine-tuned is PhysMAP, as it is UMAP-based and therefore non-differentiable. For PhysMAP comparisons, we utilize the weighted graph approach with alignment mapping provided in (Lee et al., 2024) for all comparisons. The macro-averaged F1 score, calculated as the unweighted mean of F1 scores for each class, and balanced accuracy, which measures average accuracy per class, are selected as the primary metrics for our classification tasks. This choice addresses data imbalance, a common issue in opto-tagged datasets and brain region classification. For additional details about baseline hyperparameters, see Supplement B.

Table 2: **single-neuron brain region classification for the IBL dataset.** The accuracy and F1-scores are reported for three conditions: (i) a linear layer and (ii) MLP on top of the frozen pretrained representations (for VAE and NEMO), and (iii) after MLP finetuning. We only ran PhysMAP one time as it is deterministic.

Model	**Linear**		**MLP**		**MLP fine-tuned**	
	Acc	**F1**	**Acc**	**F1**	**Acc**	**F1**
Supervised	N/A	N/A	N/A	N/A	0.45 ± 0.01	0.42 ± 0.01
PhysMAP	0.31[1]	0.27[1]	N/A	N/A	N/A	N/A
VAE	0.33 ± 0.00	0.29 ± 0.00	0.40 ± 0.01	0.37 ± 0.00	0.43 ± 0.00	0.40 ± 0.00
NEMO	$\mathbf{0.42 \pm 0.00}$	$\mathbf{0.40 \pm 0.00}$	$\mathbf{0.45 \pm 0.01}$	$\mathbf{0.42 \pm 0.00}$	$\mathbf{0.47 \pm 0.01}$	$\mathbf{0.44 \pm 0.01}$

5.3 Experiments

NP Ultra opto-tagged dataset. For the NP Ultra dataset, we pretrain NEMO and the VAE-based method on 8699 unlabelled neurons. This pretraining strategy is important for reducing overfitting to the small quantity of labeled cell-types. To evaluate each model after pretraining, we perform the three evaluation schemes introduced in Section 5.2: freezing + linear classifier, freezing + MLP classifier, and full end-to-end finetuning. For PhysMAP, we utilize the anchor alignment technique

[1] We utilize PhysMAP's anchor alignment technique to evaluate its performance (not a linear classifier).

Table 3: **multi-neuron brain region classification for the IBL dataset.** The accuracy and F1-scores for brain region classification are reported for three conditions: (i) a linear layer and (ii) MLP on top of the frozen pretrained representations (for VAE and NEMO), and (iii) after MLP finetuning.

Model	**Linear**		**MLP**		**MLP fine-tuned**	
	Acc	**F1**	**Acc**	**F1**	**Acc**	**F1**
Supervised	N/A	N/A	N/A	N/A	0.50 ± 0.00	0.48 ± 0.01
VAE	0.36 ± 0.00	0.32 ± 0.00	0.45 ± 0.00	0.42 ± 0.00	0.48 ± 0.00	0.46 ± 0.00
NEMO	$\mathbf{0.48 \pm 0.00}$	$\mathbf{0.45 \pm 0.00}$	$\mathbf{0.50 \pm 0.00}$	$\mathbf{0.48 \pm 0.00}$	$\mathbf{0.51 \pm 0.00}$	$\mathbf{0.50 \pm 0.00}$

introduced by (Lee et al., 2024). For all methods and evaluation schemes, we perform a 5-fold cross-validation with 10 repeats to evaluate each model.

IBL Brain-wide Map. For the IBL dataset, we randomly divide all insertions (i.e., Neuropixels recordings) into a 70-10-20 split to create a training, validation, and test set for each method. We then pretrain NEMO and the VAE-based method on all neurons in the training split. We then perform the three evaluation schemes introduced in Section 5. For PhysMAP, we utilize the anchor alignment technique. We train both a single-neuron and multi-neuron classifier using the representations learned by NEMO and the VAE-based method. For PhysMAP, we only evaluate the single-neuron classifier. We compute the average and standard deviation of the metrics using five random seeds.

6 Results

6.1 Classification

NP Ultra cell-type classifier. The results for the NP Ultra opto-tagged dataset are shown in Table 1 and Figure 2. For all three evaluation schemes, NEMO achieves the highest macro-averaged F1 score and balanced accuracy by a significant margin. Surprisingly, the VAE-based method, even with MLP-based fine-tuning, is outperformed by a frozen NEMO model with a linear decoder (by $\sim 5\%$) . With MLP fine-tuning, NEMO reaches .88 F1 score and balanced accuracy which is an $\sim 11\%$ improvement over the baselines. For the VAE-based method, we found that utilizing the 500D representations *before* the latent space performed better than using the 10D latent space and also outperformed a VAE trained with a 500D latent space so we included this as a baseline. Except for NEMO, all baseline models struggle to differentiate VIP and SST cells as they have the fewest labels and the VIP cells display bimodality in their waveform distribution. These results demonstrate that NEMO is an accurate method for cell-type classification in visual cortical microcircuits.

IBL single-neuron region classifier. We then aim to investigate how much relevant information NEMO extracts from each neuron about its anatomical location, i.e., brain region. We investigate this by training classifiers that use single neuron features to identify anatomical regions for the IBL dataset (see Table 2 and Figure 3b for results). We again find that NEMO outperforms all other methods using both linear and MLP-based classification schemes. While NEMO outperforms the supervised baseline, we find that the VAE-based method performs worse than a fully supervised model trained on the same raw data. Without end-to-end fine-tuning, NEMO with an MLP classification head is already on par with the supervised MLP. NEMO's success with both the linear and MLP classifier with frozen encoder weights indicates that NEMO is able to extract a region-discriminative representation of neurons without additional fine-tuning. This representation can be further improved by fine-tuning NEMO and the classifier end-to-end. The confusion matrix for PhysMAP is shown in Supplementary Figure 8.

IBL multi-neuron region classifier. We further investigate whether combining information from multiple neurons at each location can improve brain region classification. We use the nearest-neurons ensembling approach as described in 4.5 and shown in Figure 3a. Averaging the logits of predictions from single neurons improves classification performance over the single-neuron model. NEMO still has the best region classification performance (see Table 3 and Figure 3c for results).

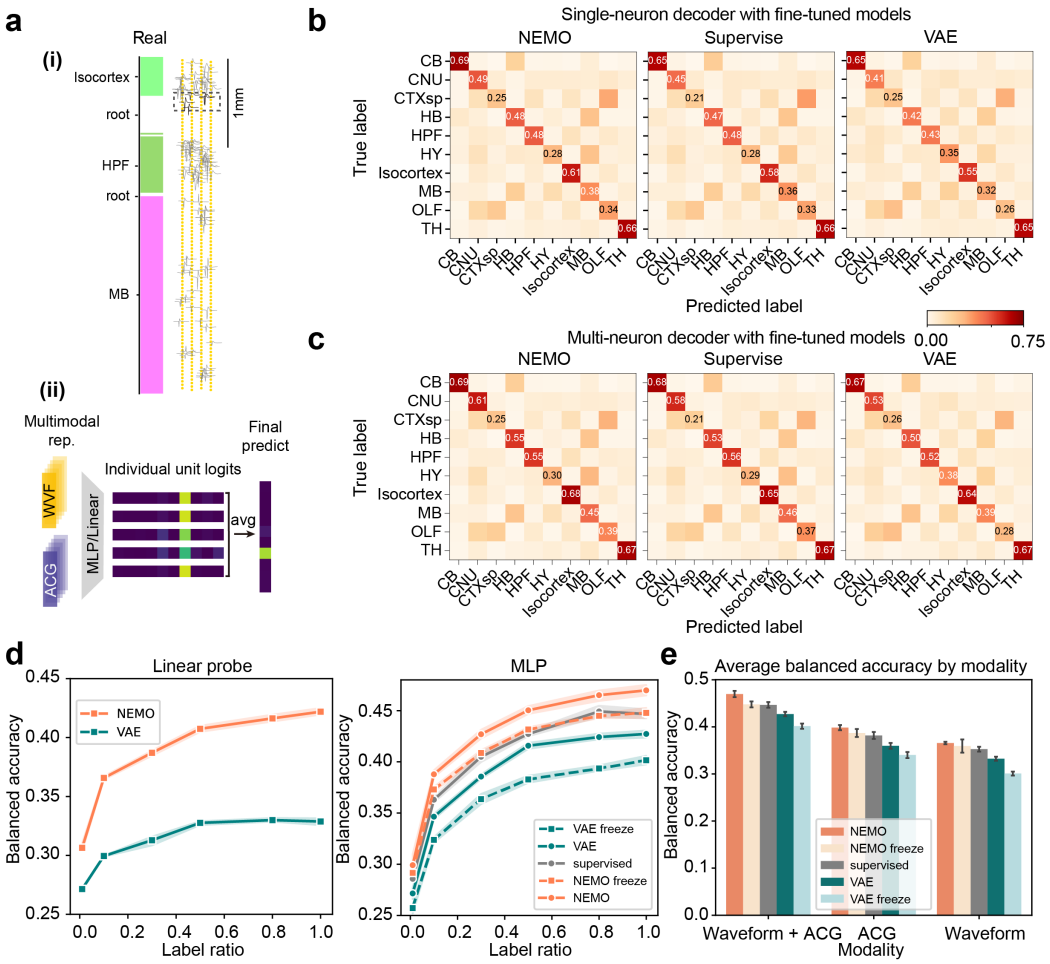

Figure 3: **Results for NEMO on the IBL brain region classification task.** (a) Schematic for multi-neuron classifier. (i) At each depth, the neurons within 60 µm were used to classify the anatomical region. Only the nearest 5 neurons were selected if there were more than 5 neurons within that range. (ii) For logits averaging, single-neuron classifier logits are predicted using a linear model/MLP trained on the representations of our two physiological modalities. The final prediction is based on the average of the individual logits. (b) Confusion matrices for the single-neuron region classifier with fine-tuned NEMO pretrained encoders, the fully supervised model, and with VAE pretrained encoders, averaged across 5 runs. (c) Confusion matrices for the multi-neuron region classifier, averaged across 5 runs. (d) Single neuron balanced accuracy with linear classifier and the MLP head for each model trained/fine-tuned with different label ratios. (e) Single-neuron MLP-classification balanced accuracy for each modality separately and for the combined representation.

6.2 Clustering

We next examine the clusterability of NEMO representations for the IBL Brain-wide map. We followed the clustering strategy used in Lee et al. (2021) by running Louvain clustering on a UMAP graph constructed from the representations extracted by NEMO from the IBL training neurons. We adjusted two main settings: the neighborhood size in UMAP and the resolution in Louvain clustering. We selected these parameters by maximizing the modularity index, which had the effect of minimizing the number of resulting clusters (Figure 4c). The clustering results relative to the region labels are presented in Figures 4a and b. The UMAP visualization of the NEMO representations, colored by region label, demonstrates that the regions are separable in the representation space. Notably, there is a distinct separation of thalamic neurons from other regions, along with an isolated cluster of cerebellar neurons. Neurons from other regions are also well organized by region labels within the NEMO representation space, allowing for their clustering into several distinct clusters. Additionally, overlaying the neurons colored by their cluster IDs onto their anatomical

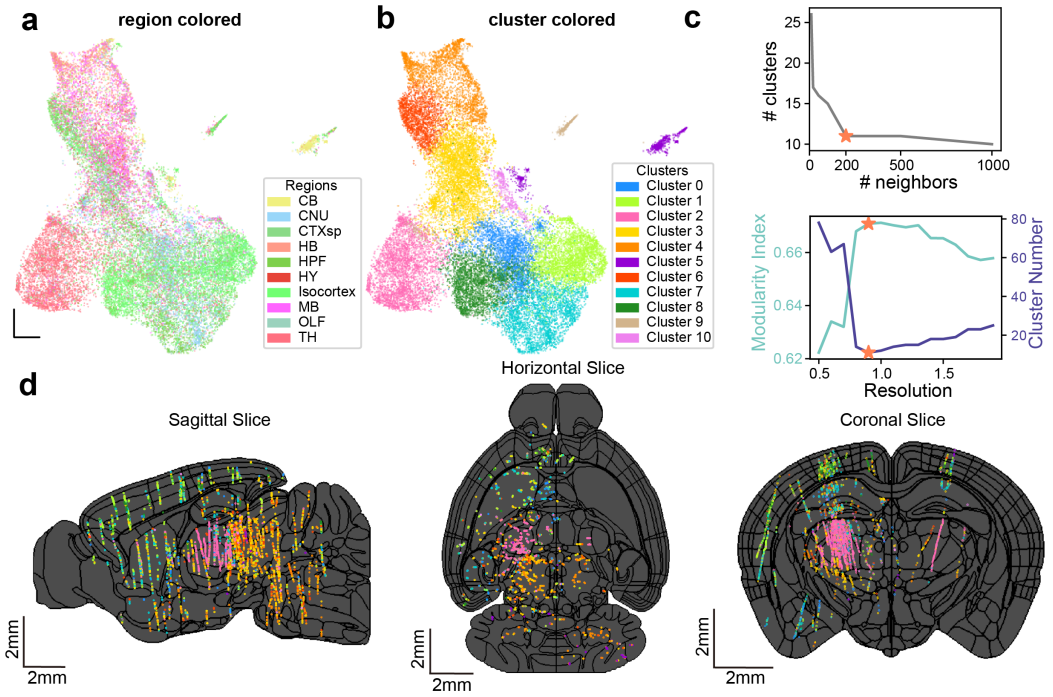

Figure 4: **IBL neuron clustering using NEMO pretraining.** (a) A UMAP visualization of the representations that NEMO extracts from the training data colored by anatomical brain region. (b) The same UMAP as shown in (a) but instead colored by cluster labels using a graph-based approach (Louvain clustering). (c) We tuned the neighborhood size in UMAP and the resolution for the clustering. These parameters were selected by maximizing the modularity index which minimized the number of clusters. (d) 2D brain slices across three brain views with the location of individual neurons colored using the cluster IDs shown in (b). The black lines show the region boundaries of the Allen mouse atlas (Wang et al., 2020). The cluster distribution found using NEMO is closely correlated with the anatomical regions and is consistent across insertions from different labs.

locations (Figure 4) reveals a cluster distribution closely correlated with anatomical regions which is consistent across insertions from different labs (Supplementary Figure 10). We find that clustering NEMO's representations leads to a more region-selective clustering than when we use the raw features directly (Supplementary Figures 11 and 12). These results suggest that NEMO is able to extract features that capture the electrophysiological diversity across regions.

6.3 Ablations

Label ratio sweep. We hypothesize that NEMO requires less labeled data for fine-tuning to achieve comparable performance to other models for brain region classification due to the contrastive pretraining. To test this, we conducted a label ratio sweep with our single-neuron region classifiers. We trained the linear classifier and the MLP classifier under two conditions: with frozen weights and with full end-to-end fine-tuning. We use 1%, 10%, 30%, 50%, 80%, and 100% of the labeled data for this experiment. The accuracy results are depicted in Figure 3d (for F1, see Supplementary Figure 5). The NEMO fine-tuned model outperforms all other methods for every label ratio. For the linear model, using just 10% of the training labels, NEMO achieves superior performance compared to using the full training set for the VAE. Similarly, NEMO paired with the MLP-classifier shows impressive results; with only 50% of the labeled data, the NEMO outperforms the supervised MLP. With 30% of the labels, NEMO exceeds the performance of the VAE model trained with all labels.

Single modality classifier. We next explore the additional region-relevant information gained by combining both modalities instead of just one, and whether NEMO enhances information etraction from each modality by aligning the embeddings of the two. We compared the classification performance of the MLP classifier with encoder weights frozen and end-to-end fine-tuned, across all models using: 1) waveforms only 2) ACGs only 3) combining waveforms and ACGs. Brain region

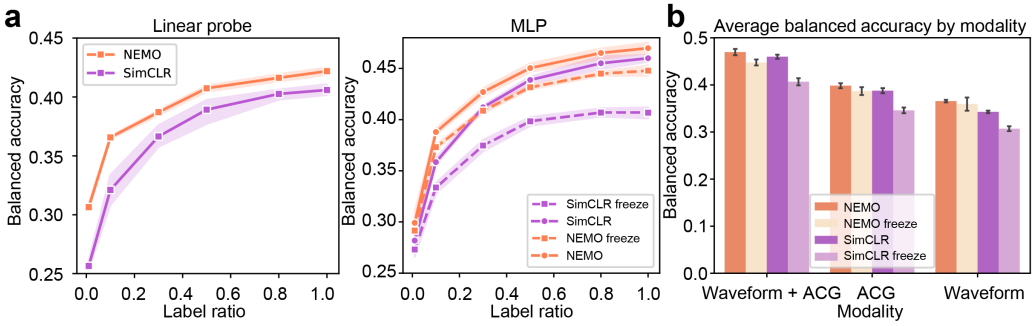

Figure 5: **Ablating joint vs. independent learning for NEMO.** To evaluate the importance of learning a shared representation between modalities, we train a version of NEMO on the IBL brain classification task where each modality is independently embedded using SimCLR. We refer to this method as SimCLR. (a) Across all label ratios and classifiers, we find that NEMO trained with CLIP outperforms the SimCLR version. (b) NEMO trained with CLIP also extracts more informative representations for each modality (especially waveforms) than when training with SimCLR.

classification balanced accuracies are shown in Figure 3e (for F1, see Supplementary Figure 5). We found that bimodal models, in general, outperform unimodal models, indicating that combining both modalities provides extra information on the anatomical location of neurons. We also find that the NEMO fine-tuned model delivers the best performance. This demonstrates that NEMO improves the information extracted from a single modality by leveraging information from the other modality.

Joint vs. independent learning for NEMO. To ablate the importance of learning a shared representation of each modality, we train a version of NEMO where we independently learn an embedding for each modality using a unimodal contrastive method, SimCLR (Chen et al., 2020). The results of the IBL brain region classification task are shown in Figure 5a where NEMO trained with CLIP outperforms NEMO trained with SimCLR for all label ratios and classification methods. NEMO trained with CLIP is also able to extract more informative representations from each modality (especially for waveforms) as shown in Figure 5b. These results demonstrate that learning a shared representation of the two modalities is important for the downstream performance of NEMO. Supplementary Table 9 shows that joint training with CLIP leads to a modest improvement over SimCLR for cell-type classification of the NP Ultra dataset.

7 Discussion

In this work, we proposed NEMO, a pretraining framework for electrophysiological data that utilizes multi-modal contrastive learning. We demonstrate that NEMO is able to extract informative representations for cell-type and brain region classification with minimal fine-tuning. This capability is particularly valuable in neuroscience research where ground truth data, such as opto-tagged cells, are costly and labor-intensive to acquire (or in some cases even impossible to acquire, for example in human datasets). Our work has some limitations. Firstly, we primarily focus on shared information between two modalities, assuming this is most informative for identifying cell identity or anatomical location. While our analysis using a single modality classifier indicates that our model captures modality-exclusive information, it would be beneficial to distinguish between the shared and private information within each modality (Liu et al., 2024; Liang et al., 2024). Additionally, NEMO utilizes the activity of each neuron independently. Recent work has demonstrated that there is additional information in neuron correlations that can help distinguish different cell-types (Mi et al., 2023).

NEMO opens up several promising avenues for future research. Our framework can be adapted for studies of peripheral nervous systems, such as the retina (Wu et al., 2023). NEMO can also be combined with RNA sequencing to find features that are shared between RNA and electrophysiological data (Li et al., 2023). It will also be possible to correlate the cell-types discovered using NEMO with animal behavior to characterize their functional properties. Finally, we imagine that the representations found by NEMO can be integrated with current large-scale pretraining approaches for neural population activity to provide additional cell-type information which could improve generalizability to unseen sessions or animals (Azabou et al., 2023; Ye et al., 2023a; Zhang et al., 2024).

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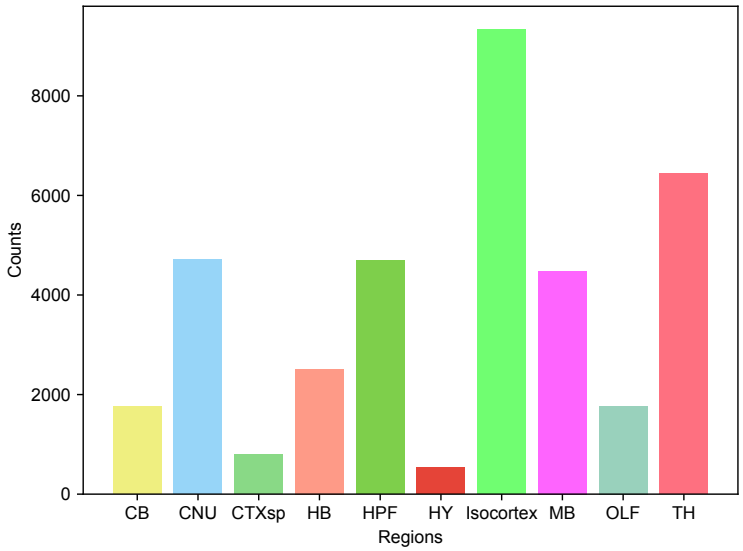
A IBL BRAIN-WIDE-MAP DATASET

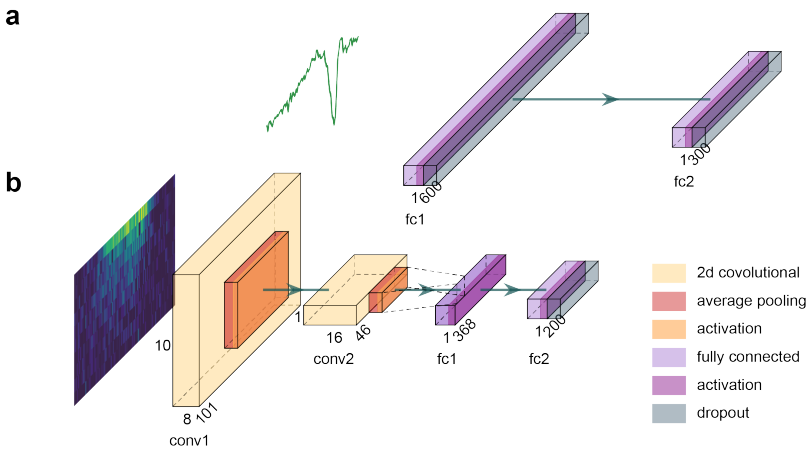
The full IBL Brain-Wide-Map dataset contains 699 insertions. Among those, only 675 successfully went through the pre-processing, with 37017 units that pass the IBL spike-sorting quality control. The region distribution of those units are shown in Supplementary Figure 1.



Supplementary Figure 1: Region distribution of the IBL Brain-Wide-Map dataset. The dataset is very imbalanced and has a small number of units in hypothalamus and cortical subplates.

B BASELINES AND HYPERPARAMETERS

B.1 MODEL ARCHITECTURE



Supplementary Figure 2: Encoder architecture illustration: a) A two-layer multilayer perceptron for waveform encoding, and b) A convolutional neural network for ACG image encoding.

B.2 MODEL PARAMETERS

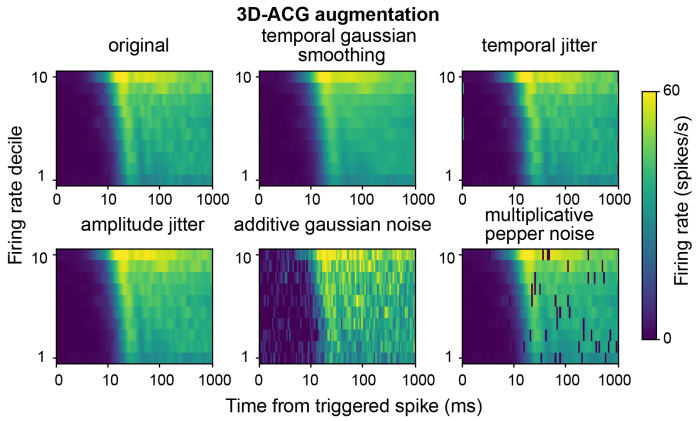
We apply the model augmentations in table 1 on the training data during NEMO. p is the probability an augmentation gets applied to an instance. We use the Adam optimizer with learning rate 0.0005 and Cosine Annealing scheduler with $T_0 = 20$. Other model hyperparameters are in table 2.

Supplementary Table 1: Model Augmentations

Augmentation	**Description**	**Type**	**p**
Gaussian noise	Gaussian noise with mean 0 and std 0.1×std of WVF	WVF	0.3
Temporal Gaussian smoothing	Smooths an ACG using a Gaussian filter along the temporal axis with $\sigma = 2$	ACG	0.5
Temporal jittering	Jitters the temporal axis of an ACG by a random integer between -3 and 3 inclusive	ACG	0.5
Amplitude scaling	Scales the amplitude of an ACG by a random number between 0.9 and 1.1	ACG	0.5
Additive Gaussian noise	Adds Gaussian noise with mean 0 and std 0.1×maximum of ACG	ACG	0.5
Multiplicative pepper noise	Each value in the ACG has a 5% of being set to 0	ACG	0.5

B.3 MLP FINE-TUNING

For cell-type classification, we fix the hyperparameters of the MLP classifier for all methods to those used in Beau et al. (2024). This is because we have very few labels for evaluation and it is challenging to hold out any for hyperparameter tuning. For the brain region classification experiments, we tune the optimizer, learning rate, model size and dropout using the validation set of insertions.



Supplementary Figure 3: Illustration of the augmentations on spiking activity ACGs used by NEMO.

Supplementary Table 2: Model Hyperparameters

Parameter	**Description**	**Value**
Epochs	Numbers of epochs to run	6000
Batch Size	Number of items processed in a single operation	1024
Learning Rate	Learing rate of the model	0.0005
Log every n steps	Save model and cross validate results every n epochs	100
dim_embed	Latent dimension size	512
Temperature	τ in the formula for contrastive loss	0.5
ACG dropout	Dropout probability in ACG encoder	0.2
Waveform dropout	Dropout probability in waveform encoder	0.1

B.4 SimCLR-based NEMO

For the SimCLR-based NEMO, we randomly apply the same set of augmentation methods to each data point, generating two correlated augmented views. We then train each encoder and a linear projection layer to maximize agreement using contrastive learning. Compared to other methods, we use stronger augmentations for SimCLR. This is because SimCLR focuses on the differences between augmentations from the same data point, whereas CLIP-based learning strategies compare augmentations across different modalities. Stronger augmentations improved SimCLR's performance.

Both methods use contrastive learning to maximize similarity between augmented views of the same neuron, while keeping views of different neurons distinct. The main difference between the SimCLR-based method and the CLIP-based method is that CLIP is multimodal and SimCLR is unimodal. For the SimCLR-based method, we use the contrastive objective from (Chen et al., 2020). Let z_{view1} and z_{view2} be the normalized projections of two augmented views from the same modality. For a batch B, the objective is as follows:

$$\mathcal{L} = -\frac{1}{2|B|}\sum_{i=1}^{|B|}[\log \frac{\exp(z_{\text{view1}_i} \cdot z_{\text{view2}_i}/\tau)}{\sum_{j=1}^{|B|} \mathbb{1}_{\{j\neq i\}} \exp(z_{\text{view1}_i} \cdot z_{\text{view2}_j}/\tau) + \sum_{j=1}^{|B|} \mathbb{1}_{\{j\neq i\}} \exp(z_{\text{view1}_i} \cdot z_{\text{view1}_j}/\tau)} + \log \frac{\exp(z_{\text{view2}_i} \cdot z_{\text{view1}_i}/\tau)}{\sum_{j=1}^{|B|} \mathbb{1}_{\{j\neq i\}} \exp(z_{\text{view2}_i} \cdot z_{\text{view1}_j}/\tau) + \sum_{j=1}^{|B|} \mathbb{1}_{\{j\neq i\}} \exp(z_{\text{view2}_i} \cdot z_{\text{view2}_j}/\tau)}] \tag{2}$$

For SimCLR, we use the same types of augmentation as CLIP with increased strength: for the EAPs, we applied Gaussian noise with standard deviation 1 with probability 0.5, rather 0.1 standard with a probability of than a 0.3. For the ACG images, we keep the augmentation methods consistent across all models.

In our SimCLR-based method, we use the pretrained encoder to obtain separate embeddings for each modality, which are then combined to train downstream tasks such as cell-type and brain region classification.

C Augmentation Ablation Study

D Different projection techniques for SimCLR

We explored different projection techniques for SimCLR. One approach was to use a SimCLR model with a smaller projection size than the representation, reducing the projection size from 512 to 128. Another approach was to use a 2-layer MLP with a 512-dimensional hidden layer and a 128-dimensional projection size instead of a linear projection layer. However, our original linear projector still achieved the highest validation metrics, as shown in Supplementary Table 3. Therefore, we continued to use the original linear projector for all experiments.

Supplementary Table 3: Linear brain region classification for IBL dataset

Model	**Acc**	**F1**
original	0.388	0.338
128-dim linear projection	0.371	0.326
mlp-based projection	0.377	0.328

E Dataset split and validation strategy

Given the limited quantity of ground-truth labels for the NP Ultra dataset, we evaluate the performance of each model using a 5-fold cross-validation of the labeled cells with 10 repeats. Since we do not have a separate validation set while training NEMO, we utilize a nested cross-validation approach to choose an evaluation checkpoint for NEMO. In other words, for each cross-validation

fold, we perform a nested 5-fold cross-validation on the *training* folds with a linear classifier to choose the best checkpoint for NEMO. We then use this NEMO checkpoint to perform the evaluation on the heldout fold of the original 5-fold cross-validation. This checkpoint choosing procedure is done only on the training folds and does not use any information from the testing fold. Without this procedure, we are still able to achieve high performance.

For the IBL dataset, we use a standard 70-10-20 split for the training, validation, and test sets, respectively. During training, we monitor the performance of the model using a linear classifier trained on the training set and validated on the validation set. We then compute our evaluation metrics on the test set using the checkpoint with the highest validation F1.

F MULTICHANNEL

Using multi-channel templates has the potential to be more informative for identifying cell-types and classifying brain regions. In the following subsections, we demonstrate that using multi-channel templates does improve these downstream tasks, however, more work is needed to generalize this paradigm to different probe geometries.

F.1 IBL MULTICHANNEL

For the IBL multichannel experiments, we use 25 channel templates centered on the peak channel if possible (this is hard for edge spikes). We add two additional template augmentations: amplitude jitter (Vishnubhotla et al., 2024) and electrode dropout. Amplitude jitter rescales a channel's amplitude by a uniform value between 0.9 and 1.1 with $p = 0.3$. Electrode dropout zeros out a channel with $p = 0.1$. If all channels are zeroed out, we leave the peak channel. Our model architecture is the same as NEMO with the only difference being the input size of the waveform encoder.

We tune hyperparameters for both the linear and MLP classification models. For the linear model, we tune the inverse of the regularization strength between $1e-5$ and $1e4$ using the python module optuna (Akiba et al., 2019). For the MLP model, we do a grid search over the dropout probability$(0.1, 0.2, 0.3, 0.4)$, learning rate$(1e-4, 1e-5, 1e-6)$, number of layers$(1, 2, 3)$ and layer size$(128, 256, 512)$.

Supplementary Table 4: Multi-channel, single-neuron brain region decoding for the IBL dataset

Model	**Linear**		**MLP**		**MLP fine-tuned**	
	Acc	**F1**	**Acc**	**F1**	**Acc**	**F1**
NEMO	0.42 ± 0.00	0.40 ± 0.00	0.45 ± 0.01	0.42 ± 0.00	0.47 ± 0.01	0.44 ± 0.01
NEMO (25-channel)	$\mathbf{0.45 \pm 0.01}$	$\mathbf{0.42 \pm 0.00}$	$\mathbf{0.46 \pm 0.00}$	$\mathbf{0.43 \pm 0.00}$	$\mathbf{0.48 \pm 0.00}$	$\mathbf{0.45 \pm 0.00}$

Supplementary Table 5: multi-neuron, multi-channel brain region classification for the IBL dataset

Model	**Linear**		**MLP**		**MLP fine-tuned**	
	Acc	**F1**	**Acc**	**F1**	**Acc**	**F1**
NEMO	0.48 ± 0.00	0.45 ± 0.00	0.50 ± 0.00	$\mathbf{0.48 \pm 0.00}$	0.51 ± 0.00	0.50 ± 0.00
NEMO (25-channel)	$\mathbf{0.50 \pm 0.01}$	$\mathbf{0.47 \pm 0.00}$	$\mathbf{0.51 \pm 0.01}$	$\mathbf{0.48 \pm 0.01}$	$\mathbf{0.52 \pm 0.00}$	$\mathbf{0.51 \pm 0.01}$

F.2 NP-ULTRA MULTICHANNEL

For NP-Ultra multichannel, we tested using 9 or 25 channels centered on the peak channel if possible. We use the same augmentations and encoding model architecture as IBL multichannel and we do not finetune the hyperparameters in the MLP or linear model.

G PICKING PARAMETERS FOR IBL UNIT REPRESENTATION CLUSTERING

We used the Python-implemented UMAP package (McInnes et al., 2018b) and the Python-Louvain package (Aynaud, 2020) for our clustering analyses. For our clustering analysis, we aim to find the

Supplementary Table 6: Multichannel cell-type classification for the NP-Ultra dataset

Model	Linear		MLP (5-fold)		MLP fine-tuned(5-fold)	
	Acc	F1	Acc	F1	Acc	F1
NEMO (500d rep)	0.833 ± 0.008	0.833 ± 0.007	0.839 ± 0.007	0.839 ± 0.008	0.878 ± 0.005	0.878 ± 0.005
NEMO (25-channel)	0.853 ± 0.006	0.85 ± 0.006	0.865 ± 0.007	0.862 ± 0.007	0.875 ± 0.008	0.875 ± 0.007
NEMO (9-channel)	**0.854 ± 0.014**	**0.851 ± 0.015**	**0.869 ± 0.003**	**0.865 ± 0.004**	**0.886 ± 0.003**	**0.883 ± 0.004**

most informative clustering with the smallest number of clusters. There are two parameters to tune: 1) the size of local neighborhood used for the UMAP graph manifold approximation resolution that controls the community size in Louvain clustering ($n_{neighbor}$), and 2) the resolution γ that determines the size of the communities. We tuned $n_{neighbor}$ while keeping the resolution to be 1.0 and tracked the final number of clusters. We picked the 'elbow' that has the smallest $n_{neighbor}$ (200). The resolution was chosen by maximizing the modularity index Q of Louvain clustering with $n_{neighbor} = 200$. The modularity index of a graph is defined as:

$$Q = \sum_{c=1}^{n} \left[\frac{L_c}{m} - \gamma \left(\frac{k_c}{2m} \right)^2 \right]$$

where each c represents a community in the graph, m is the number of edges, L_c is the number of intra-community links for community c, k_c is the sum of degrees of the nodes in community c, and γ is the resolution parameter.

H Parameters for classifiers and fine-tuning methods

Supplementary Table 7: Linear probe best hyperparameters

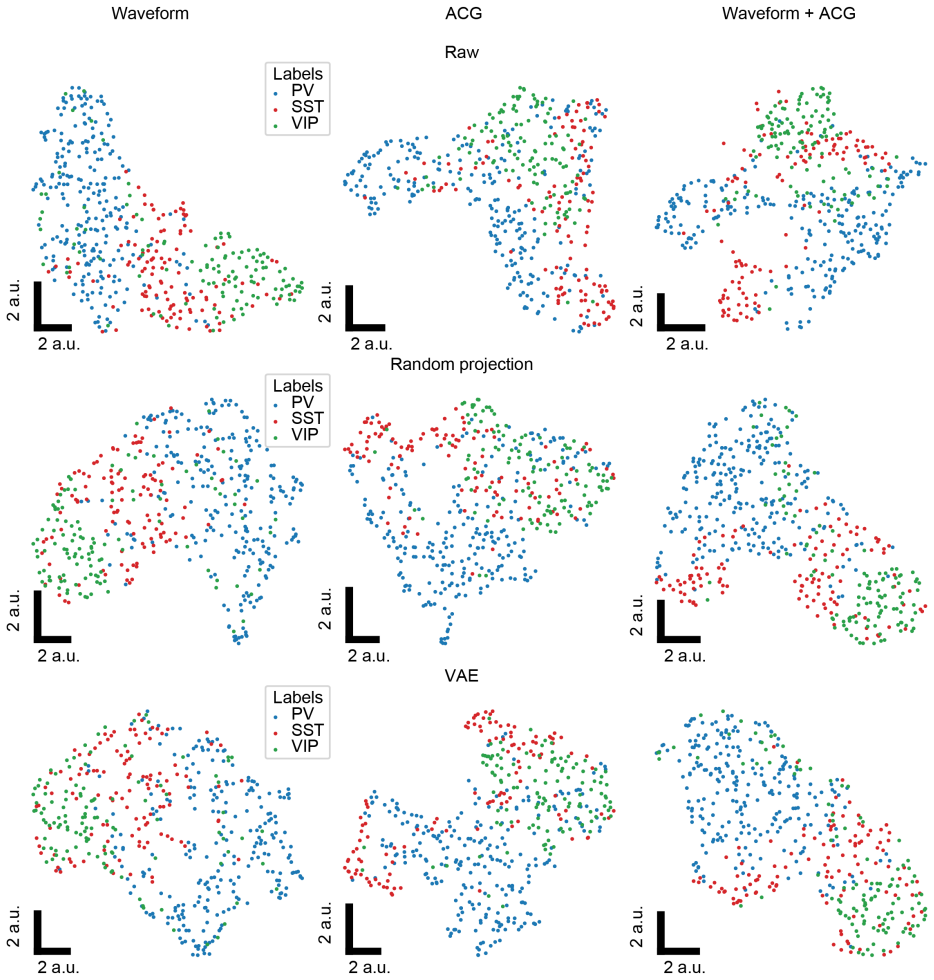
Hyperparameter	Value
max_{iter}	1000
tol	1e-5
NP-Ultra Celltype c	0.02
IBL NEMO joint c	0.02
IBL VAE c	0.001
IBL supervise c	0.001
IBL NEMO independent c	2.5

Supplementary Table 8: IBL MLP hyperparameters

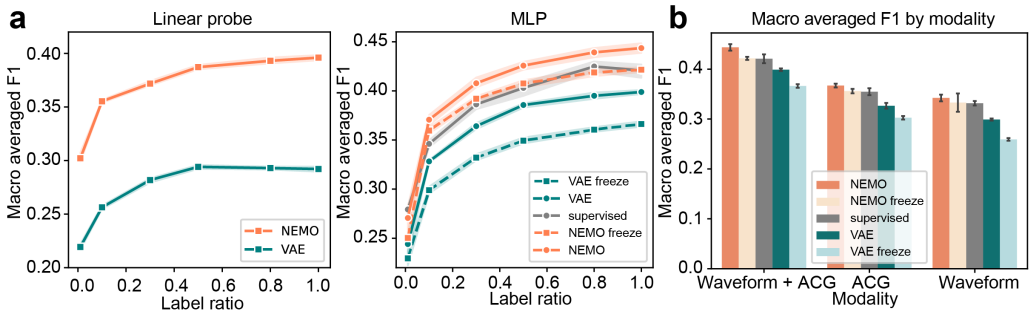
Hyperparameter	Value
n_{layer}	1
layer0 size	256
Dropout rate	0.2
Supervised scheduler	CosineAnnealingWarmRestarts
Supervised T_0	20
Supervised T_{mult}	1
Other models' schedular	StepLR
StepLR $step_{size}$	200
StepLR γ	0.8
Supervised lr	1×10^{-4}
Other models' lr	1×10^{-5}

H.1 UMAP EMBEDDINGS WITH RAW FEATURES, RANDOM PROJECTION AND VAE

We visualize the UMAP embeddings of the raw features, randomly initialized encoder projection and the VAE projection in Supplementary figure 4. We find that NEMO representations are visually more structured.



Supplementary Figure 4: UMAP visualizations with raw features, random projections, and from the VAE. The random projections are representations which are passed through randomly initialized encoders with the same architecture as NEMO.

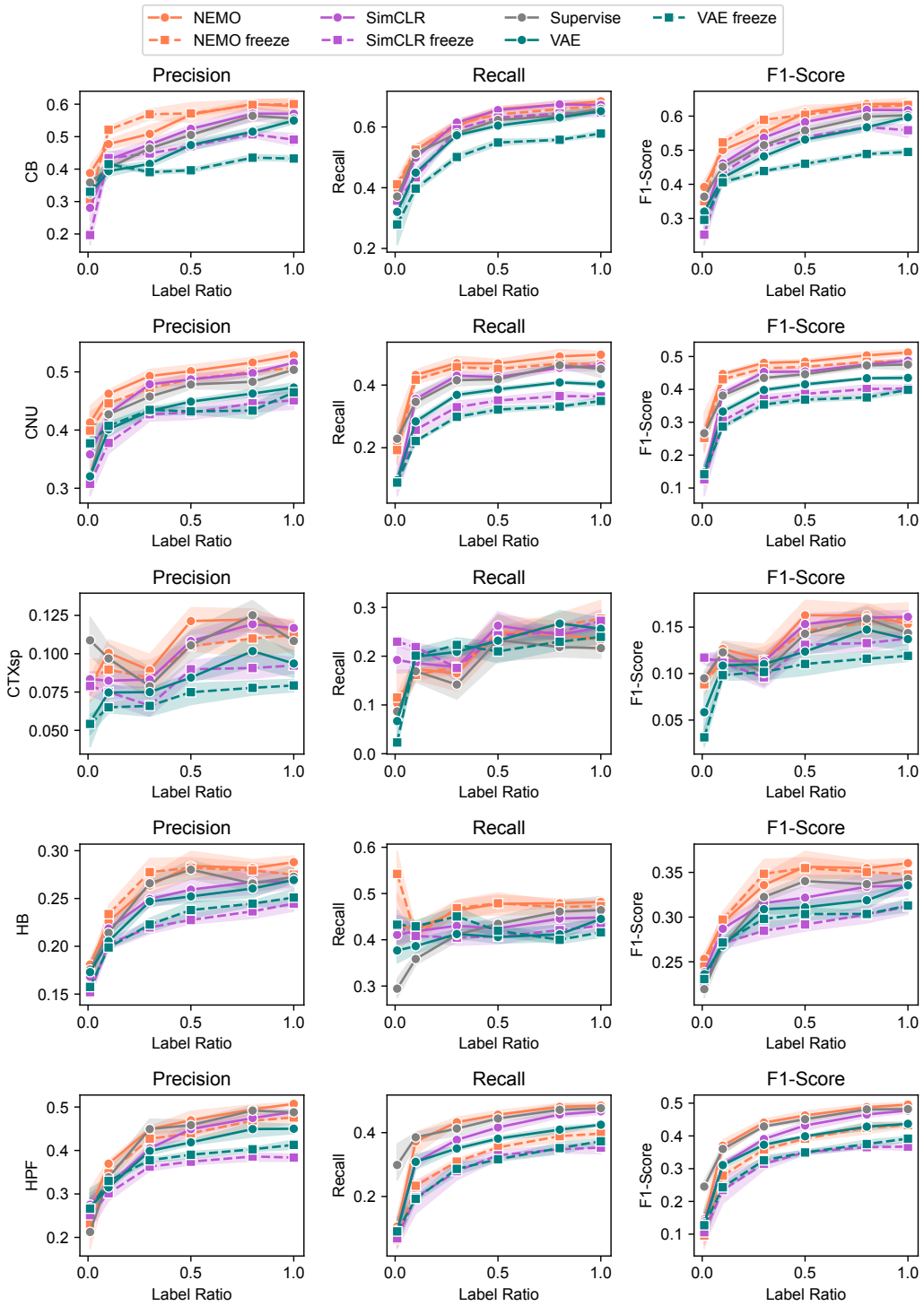


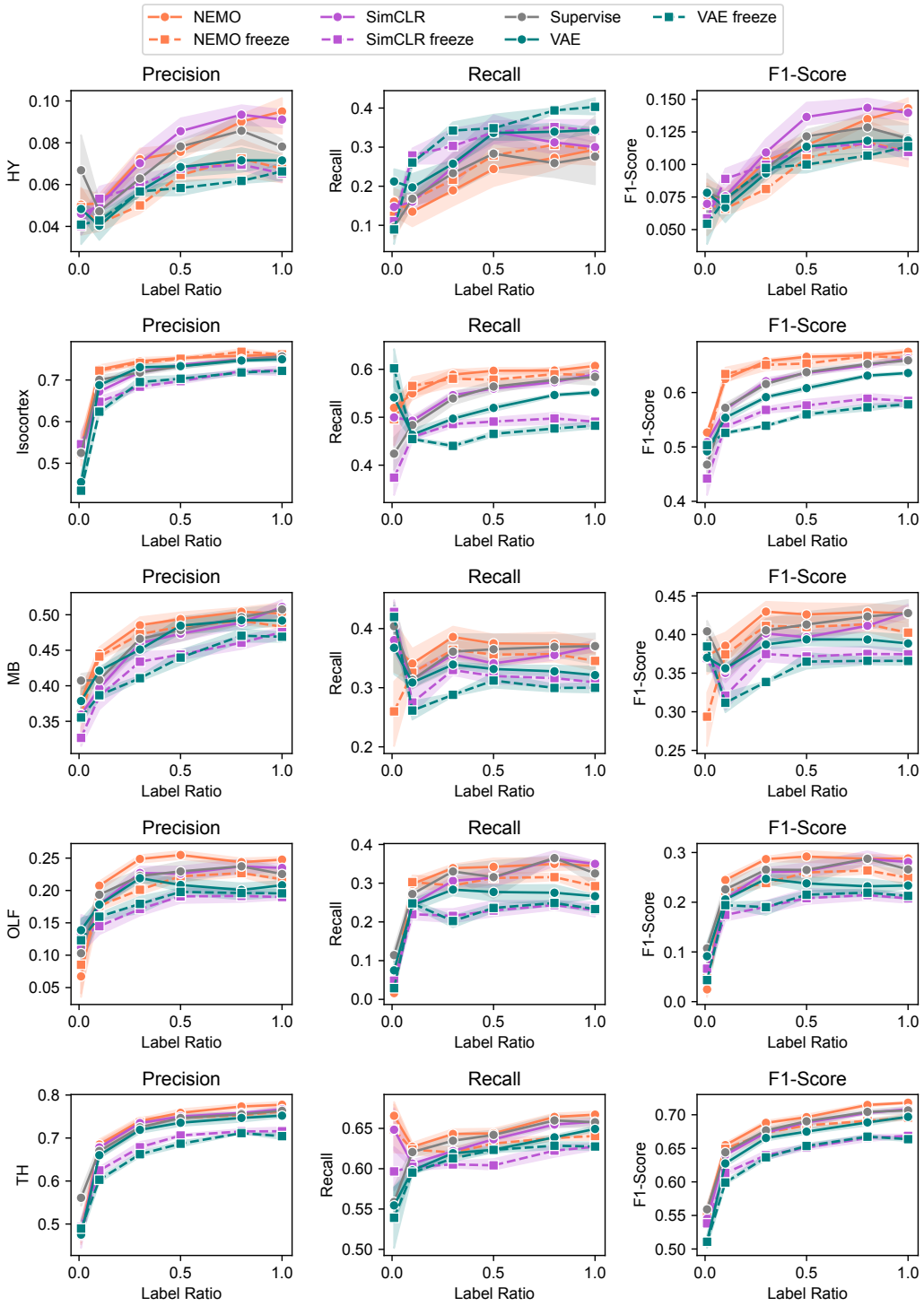
Supplementary Figure 5: Left and middle, macro-averaged F1 scores for linear and MLP-based single-neuron classification of brain region by label ratio, as in Fig. 3(d), replacing accuracy with F1 score. Right, single-neuron MLP-classification balanced F1 scores for uni- and bimodal models, as in Fig. 3(e), replacing accuracy with F1 score.

H.2 IBL REGION CLASSIFICATION WITH LABEL RATIO SWEEP

In Supplementary Figure 5, we show the macro-averaged F1 scores for single neuron classification of brain region by label ratio as complementary to Figure 3.

In Supplementary Figure 6, we study the effect of varying the ratio of labeled data used to train the brain region classifier. Fully supervised methods used the same labeled examples, but were not pretrained. Means and standard-deviation bands are computed over five random initializations for each label ratio.


NEMO
NEMO freeze
SimCLR
SimCLR freeze
Supervise
VAE
VAE freeze
Precision
Recall
F1-Score
CB
GNU
CTXsp
HB
HPF
Label Ratio



Supplementary Figure 6: Detailed comparison of NEMO and SimCLR for region classification performance on IBL data with varying label ratio. We show the precision, recall and F1 score for each class. NEMO shows superior performance in most of the classes, except hypothalamus (HY), which has a small sample size compared to other classes.

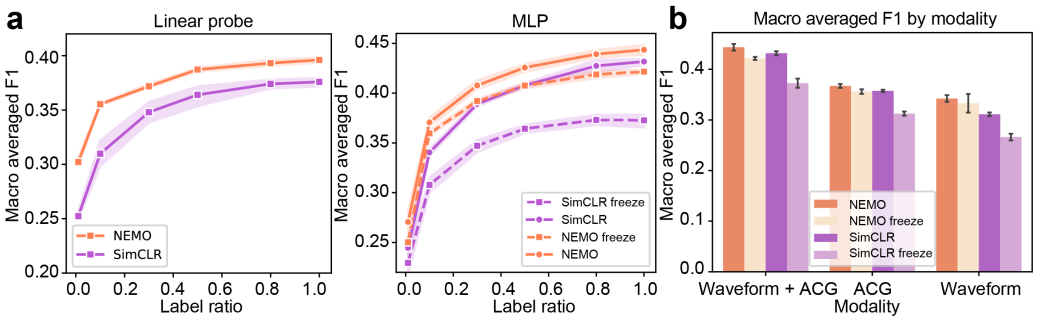
I INDEPENDENT LEARNING ABLATION FOR NEMO

We compared the cell-type classification performance between NEMO with independent and joint learning. The results are shown in Supplementary table 9.

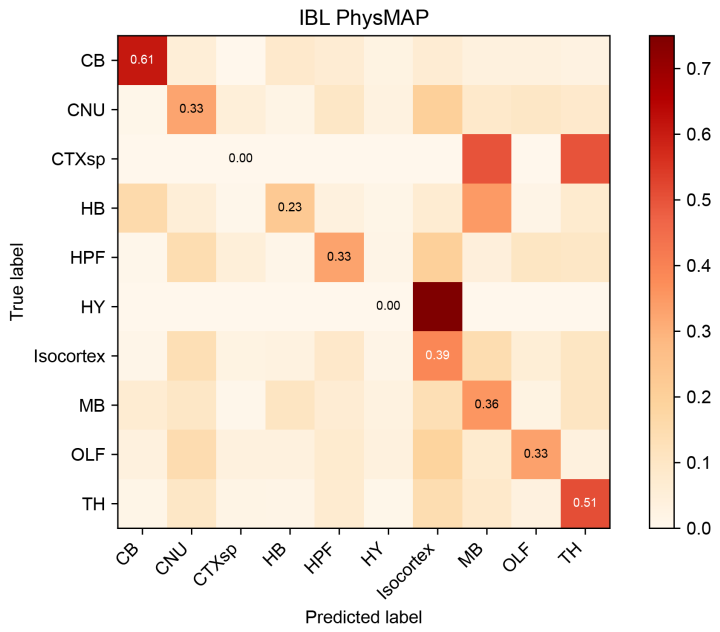
Supplementary Table 9: Cell-type classification for the NP-Ultra dataset for independent vs. joint learning NEMO.

Model	**Linear**		**MLP**		**Finetuned MLP**	
	Acc	**F1**	**Acc**	**F1**	**Acc**	**F1**
independent NEMO	**0.84 ± 0.00**	**0.83 ± 0.00**	0.83 ± 0.00	0.82 ± 0.01	0.87 ± 0.01	0.87 ± 0.01
joint NEMO	0.83 ± 0.01	**0.83 ± 0.01**	**0.84 ± 0.01**	**0.84 ± 0.01**	**0.88 ± 0.01**	**0.88 ± 0.01**

Supplementary Figure 7 shows the macro-averaged F1 scores for single neuron classification of brain region by label ratio with the IBL dataset, complementary to Figure 5.



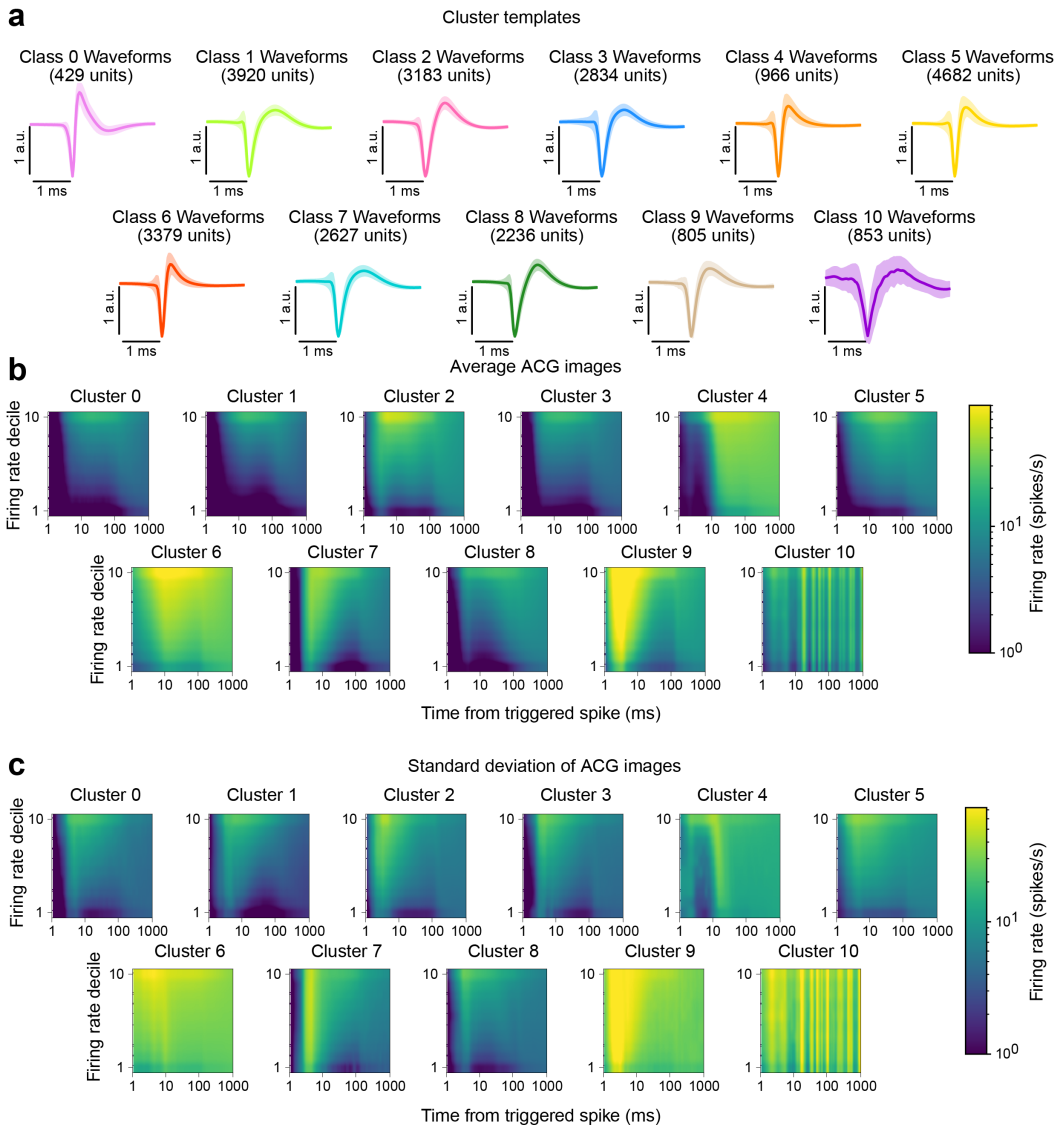
Supplementary Figure 7: Left and middle, macro-averaged F1 scores for linear and MLP-based single-neuron classification of brain region by label ratio for independent learning ablation, as in Fig. 5(a), replacing accuracy with F1 score. Right, single-neuron MLP-classification balanced F1 scores for uni- and bimodal models, as in Fig. 5(b), replacing accuracy with F1 score.



Supplementary Figure 8: Classification results of PhysMAP on the IBL brain region classification task. Due to label imbalance, PhysMAP is unable to predict CTXsp and HY, which leads to a low balanced accuracy and F1 score.

J IBL UNIT CLUSTERING RESULTS WITH NEMO

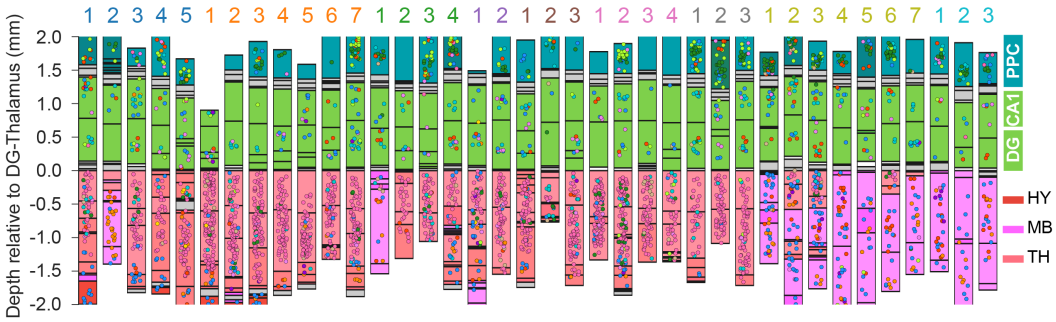
Supplementary Figure 9 shows averages and standard deviations for each cluster's template waveforms and ACG images. Supplementary Figure 10 shows the distribution of cluster labels over brain regions, separated by individual insertions and by the recording lab.



Supplementary Figure 9: Average and standard deviation of the template waveforms and ACGs for the units in each cluster, as shown in Figure 4. (a) The waveforms are consistent within clusters and distinct across clusters. (b) The ACG images are are also distinct across clusters. These results suggest that NEMO is able to find distinct clusterings of neurons across the whole-brain.

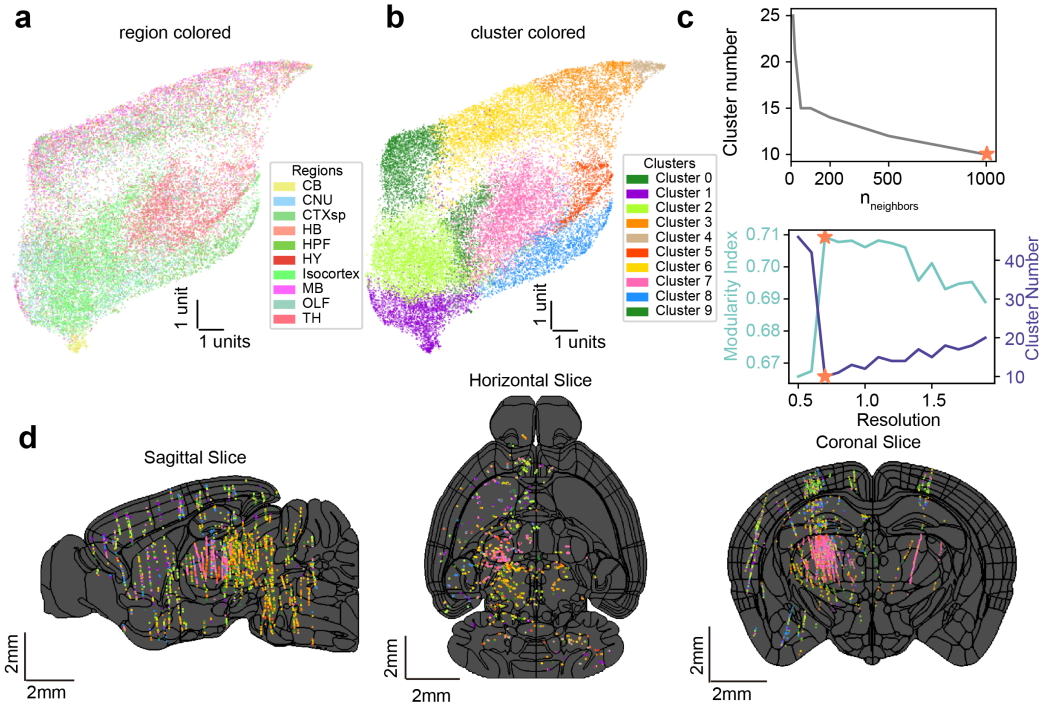
K IBL UNIT CLUSTERING RESULTS WITH RAW FEATURE INPUT

Supplementary Figure 11 shows clustering results for the IBL dataset. The hyper-parameters were selected with similar criteria as in Section 6. Since the number of clusters does not show a similar 'elbow,' but keeps decreasing as $n_{neighbor}$ increases, we picked $n_{neighbor} = 1000$ and used a resolution γ that maximizes the the modularity and minimizes the number of clusters. These clusters are

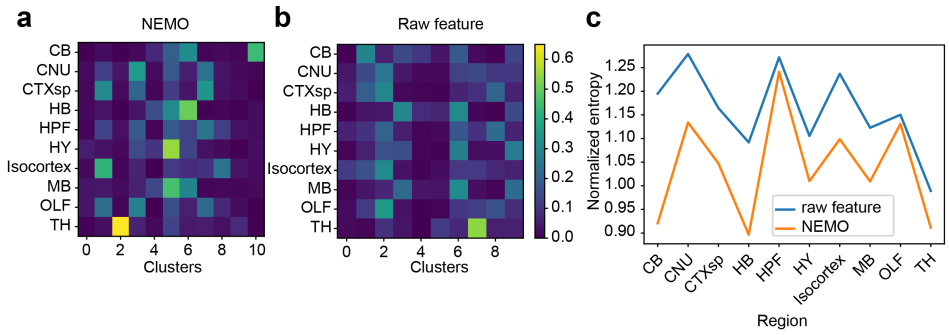


Supplementary Figure 10: Visualization of clustering results across repeated site in the IBL dataset (IBL et al., 2022). The Neuropixels probes target the same brain regions (including posterior parietal cortex, hippocampus, and thalamus) in these insertions. The color of the labels on top of each column indicates the lab ID of each insertion. Our results reveal that the clusters are highly distinguishable by region, with each region containing a distinct group of neurons. Moreover, the dominant cluster IDs for the same region remain consistent across different insertions.

less spatially organized compared to the clusters clustered using NEMO. As shown in Supplementary Figure 12, the clustering result based on NEMO overall shows lower entropy, which indicates higher region-selectivity.



Supplementary Figure 11: IBL neuron clustering using raw features. (a) A UMAP visualization of raw features on the training data colored by anatomical brain region. (b) The same UMAP as shown in (a) but instead colored by cluster labels using a graph-based approach (Louvain clustering). (c) We tuned the neighborhood size in UMAP and the resolution for the clustering. These parameters were selected by maximizing the modularity index which minimized the number of clusters. (d) 2D brain slices across three brain views with the location of individual neurons colored using the cluster IDs shown in (b). The black lines show the region boundaries of the Allen mouse atlas.

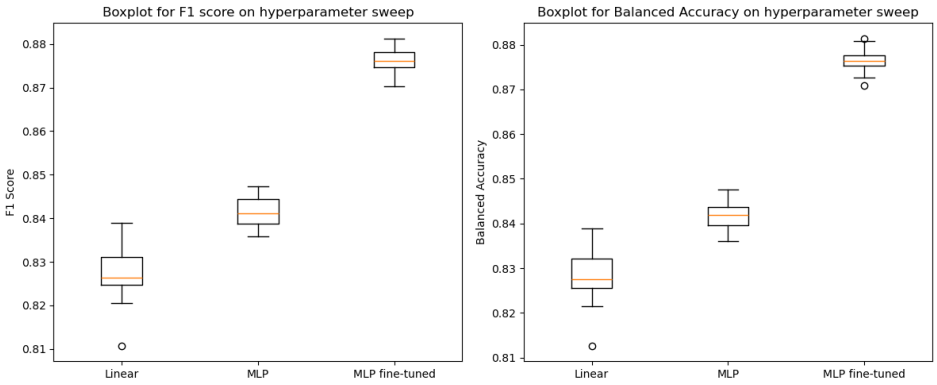


Supplementary Figure 12: Region distribution of clustering results using different NEMO features and raw features (normalized by region). For each region, we get a cluster distribution vector. We then computed the normalized entropy of that distribution. Cluster result based on NEMO overall shows lower entropy, which indicates higher region-selectivity.

L SENSITIVITY TO PARAMETERS

L.1 NEMO HYPERPARAMETER SWEEP

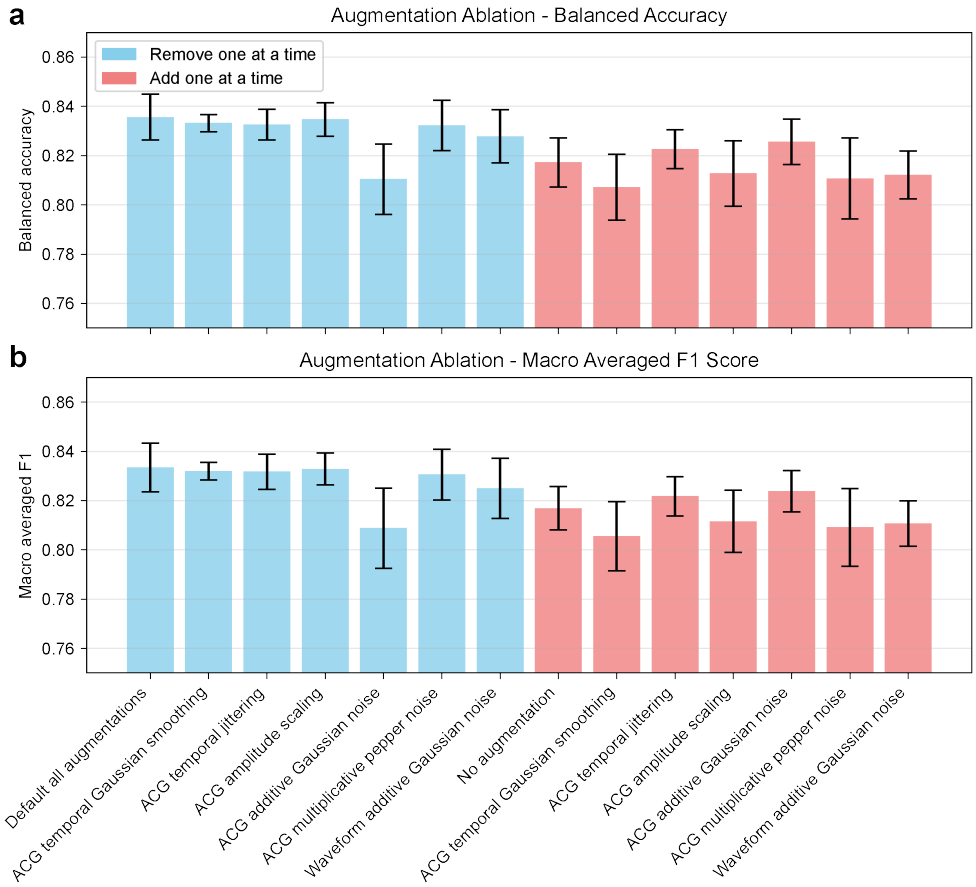
For assess the importance of hyperparameters in NEMO, we do a sweep over the learning rate (1e-4,5e-4,1e-3), the latent dimension size (256,512,1024), and the dropout shift (-0.1,0,0.1) on the single channel NP-Ultra model with linear and MLP decoders. The dropout shift is added to the default waveform and acg dropouts. The boxplots of the resulting balanced accuracy and macro-averaged F1 scores are presented in Figure 13. The results show that hyperparameters do not affect performance that much.



Supplementary Figure 13: Boxplot of Macro-averaged F1 score and Balanced accuracy over 5 seeds of NEMO with linear, MLP and MLP fine-tuned decoders over NEMO hyperparameter sweep.

L.2 AUGMENTATION ABLATION STUDY

To assess the importance of augmentations, we conducted an ablation experiment in two parts: (1) starting with the full set of augmentations, we removed one augmentation at a time, and (2) starting with no augmentations, we added one augmentation at a time. The resulting balanced accuracy and macro-averaged F1 scores are presented in Supplementary Figure 14. The results indicate that additive Gaussian noise in the ACG images is the most impactful augmentation, while the absence of augmentations significantly degrades performance. For other augmentations, the combined effect is large while the individual contribution is smaller.



Supplementary Figure 14: To assess the impact of our augmentations, we performed two analyses on the UHD cell-type classification dataset: (1) removing one augmentation at a time, starting from the full set of augmentations, and (2) applying one augmentation at a time, starting from no augmentations. For each condition, we calculated the mean and standard deviation of (a) balanced accuracy and (b) macro-averaged F1 score with linear classifier.

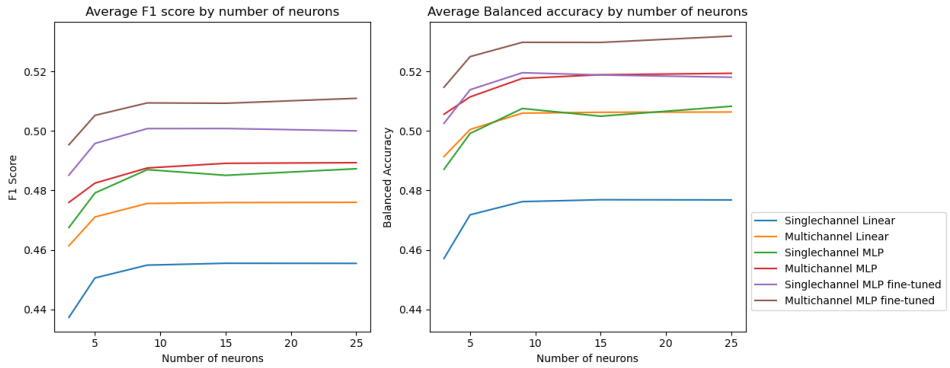
L.3 Multi-neuron neighbor sweep

To evaluate the significance of the number of neurons we choose in our multi-neuron brain region classification model, we conduct a sweep over the maximum number of neighboring neurons (3, 5, 9, 15, 25) that can be selected within a 60-micron radius on the IBL dataset. As illustrated in Supplementary Figure 15, the overall classification performance improves as the maximum number of neighboring neurons increases, reaching a saturation point when this number reaches $\sim$10.

L.4 IBL clustering with different random seeds

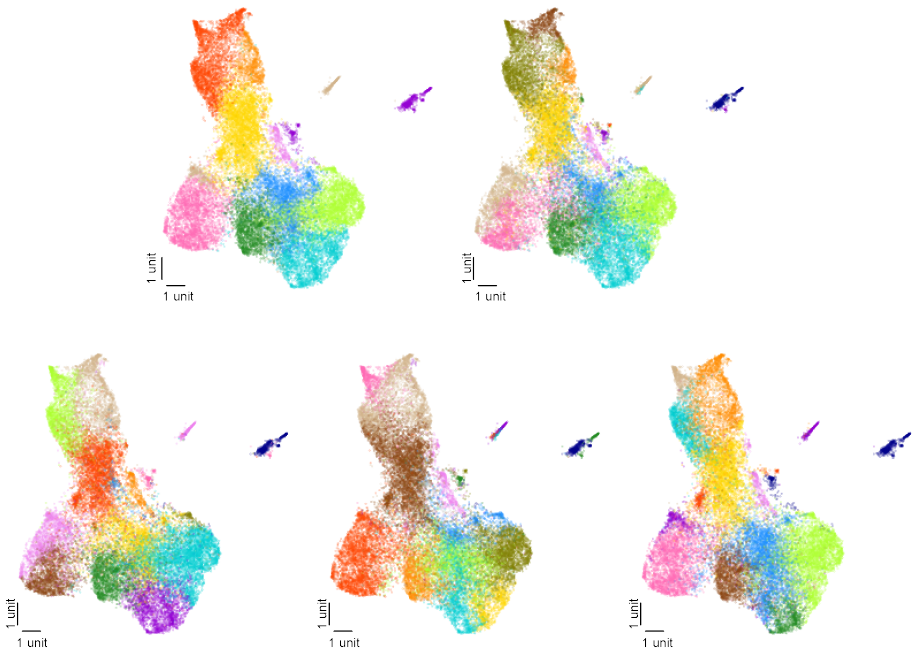
We trained five instances of NEMO with different random seeds and clustered the neurons using the same hyperparameters. The clustering results are visualized in Supplementary Figure 16, where UMAP was applied for dimensionality reduction, with colors distinguishing clusters. To ensure direct comparison, we maintain consistent UMAP visualizations across clustering results. The average adjusted Rand Index (ARI) for these outcomes is 0.48 ± 0.04, while the adjusted mutual information (AMI) score is 0.58 ± 0.02. These metrics indicate a moderate level of agreement among the clustering results.

The boundaries of the clusters are subtle, introducing stochasticity into the clustering results. In addition, clustering the full brain dataset results in a coarse segmentation, while significant vari-



Supplementary Figure 15: Macro-averaged F1 score and Balanced accuracy over 5 seeds of NEMO with multineuron classifier over different number of max neighboring neurons for single and multi-channel IBL with linear, frozen MLP, and MLP fine-tuning.

ability is expected within individual regions. Therefore, these clustering results should mainly be interpreted as exploratory.



Supplementary Figure 16: Clustering results from five NEMO instances trained with different random seeds, visualized using UMAP with consistent dimensionality reduction across runs. Colors indicate distinct clusters.

M STATISTICAL TESTS

To evaluate performance differences between models, we conducted two-sample one-tailed t-tests (significance threshold $p < .05$) on the macro-averaged F1 scores and balanced accuracies across five random seeds. The results are summarized in a significance matrix comparing models: red indicates the model in the column is significantly better than the model in the row, blue indicates it is significantly worse, and white indicates no significant difference.

M.1 CELL-TYPE CLASSIFICATION

For both linear and MLP classifiers, NEMO significantly outperforms all baselines except SimCLR (Supplementary Figure 17). Specifically, NEMO significantly outperforms SimCLR on balanced accuracy when using the fine-tuned MLP and the frozen MLP, but the difference is not significant with the linear classifier. On the F1 score, NEMO significantly outperforms SimCLR with the frozen MLP but not with the fine-tuned MLP or the linear classifier (though the p-value is 0.0505 for the fine-tuned MLP, narrowly missing the $p < .05$ threshold).

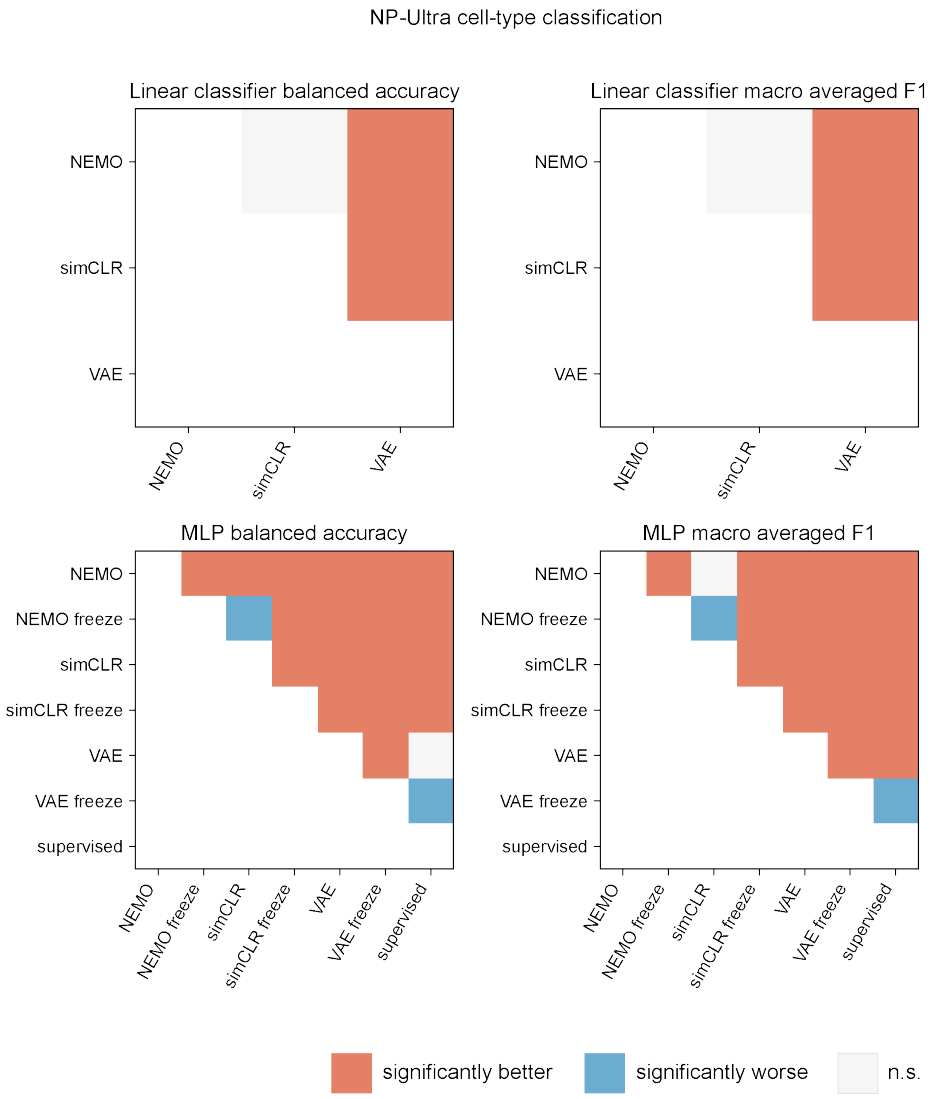
M.2 BRAIN REGION CLASSIFICATION

NEMO consistently outperforms all other baselines for both linear and MLP classifiers (Supplementary Figure 18). For single-modality brain region classification with the MLP classifier (Supplementary Figure 19), NEMO with the fine-tuned MLP either significantly outperforms or shows no significant difference compared to all other baselines.

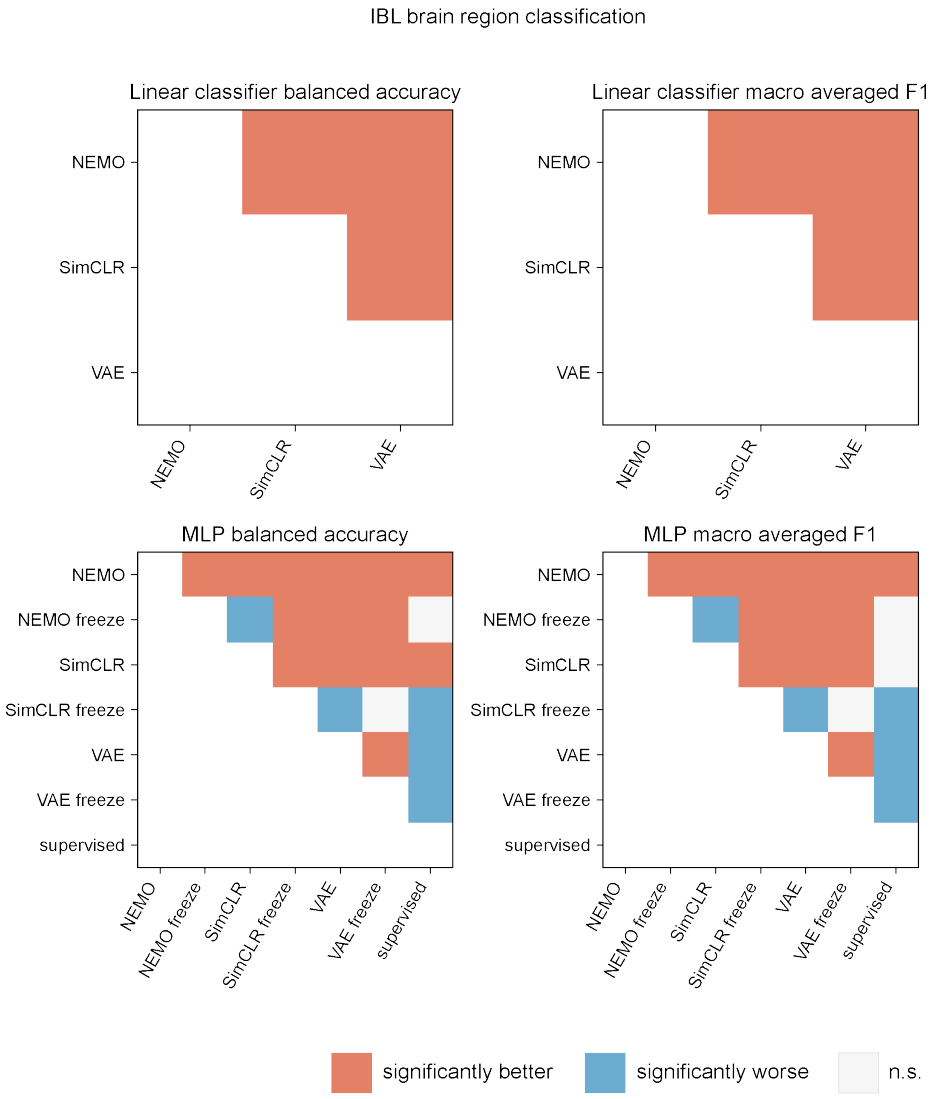
M.3 COMPARISON ACROSS LABEL RATIOS

We further compared classifier performance under varying label ratios. For both the linear classifier (Supplementary Figures 20 and 21) and MLP classifier (Supplementary Figures 22 and 23), NEMO significantly outperforms other baselines at each label ratio. Notably, with only 50% of the labels, the linear classifier with NEMO either significantly outperforms or shows no significant difference compared to all baselines using 100% of the labels. Similarly, the MLP classifier with NEMO achieves comparable or significantly better performance than all baselines using 100% of the labels with only 80% of the labels.

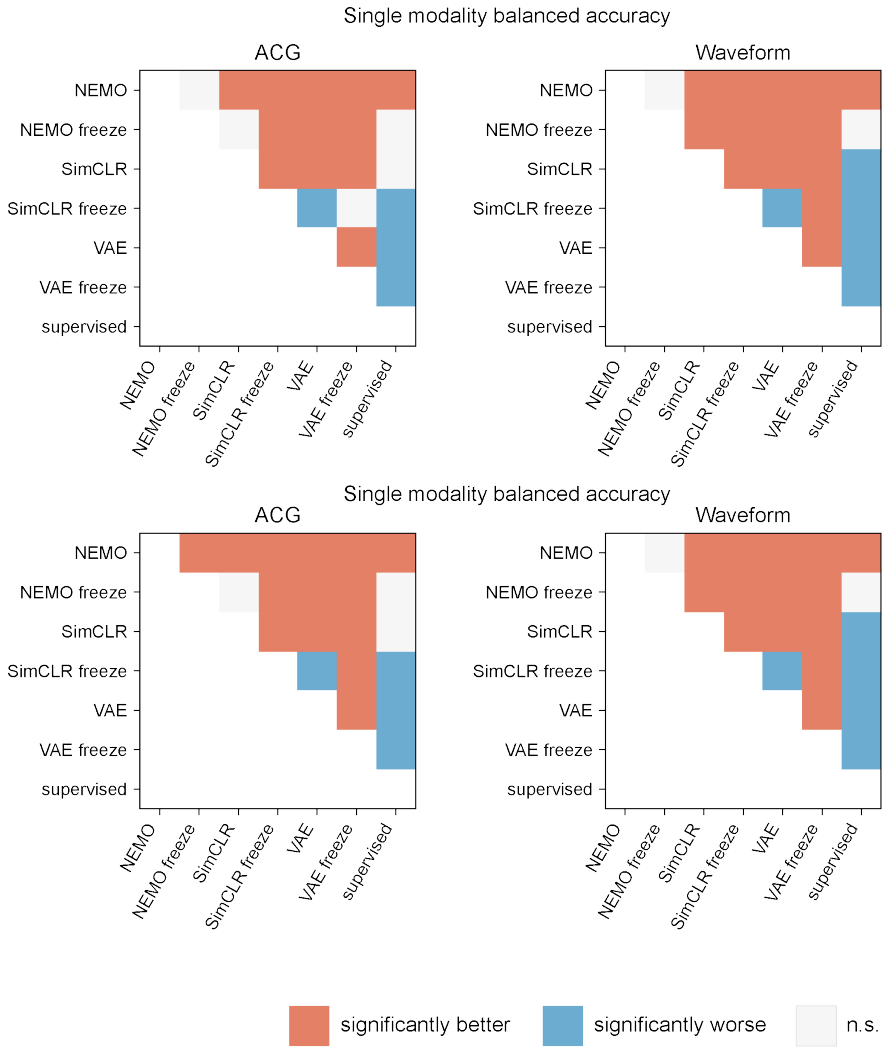
Overall, these results demonstrate that NEMO consistently outperforms or matches the performance of other baselines across multiple evaluation criteria, including cell-type classification and brain region decoding tasks. Its robustness is evident across classifiers, label ratios, and metrics, establishing NEMO as a highly effective model for these challenging neurobiological classification tasks.



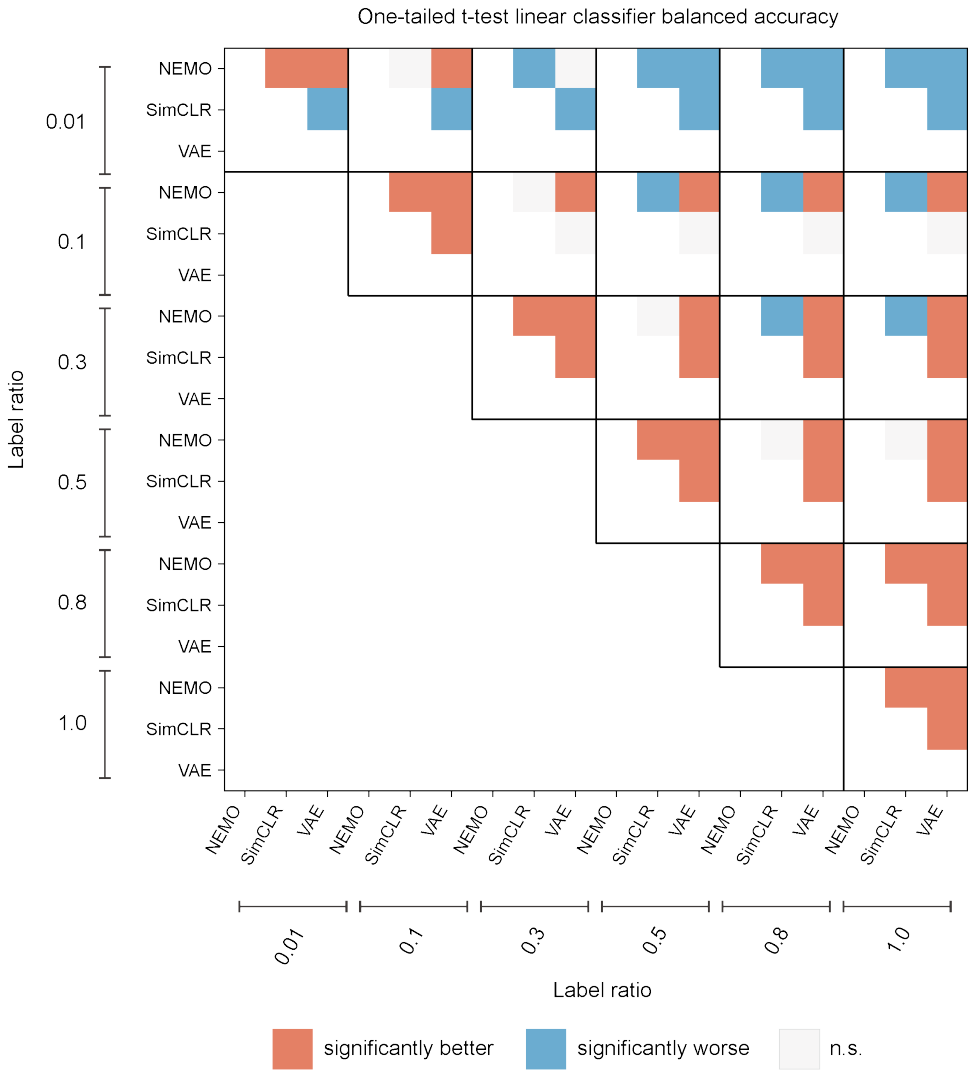
Supplementary Figure 17: One-tailed t-test results for NP-Ultra cell-type classification. For both linear and MLP classifiers, NEMO significantly outperforms or matches all baselines.



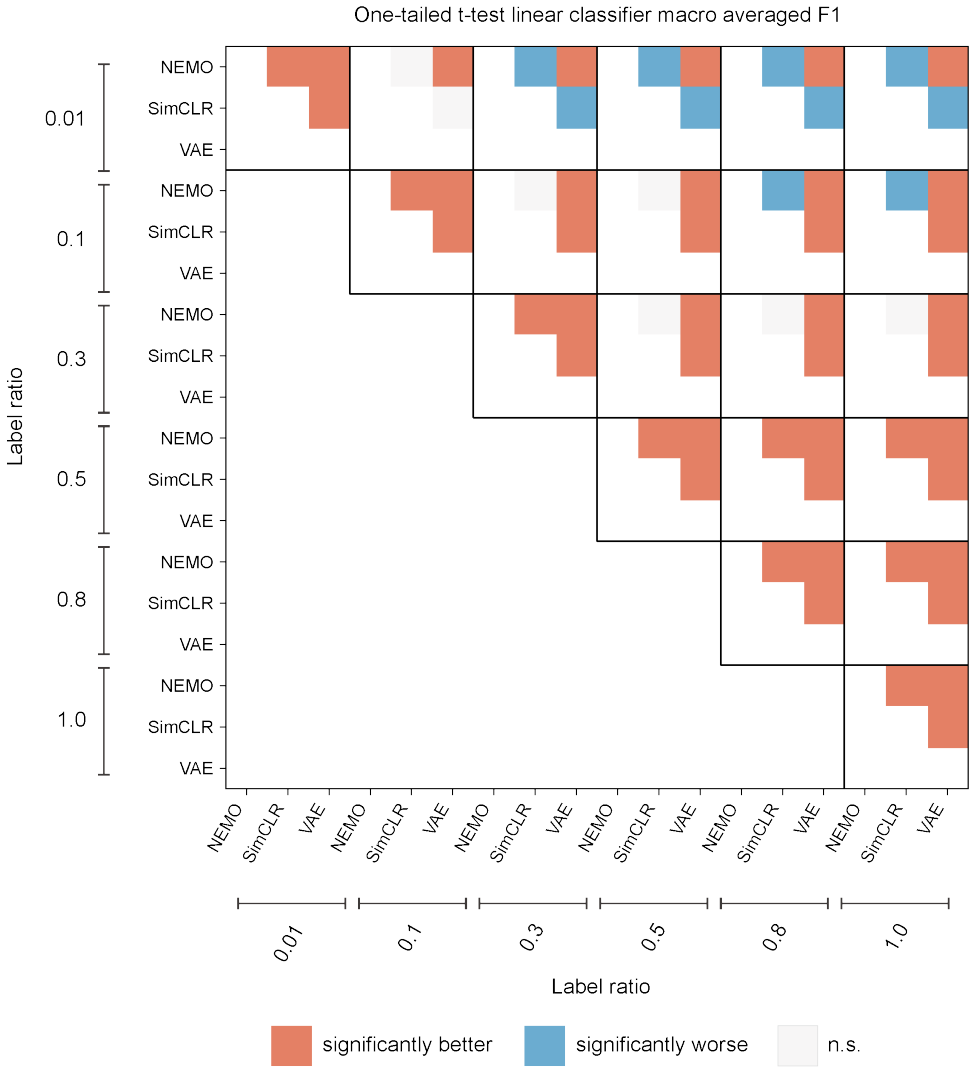
Supplementary Figure 18: One-tailed t-test results for IBL brain region classification. For both linear and MLP classifiers, NEMO significantly outperforms all baselines.



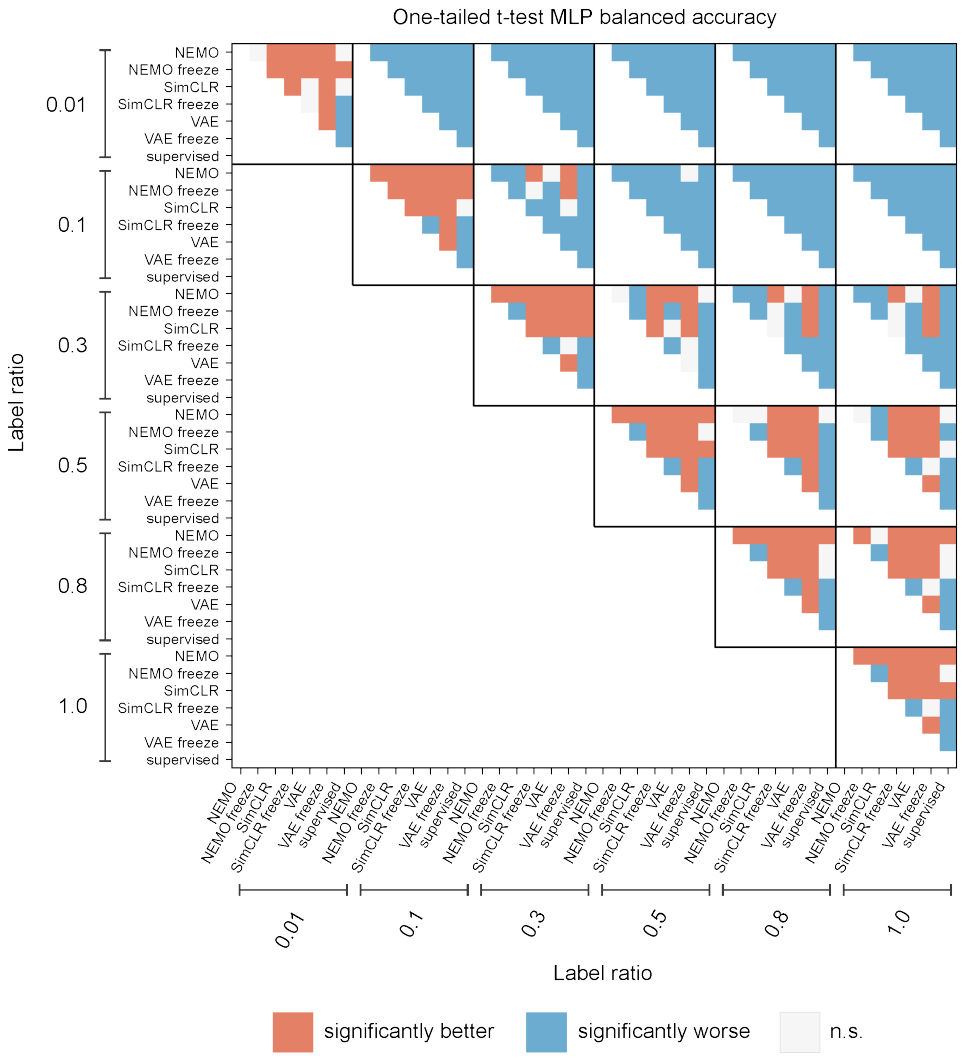
Supplementary Figure 19: One-tailed t-test results for IBL brain region classification with single modality and MLP classifier with end-to-end fine-tuning. For both modality, NEMO significantly outperforms or matches all baselines.



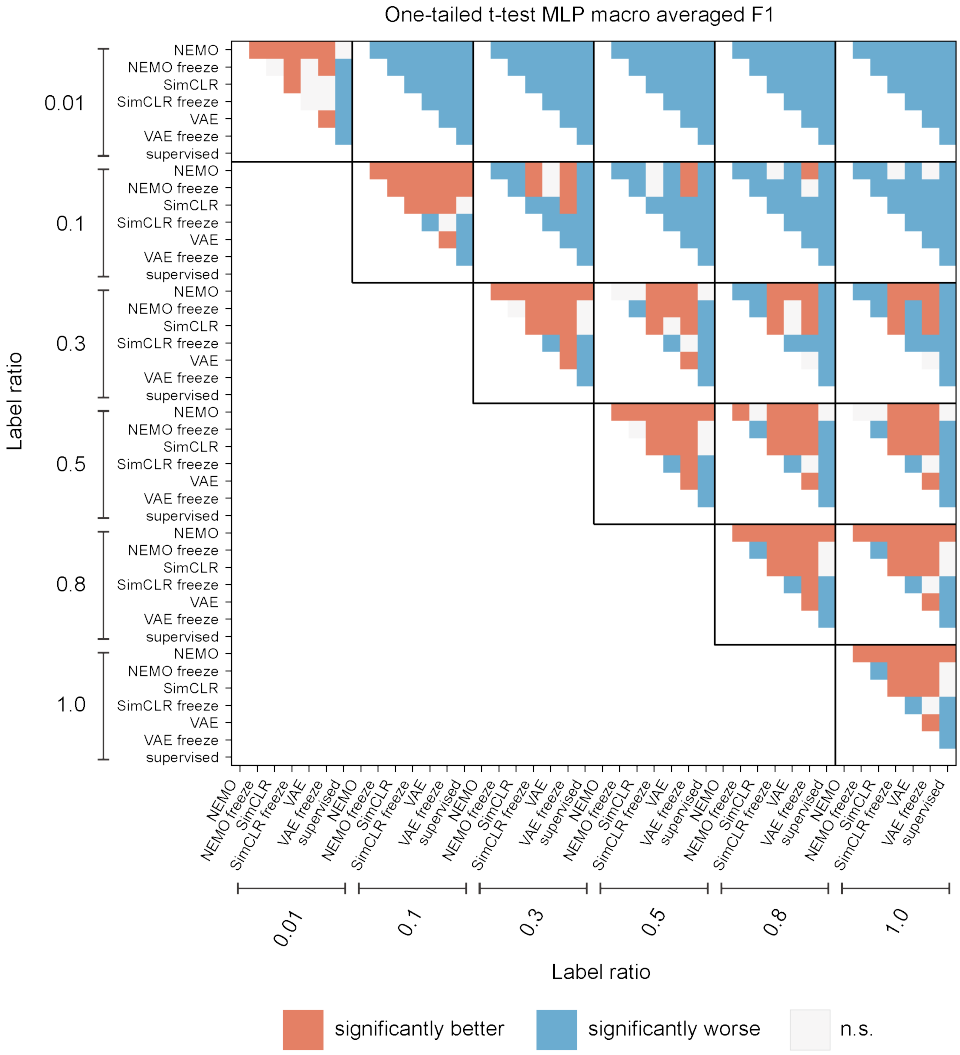
Supplementary Figure 20: One-tailed t-test results on balanced accuracy for IBL brain region classification with linear classifier across different label ratios. For all label ratios, NEMO significantly outperforms all other baselines. With only 50% of the labels, the linear classifier with NEMO either significantly outperforms or shows no significant difference compared to all baselines using 100% of the labels.



Supplementary Figure 21: One-tailed t-test results on macro averaged F1 score for IBL brain region classification with linear classifier across different label ratios. For all label ratios, NEMO significantly outperforms all other baselines. With only 50% of the labels, the linear classifier with NEMO significantly outperforms all baselines using 100% of the labels.



Supplementary Figure 22: One-tailed t-test results on balanced accuracy for IBL brain region classification with MLP classifier across different label ratios. For almost all label ratios (except 0.01), NEMO significantly outperforms all other baselines. With only 80% of the labels, the linear classifier with NEMO either significantly outperforms or shows no significant difference compared to all baselines using 100% of the labels.



Supplementary Figure 23: One-tailed t-test results on macro F1 score for IBL brain region classification with MLP classifier across different label ratios. For almost all label ratios (except 0.01), NEMO significantly outperforms all other baselines. With only 50% of the labels, the linear classifier with NEMO either significantly outperforms or shows no significant difference compared to all baselines using 100% of the labels.