BENCHMARKING SINGLE-MODAL MOLECULAR REPRESENTATIONS ACROSS DIVERSE MODALITIES

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ABSTRACT

Molecular representation learning (MRL) plays a vital role in high-precision drug discovery. Currently, people represent molecules in different modalities (such as sequences, graphs, and images), and have developed many MRL methods. However, three key challenges hinder further progress in the field of MRL: (i) Lack of systematic and unified evaluation on models of different modalities, resulting in unfair comparisons or being affected by randomness; (ii) The specific advantages between different molecular modalities are unclear; (iii) Lacking a unified platform to integrate data of different modalities and a large number of MRL methods. Therefore, we propose the first MRL platform supporting different modalities, called BenchMol, to integrate a large number of sing-modal MRL methods with different modalities and evaluate them systematically and fairly. BenchMol has four attractive features: (i) Rich modalities: BenchMol supports 7 major modalities of molecules, such as fingerprint, sequence, graph, geometry, image, geometry image, and video; (ii) Comprehensive methods: BenchMol integrates 23 mainstream MRL methods to process these modalities; (iii) New benchmarks: BenchMol constructs two new benchmarks based on PCQM4Mv2 and ChEMBL 34, called MBANet and StructNet, for a more systematic evaluation. (iv) Comprehensive evaluation: evaluation covers different aspects of molecules, such as basic attributes and molecular types. Through BenchMol, we conduct large-scale research on methods of different modalities and report many insightful findings. We hope that BenchMol can help researchers quickly use MRL methods with different modalities on the one hand; and on the other hand, provide meaningful insights into multi-modal MRL and help researchers choose appropriate representations in downstream tasks. We open-sourced BenchMol in Github.

1 INTRODUCTION

Molecular representation learning (MRL) is a prerequisite for high-precision drug discovery (Li et al., 2022; Catacutan et al., 2024). With the development of deep learning, researchers have developed a large number of MRL methods in recent years (Yi et al., 2022). According to the different representation forms of molecules, existing methods represent molecules in 7 different modalities (as shown in Figure 1) and use modality-specific techniques to extract molecular representations, namely molecular sequence (Kim et al., 2021; Ross et al., 2022), graph (Hu et al., 2020a; Liu et al., 2022a), geometry (Fuchs et al., 2020; Satorras et al., 2021), image (Xiang et al., 2023), geometry image (Xiang et al., 2024a) and video (Xiang et al., 2024b).

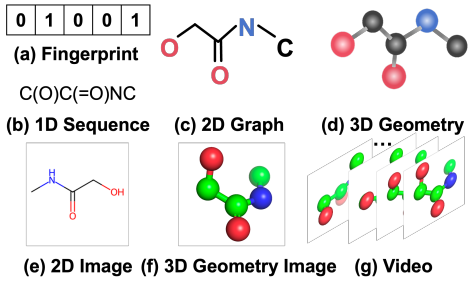

Figure 1: Schematic diagram of "C(O)C(=0)NC" with different modalities. The molecule is represented in 7 different modalities: (a) Fingerprint, (b) Sequence, (c) Graph, (d) Geometry graph, (e) 2D image, (f) 3D geometry image, and (g) video.

Challenges. Despite the remarkable success of MRL, there are still three key challenges in its development that hinder the further development of multi-modal MRL methods:

I. **Unfair comparison.** Differences in evaluation strategies lead to incomparable or unfair comparisons between methods. We summarize four differences in the evaluation process, including the method for dividing the dataset (Rong et al., 2020; Ross et al., 2022), the range of parameter optimization (Hu et al., 2020a; Wang et al., 2022), the standardization of labels (Ross et al., 2022; Zhou et al., 2023), and the selection of random seed (Zeng et al., 2022; Wang et al., 2022; Xia et al., 2023). For example, MoLFormer (Ross et al., 2022) uses random scaffold split to divide the dataset and is compared with MolCLR (Wang et al., 2022) and GraphMVP-C (Liu et al., 2022a) which use strict scaffold split. In general, it is easier to achieve good performance on datasets with random scaffold split than on datasets with scaffold split. Compared to previous studies (Hu et al., 2020a) that use the same hyperparameters for different tasks and run 10 replicates, MolCLR performs independent hyperparameter optimization for different tasks and runs 3 replicates. BARTSmiles (Chilingaryan et al., 2022), Uni-Mol (Zhou et al., 2023) and MoLFormer use label regularization to compare with other methods. Hu et al. and Xia et al. use consistent random seeds from 0 to 9 to initialize the model while a large number of studies (Rong et al., 2020; Zeng et al., 2022; Wang et al., 2022) do not explicitly state the random seeds, which may result in a benchmark deviation. Therefore, *it is necessary to build a platform to fairly evaluate methods with different modalities*, which will pave the way for researchers to explore scientific questions instead of falling into biases caused by experimental differences.

II. **Incomplete evaluation for different modality data.** MRL is evolving towards a multi-modal direction, relying on various technology stacks, such as sequence modalities based on Natural Language Processing (NLP) (Devlin et al., 2019; Lewis et al., 2020), graph modality based on graph deep learning (Xu et al., 2018), geometry modality based on geometry deep learning (Monti et al., 2017; Atz et al., 2021), and image, geometry image and video modalities based on Computer Vision (CV) (He et al., 2020; Kirillov et al., 2023). Currently, a large number of methods based on different modalities have been developed in the field of MRL (Hu et al., 2020a; Ross et al., 2022; Zeng et al., 2022). Intuitively, the data and encoding methods between different modalities are different, which may lead to their preference for different types of molecules. However, *this preference is still unclear and deserves further exploration*. In addition, researchers often focus on the task of molecular property prediction from MoleculeNet Wu et al. (2018) in the evaluation of MRL (Xiang et al., 2023; Xia et al., 2023; Xiang et al., 2024a). However, *a single evaluation is not comprehensive for studying preferences in molecular representation of different modalities*. Here, We introduce the Molecular Basic Attribute (MBANet) benchmark built from IEM (Xiang et al., 2024a) and the StructNet benchmark built from the ChemBL 34 database (Zdrazil et al., 2024), as shown in Figure 2, to further evaluate the ability of these models to identify essential molecular attributes and mine information from different types of molecules.

III. **Lack of a unified platform supporting diverse modalities.** MRL methods of different modalities are scattered in various corners of the Internet with different development environments and different running pipelines. *It is challenging to integrate various methods with different modalities into a unified platform to support multiple modality data*. Currently, there remains a blank in the molecular-based platform supporting diverse modalities. We hope to propose a unified molecular platform to provide meaningful insights for multi-modal MRL and facilitate the use and development of multi-modal molecules by researchers.

Contributions. In this work, we aim to provide the first MRL platform unified diverse modalities (called BenchMol) covering a large number of existing algorithms and re-evaluate existing methods in a fair and comprehensive manner. Our contributions are summary as follows:

- **Unified and flexible platform for supporting different molecular modalities.** BenchMol is a flexible and easy-to-use toolkit, which integrates 7 molecular modalities into a unified framework. Meanwhile, BenchMol provides a complete pipeline from raw data to the evaluation of the final model, which includes data preprocessing (57 modality extractors), predefined models (6 sequence models, 13 graph models, 9 geometry models and at least 900 visual models), training strategies (linear probing and fine-tuning) and a large number of evaluation metrics.
- **Novel benchmarks.** We propose two benchmarks, MBANet and StructNet, to explore the advantages of existing models in basic molecular information (12 atoms, 4 bonds and 8 attributes) and

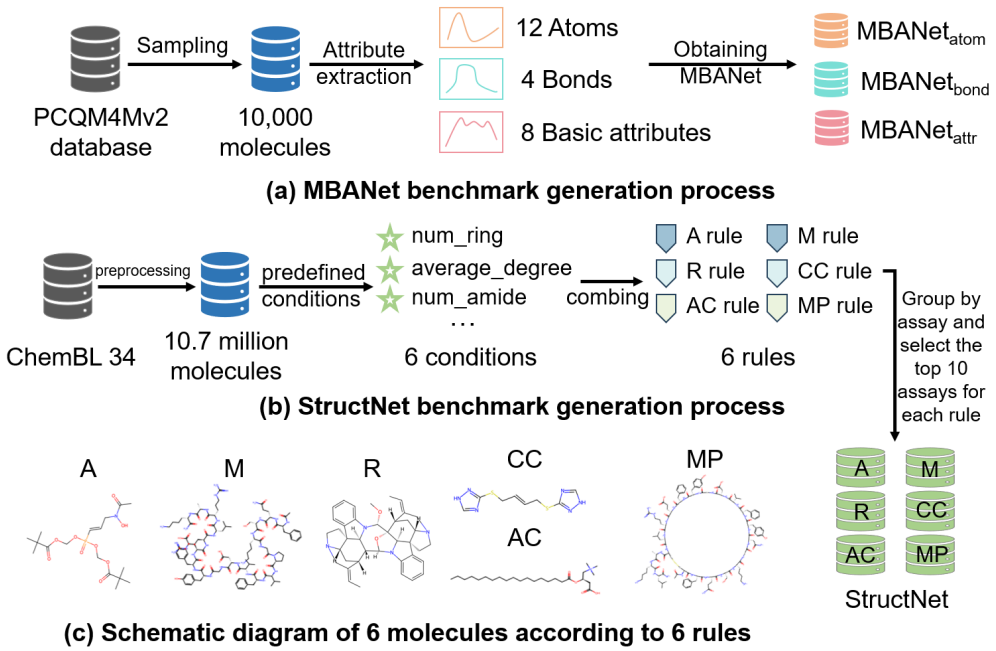

Figure 2: (a) Schematic diagram of MBANet. (b) Schematic diagram of StructNet. 6 conditions are num_ring, average_degree, num_amide, has_branch, num_amide and molecular weight. 6 rules are acyclic (A) rule, complete chain (CC) rule, acyclic chain (AC) rule, macrocyclic peptide (MP) rule, macromolecule (M) rule and reticular (R) rule. (c) Schematic diagram of the molecular structure corresponding to the 6 rules in StructNet.

the preferences for 6 different types of molecules (acyclic chain, acyclic, complete chain, macro, macrocyclic peptide, reticular molecules), respectively.

- **Experiment comprehensively and fairly.** With BenchMol, we train at least 57,060 models, including 6,960 models on linear probing of 12 MoleculeNet, 900 models on fine-tuning of 3 datasets from MBANet, and 49,200 models on fine-tuning of 60 datasets from StructNet.
- **Meaningful insights.** Based on extensive experiments and rigorous comparisons, we provide many meaningful insights, including 9 main finds. Here, we highlight several important conclusions: (1) Multi-conformation ensemble is expected to significantly improve the performance of models based on visual modality, which provides direction for further development of molecular vision; (2) Video modality excels at atomic-level and attribute-level tasks and graph/geometry modality excels at bond-level tasks, which provides guidance for the choice of modality; (3) With regard to molecular preferences, the geometry modality prefers acyclic molecules, the fingerprint and graph modalities prefer cyclic molecules, and the vision-based modalities prefer macrocyclic and reticular molecules, which provides guidance for the multi-modal fusion of molecules.

2 Related Works

Molecular representation learning (MRL). Existing MRL can be mainly divided into the following categories, including sequences, 2D topological graphs, 3D geometric graphs, 2D images, 3D geometry images, and videos. Sequence-based methods represent molecules as 1-dimensional strings (such as SMILES) and use NLP-related techniques to learn molecular representations (Ross et al., 2022; Zheng & Tomiura, 2024). Graph-based methods treat the atoms and bonds of molecules as nodes and edges in a graph and use GNN and its variants to extract features (Hu et al., 2020a; Wang et al., 2022). Image-based methods treat molecules as a flat image and use computer vision (CV)-related techniques for processing (Zeng et al., 2022; Xiang et al., 2023; Zhang et al., 2023). Subsequently, considering the importance of geometric information in molecules, geometric deep learning methods represent molecules as geometric graphs and extract information from them (Schütt et al., 2021; Liu et al., 2022b). Meanwhile, image-based methods render the geometric information of molecules into geometry images and perform feature extraction (Xiang et al., 2024a).

Recently, video-based methods have been proposed, which represent the conformation of molecules as a video and extract features (Xiang et al., 2024b). Given that MRL is rapidly evolving towards different modalities, it is necessary to aggregate these methods with different modalities into a unified platform and accelerate the development and use of researchers.

Molecular benchmark platforms. OGB (Hu et al., 2020b) proposes a set of diverse, challenging and realistic benchmark datasets covering molecular graphs, which mainly focus on graph data. Molecule3D (Xu et al., 2021) develops a benchmark that includes a dataset with precise ground-state geometries of approximately 4 million molecules, and provides a few baseline methods based on DeeperGCN (Li et al., 2023) and DAGNN (Yang et al., 2021). Deng et al. (2023) focuses on the evaluation of molecular fingerprints and 2D graphs. MOLGRAPHEVAL (Wang et al., 2024) focuses on evaluating the impact of different pre-training strategies based on 2D graphs. Geom3D (Liu et al., 2024) focuses on geometric data of different biological entities (such as molecules, proteins, crystalline materials). However, with the increase in modality data, the integration of multiple fields has raised concerns about fairness and comprehensiveness in evaluation protocols. Different from the above mentioned methods, BenchMol is the first molecular benchmark platform unified multiple molecular modalities, which integrates 7 different modalities (fingerprint, sequence, graph, geometry, image, geometry image, and video) and provides a easy-to-use interface for access.

3 PRELIMINARIES

There is a molecule m with n_a atoms and n_b bonds and the representation of different modalities is as follows:

Fingerprint. Molecular Fingerprints $\mathcal{F}$ are a compact, fixed-size representation, where $\mathcal{F} \in \mathbb{R}^{n_{fp}}$ and n_{fp} represents the dimension of molecular fingerprint. Currently, there are many fingerprints developed (Mason et al., 2001), such as 167-dimensional MACCS (Molecular ACCess System) (Durant et al., 2002), ECFPx with custom dimension (Rogers & Hahn, 2010), 210-dimensional RDKit2D (Landrum et al., 2016). You can read (Hou et al., 2024) for more details about fingerprints.

Sequence. A molecule is regarded as a string sequence, such as SMILES (Weininger et al., 1988), SELFIES (Krenn et al., 2020) and IUPAC (Kuhn et al., 2004). These sequences are split into tokens by a tokenizer with word segmentation rules, which is formalized as $\mathcal{S} = \{s_0, s_1, ..., s_{n_a}\}$, where s_{n_a} represents a token. Currently, the most commonly used molecular sequence is SMILES and a large number of technologies (Kim et al., 2021; Ross et al., 2022) are developed based on it. In this paper, we focus on the study of molecular SMILES because of its popularity.

Graph. A molecular graph $\mathcal{G} = (\mathcal{V}, \mathcal{E})$ consists of a set of nodes $\mathcal{V} \in \mathbb{R}^{n_a \times n_f}$ and edges $\mathcal{E} \in \mathbb{R}^{n_b}$, where n_f represents the feature number of atom (e.g., atom type) (Hu et al., 2020a; Xia et al., 2023). Assume that there are two atoms v and u in $\mathcal{V}$, $\mathcal{E}$ represents an adjacency matrix $\boldsymbol{A} \in \mathbb{R}^{n_a \times n_a}$ that indicates whether v and u are connected, where $\boldsymbol{A}[v, u] = 0$ means there is no edge (bond) otherwise it means there is an edge (bond). In practical applications, since bonds have multiple chemical properties (e.g., bond types), the adjacency matrix $\boldsymbol{A}$ can be easily extended to $\boldsymbol{A}^* \in \mathbb{R}^{n_a \times n_a \times d_b}$, where d_b represents the number of chemical properties of the bond (Liu et al., 2022a; Wang et al., 2022). For example, the type of the bond formed by nodes v and u can be formalized as $\boldsymbol{A}^*[v, u, i_t] = \{0, 1, 2, 3, 4\}$, where i_t represents the index describing the bond type, 0 represents no bond, 1 represents a single bond, 2 represents an aromatic bond, and so on.

Geometry. Geometry graph introduces the 3-dimensional coordinates of atoms based on graph (Zhou et al., 2023; Satorras et al., 2021), which is formalized as $\hat{\mathcal{G}} = (\hat{\mathcal{V}}, \hat{\mathcal{E}})$, where $\hat{\mathcal{V}} \in \mathbb{R}^{n_a, n_f+3}$. Please note that in practical applications, models do not include edge information or use fully connected adjacency matrices to represent edge information when processing geometric graphs.

Image, Geometry Image and Video. Molecular images (Xiang et al., 2023), geometry images Xiang et al. (2023) and videos (Xiang et al., 2024b) are based on visual representations of molecules, which are atom- and bond-independent and are made up of a bunch of pixels. We describe the importance of the visual modality in Appendix C.3. Using RDKit (Landrum, 2013), the SMILES of a molecule can be converted into a molecular image $\mathcal{U}^I \in \mathbb{R}^{224 \times 224 \times 3}$ (Figure 1(e)). The geometry image (Figure 1(f)) and video (Figure 1(g)) of the molecule take into account the 3D structural information of the molecule and are generated using PyMOL (DeLano et al., 2002). See Appendix B for details of visual rendering. Formally, the geometry image and video can be represented as

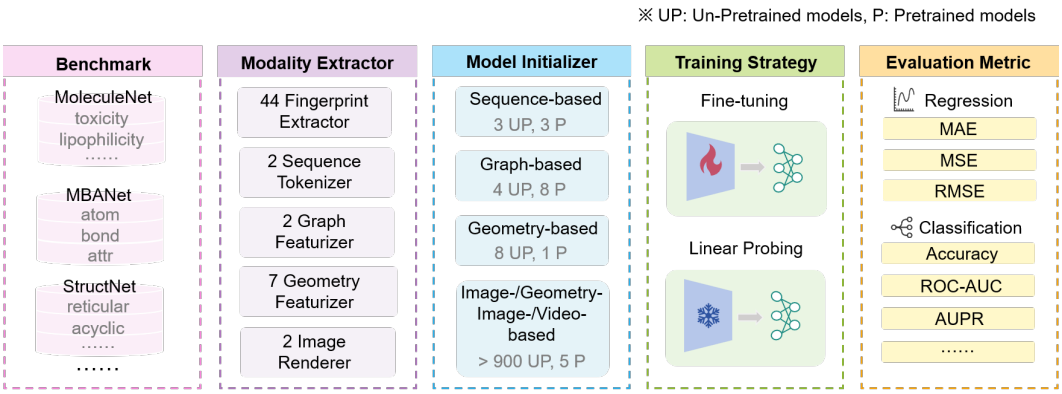

Figure 3: Overview of the proposed BenchMol.

$\mathcal{U}^G \in \mathbb{R}^{4\times224\times24\times\times3}$ and $\mathcal{U}^V \in \mathbb{R}^{60\times224\times24\times\times3}$, respectively, where 4 and 60 represent the number of views in the geometry image and the number of frames in the video.

4 BENCHMOL: BENCHMARK PLATFORM SUPPORTING MULTIPLE MOLECULAR MODALITIES

4.1 OVERVIEW OF BENCHMOL

As shown in Figure 3, BenchMol consists of 5 modules. The *Dataset Collector Module* provides 3 types of benchmarks (Section 4.2). The *Modality Extractor Module* is used for data preprocessing, which converts raw data into input for models of specific modalities (4.3). The *Model Initializer Module* is used to initialize a large number of models (Section 4.4). The *Training Strategy Module* is used to provide model trainers, including linear probing and fine-tuning (Section 4.5). The *Evaluation Metric Module* provides the indicators required for classification and regression tasks (Section 4.6). For a discussion of the motivation and potential impact of BenchMol, see Appendix C.1.

4.2 DATASET COLLECTOR MODULE

In BenchMol, we provide three benchmarks, MoleculeNet (Wu et al., 2018), MBANet and StructNet. We contribute MBANet and StructNet to systematically analyze the preferences and performance of different modal methods. See Appendix C.2 for details of the motivation and practicality of the benchmarks. Next, we introduce the construction process of MBANet and StructNet.

MBANet. MBANet aims to study the ability of different methods to capture molecular basic information, including atom distributions, bond distributions, and basic attributes. As shown in Figure 2(a), we first sample 10,000 molecules from PCQM4Mv2 (Hu et al., 2017). Then, we count the number of atoms and bonds in each molecule, which are formalized as $\mathcal{K}^a = \{k_1^a, k_2^a, ..., k_{12}^a\}$ and $\mathcal{K}^b = \{k_1^b, k_2^b, k_3^b, k_4^b\}$, respectively, where $\mathcal{K}^a \in \mathbb{Z}^{12}$ and $\mathcal{K}^b \in \mathbb{Z}^4$ represent the count of 12 types of atoms (C, N, O, F, S, Cl, Br, P, Si, B, Se, Ge) and 4 types of bonds (SINGLE, AROMATIC, DOUBLE, TRIPLE). Subsequently, we further extracted 8 basic attributes, including {molecular weight, MolLogP, MolMR, BalabanJ, NumHAcceptors, NumHDonors, NumValenceElectrons, TPSA}, which are formalized as $\mathcal{K}^k \in \mathbb{R}^8$. Finally, we can generate three different types of datasets: MBANet$_{atom} = \{m, \mathcal{K}^a\}$, MBANet$_{bond} = \{m, \mathcal{K}^b\}$, and MBANet$_{attr} = \{m, \mathcal{K}^k\}$, where m represents molecules. See Appendix D.1 for details and limitation analysis of the MBANet.

StructNet. StructNet is designed to evaluate the preference of models with varying modalities for molecule types. As shown in Figure 2(b), we first collect over 14.4 million the latest molecules from ChemBL 34. Then, we predefine 6 different conditions, including the number of rings, average degree, presence of branches, maximum number of rings, number of amides, and molecular weight. Based on the molecular SMILES sequences, we leverage RDKit to generate these conditions. Then, we formulate 6 rules to classify molecules into different types: acyclic, complete chain, acyclic chain, macrocyclic peptide, reticular, and macromolecule. As shown in Figure 2(c), the application of these rules enables us to identify molecules exhibiting diverse characteristics. For example, a molecule generated by the acyclic chain rule is a chain-like molecule without any rings. Ultimately,

we group the molecules based on the Assay ChEMBL ID defined in ChemBL 34 and select the top 10 assays with the largest number of samples to construct StructNet. It is worth noting that these result in the creation of 60 datasets within StructNet, with 10 for each molecule type. See Appendix D.2 for details of the StructNet.

Table 1: The supported models in BenchMol.

Modality Type	Pre-training Models	Models w/o Pre-training
1D Sequence	CHEM-BERT (Kim et al., 2021), CHEM-RoBERTa (Kim et al., 2021), MoLFormer (Ross et al., 2022)	BERT (Devlin, 2018), RoBERTa (Liu, 2019), Transformer_Rotate (Ross et al., 2022)
2D Graph	EdgePred (Hu et al., 2020a), ContextPred (Hu et al., 2020a), infomax (Hu et al., 2020a), masking (Hu et al., 2020a), GraphMVP (Liu et al., 2022a), MolCLR (Wang et al., 2022), CGIP-Graph (Xiang et al., 2023), MoleBERT (Xia et al., 2023)	GIN (Xu et al., 2018), GAT (Veličković et al., 2018), GCN (Li et al., 2021), GraphSAGE (Hamilton et al., 2017), DeeperGCN (Kipf & Welling, 2016)
3D Geometry Graph	Uni-Mol (Zhou et al., 2023)	SchNet (Schütt et al., 2017), DimeNet (Gasteiger et al.), DimeNetPlusPlus (Gasteiger et al., 2020), TFN (Thomas et al., 2018), SE3_Transformer (Fuchs et al., 2020), EGNN (Satorras et al., 2021), SphereNet (Coors et al., 2018), PaiNN (Schütt et al., 2021)
2D Image	ImageMol (Zeng et al., 2022), CGIP-Image (Xiang et al., 2023), MaskMol (cheng et al., 2024)	More than 900 vision models based on timm (Wightman, 2019)
3D Geometry Image	IEM (Xiang et al., 2024a)	
Video	VideoMol (Xiang et al., 2024b)	

4.3 Modality Extractor Module

In order to obtain mode-specific input from the original molecular SMILES data, we define modality extractors as follows:

- **44 fingerprint extractors.** We predefine 6 types of 44 common molecular fingerprints in BenchMol, including circular- (ECFPx, FCFPx), path- (RDKx, HashTT), substructure- (MACCS), longer-version-, pharmacophore- (TPATF), and physicochemistry-based (RDKit descriptors) fingerprints.
- **2 types of sequence tokenizers.** We include 2 common tokenizers for processing molecular SMILES, which come from CHEM-BERT (Kim et al., 2021) and MoLFormer.
- **2 types of graph featurizers.** We incorporate 2 common graph featurizers, which come from Hu et al. (2020a) and OGB library (Hu et al., 2020b).
- **7 types of geometry featurizers.** We integrate 7 geometry graph construction methods based on Hu et al. (2020a), OGB library, Geom3D, and Uni-Mol, which covers the input formats required by existing geometric deep learning methods. In particular, for Geom3D, we generate 4 combinations of featurizers by pairwise combining whether to use fully connected edges and whether to use only node features.
- **2 types of image renderers.** We build two visualization renderers based on RDKit and PyMOL to generate 2D images, 3D geometry images, and videos, which are referenced by ImageMol, IEM, and VideoMol, respectively.

4.4 Model Initializer Module

After obtaining data of different modalities, we define several modality-specific factory to initialize model for extracting features. Table 1 shows the models based on different modalities supported in BenchMol. BenchMol not only supports existing pre-trained models, but also includes a large number of non-pre-trained basic models. Especially for vision-based models, BenchMol is compatible with the timm library (Wightman, 2019) and supports more than 900 vision models. Meanwhile, with the development of the timm library, BenchMol can automatically include the latest models.

4.5 Training Strategy Module

In training strategy module, we provide two training strategies, linear probing and fine-tuning. In linear probing, to improve efficiency, we define a modality-specific feature extractor and pre-extract features based on a given pre-trained model. Then, we define a trainer to train a fully connected layer directly on the features. In fine-tuning, given a model name, BenchMol will train the entire model on the given task.

4.6 Evaluation Metric Module

BenchMol supports multiple evaluation metrics for classification and regression tasks. For classification tasks, the metrics include Accuracy, Area Under Receiver Operating Characteristic Curve (ROC-AUC), F1-Score, Area Under the Precision-Recall Curve (AUPR), Precision, Recall, Kappa, Matthews. For regression tasks, the metrics include Mean Absolute Error (MAE), Mean Squared Error (MSE), Root-Mean Squared Error (RMSE), Spearman's Rank Correlation Coefficient, Pearson's Correlation Coefficient, Coefficient of Determination (R^2). Users have the liberty to select specific metrics for evaluating the model.

4.7 Use of BenchMol

BenchMol is a flexible and easy-to-use framework for MRL and you can find detailed user instructions in Appendix A. Through simple and direct code invocations (syntax is `from benchmol import package`), users can easily implement the entire pipeline of MRL from initial data loading to the final model evaluation. Specifically, the use case of image modality is shown in Appendix A.6. The entire pipeline just mentioned can be completed with only 4 lines of effective code.

5 Experiments

5.1 Experiments Settings

Settings. To ensure the fairness and comprehensiveness of the experimental results, unless otherwise stated, all experiments are performed under strictly consistent settings. Specifically, we use the same hyperparameter search range and report the test set results with the best validation performance on 12 molecular property prediction (MPP) benchmarks from MoleculeNet, 3 attribute datasets from MBANet, and a total of 60 molecular activity datasets of 6 different molecular types from StructNet. Meanwhile, we repeat the experiment 10 times with the same and large number of random seeds from 0 to 9 and report the mean and standard variance. See the Appendix E.1, the Appendix E.2 and the Appendix E.3 for details of baselines, hyperparameter search and training loss. The computational efficiency is discussed in the Appendix J.

Data Split and Metrics. All evaluation datasets are split into 80% training, 10% validation and 10% test sets. The 12 MPP datasets include 8 classification datasets and 4 regression datasets with a strict scaffold split (Hu et al., 2020a). We follow the

Table 2: Benchmarking 7 different modality methods on 8 classification tasks with average ROC-AUC (%) and 4 regression tasks with average RMSE performance from 12 MPP datasets. The modality types from top to bottom are fingerprint, sequence, graph, geometry graph, image, geometry image, and video. L means the number of layers, -I and -G mean the modalities are image and geometry image, respectively. Note that the geometry images and videos use the BGR format. The green background represents top-6 models in performance.

Model	Classification (↑)	Regression (↓)
mcfp4_2048	71.66	1.255
ecfp4_2048	69.81	1.300
maccs	70.54	1.302
physchem	63.16	1.454
atompair_2048	70.28	1.189
rdkDes	63.19	1.648
Chem-BERT-8L	73.41	1.093
MolFormer	69.58	1.293
EdgePred	60.73	1.665
ContextPred	66.90	1.461
infomax	66.35	1.409
masking	62.53	1.490
MolCLR	65.50	1.369
MoleBERT	72.28	1.320
GraphMVP	65.78	1.401
CGIP-Graph	67.05	1.552
Uni-Mol (10 conf)	74.13	1.144
ImageMol	62.63	1.507
MaskMol	63.03	1.441
CGIP-Image	61.94	1.556
IEM-I	60.95	1.577
IEM-G (10 conf)	70.29	1.212
VideoMol	69.03	1.222

suggestions of MoleculeNet and GraphMVP to use the ROC-AUC metric for classification tasks and the RMSE metric for regression tasks. In the remaining MBANet and StructNet benchmarks, we uniformly use RMSE for evaluation. We split MBANet benchmark by using ordered split. In StructNet benchmark, except for acyclic rule and acyclic chain rule which are random split, the others are strict scaffold split. This is because acyclic rule and acyclic chain rule cannot extract the scaffold.

Table 3: Effect of different numbers of conformations on 8 classification tasks (CLS) with ROC-AUC and 4 regression tasks (REG) with RMSE from 12 MPP datasets. -R means no pre-training and -G means the modalities are geometry image. The geometry images use the BGR format.

	Uni-Mol-R		Uni-Mol		ResNet18-G-R		IEM-G	
	CLS	REG	CLS	REG	CLS	REG	CLS	REG
1 conf	**65.80**	1.308	73.75	1.162	58.73	1.568	64.66	1.406
10 conf	65.53	**1.276**	**74.13**	**1.144**	**63.68**	**1.485**	**70.29**	**1.212**
δ	↓0.41%	↑2.51%	↑0.52%	↑1.55	↑8.43	↑5.29	↑8.71	↑13.8

5.2 Linear Probing on MoleculeNet

To evaluate the quality of features, we use the linear probing strategy to evaluate the performance of molecular encoders on 12 MPP datasets. Specifically, BenchMol extracts features from modality-specific data using a given encoder and trains and evaluates a single-layer fully connected network.

Findings. Table 2, Table 3 and Table 4 show the performance of encoders with different modalities in BenchMol. More detailed results about MPP in Appendix F and Tables S20, S21, S22, S23, S24. We summarize these findings as follows:

Table 4: The average ROC-AUC (%) and RMSE performance of non-pretrained methods on 8 classification tasks (CLS) and 4 regression tasks (REG) from 12 MPP datasets. The modality types from top to bottom are sequence, graph, geometry graph, image, geometry image, and video. L means the number of layers and -G means the modality is geometry image. Note that the geometry images and videos use the BGR format.

Model	CLS (↑)	REG (↓)
BERT-8L-R	68.94	1.264
MolFormer-R	70.40	1.319
GIN-R	63.21	1.576
Uni-Mol-R	65.80	1.308
Uni-Mol-R (10 conf)	65.53	1.276
ResNet18-R	55.34	1.682
ResNet18-G-R	58.73	1.568
ResNet18-G-R (10 conf)	63.68	1.485
VideoMol-R	61.16	1.526

1) **Models from 6 modalities are in the top 6 in performance.** In Table 2, the top 6 performances on the classification task are Uni-Mol (10 conf), Chem-BERT-8L, MoleBERT, mcfp4_2048, maccs, IEM-G (10 conf) and the top 6 performances on the regression task are Chem-BERT-8L, Uni-Mol (10 conf), AtomPair, IEM-G (10 conf), VideoMol, mcfp4_2048. Our analysis reveals that the top 6 include 6 modalities, except for the 2D image modality, indicating their advancement in linear probing. For the image modality, we find that it relies more on the fine-tuning stage. The Table S24 shows that ImageMol has a significant performance improvement from 62.5% to 71.9% after fine-tuning, with a performance improvement of 15.0%. There are two possible reasons: one is that there is too little information in 2D images to learn generalized knowledge in the pre-training stage, and the second is that the pre-training task still needs to be further improved. Our findings suggest using geometry images or videos to achieve better performance in vision-based representations. For more analysis of modal diversity, see Appendix K.1.
2) **Multiple conformations can significantly improve the performance of the geometry image.** Since molecules have multiple conformations, we compare here the performance differences between single and multiple conformations. Here we follow the suggestion of Zhou et al. and use 10 conformations. We use geometry-based methods (Uni-Mol-R, Uni-Mol) and geometery image-based methods (ResNet18-G-R, and IEM-G). Table 3 indicates that Uni-Mol-R and Uni-Mol have almost consistent performance between 1 conformation and 10 conformations, namely 65.8% v.s. 65.5% and 73.75% v.s. 74.13% (For Uni-Mol ablation on data scale see Appendix K.2). However, we observe that ResNet18-G-R and IEM-G have significant performance gains from 1 conformation and 10 conformations, i.e. 58.73% v.s. 63.68% and 64.66% v.s. 70.29%. For a detailed analysis of why multi-conformation has a large performance gain for geometric images, see Appendix K.3.

3) **Sequence-based methods have good inductive biases for molecular properties.** Table 4 shows the performance of non-pretrained models. Here, we exclude hand-crafted feature-based fingerprints and focus solely on discussing methods based on automatic feature extraction. We find that BERT-8L-R and MolFormer-R can achieve high performance on 12 MPP tasks without any pre-training, surpassing many other modality pre-training methods (such as GraphMVP, MolCLR, ImageMol, etc.). This provides evidence that the inductive bias based on the sequence is consistent with the molecule. Even without any training, the extracted features can retain the original molecular information. See Appendix K.4 and Appendix K.5 for a more detailed analysis.

4) **Geometry images and videos are not sensitive to RGB and BGR formats.** For images, RGB and BGR are two similar formats. Early OpenCV used BGR format by default and current PIL uses RGB format by default. Here we study the impact of RGB and BGR on the performance of geometry images and videos. Table S22 and Table S23 show the results for RGB. We find no significant difference between RGB and BGR for geometry images and videos. For example, VideoMol uses BGR (69.03%) better than RGB (68.53%) on classification tasks and RGB (1.190) is better than BGR (1.222) on regression tasks. The relationship between BGR and RGB is similar to image augmentation and the combination between them may further improve performance, just like TTA (Test Time Augmentation) (Kimura, 2021). We explore this observation in future research. See Appendix K.6 for a more detailed analysis.

5.3 Fine-tuning on MBANet

Findings. The MBANet benchmark defines tasks related to molecular attributes (atom, bond, molecular weight, etc.). Here we focus on deep learning representation and ignore fingerprint methods because some fingerprints directly contain this information. Table 5 shows the overall average results of models with different modalities on multi-tasks and the detailed results of each task are shown in Appendix G and Tables S25±S26, S27, S28±S29. We summarize the following findings:

Table 5: The RMSE performance on MBANet_{atom}, MBANet_{bond}, MBANet_{attr}, which are abbreviated as MBA_{atom}, MBA_{bond}, MBA_{attr}. *-R, TFN and SE3_Transformer represent non-pre-trained models, and the others are pre-trained models.

Models	MBA_{atom}	MBA_{bond}	MBA_{attr}
BERT-6L	0.522	2.641	11.448
BERT-8L	0.524	2.648	11.471
RoBERTa-12L	0.546	2.753	13.197
molformer-R	0.551	2.788	12.906
Chem-BERT-6L	0.521	2.666	11.349
Chem-BERT-8L	0.524	2.632	11.273
CHEM-RoBERTa-12L	0.534	2.776	13.145
Molformer	0.531	2.703	12.440
GIN-R	0.340	0.602	8.514
EdgePred	0.316	0.597	6.878
ContextPred	0.317	0.578	6.963
infomax	0.318	0.592	6.787
masking	0.332	0.593	6.536
MolCLR	0.325	0.631	7.993
MoleBERT	0.317	0.574	6.231
CGIP-Graph	0.231	0.588	4.255
GraphMVP	0.328	0.623	7.841
Uni-Mol-R (1 conf)	0.339	2.263	10.859
Uni-Mol (1 conf)	0.346	2.296	10.886
TFN	0.177	**0.309**	3.091
SE3_Transformer	0.503	1.892	11.659
ResNet18-I-R	0.350	1.630	6.951
ImageMol	0.352	1.601	6.509
CGIP-Image	0.337	1.591	7.018
MaskMol	0.532	2.656	11.764
IEM-I	0.348	1.670	6.769
ResNet18-G-R	0.268	1.586	4.848
IEM-G (1 conf)	0.243	1.560	4.916
ResNet18-V-R	0.156	1.048	2.660
IEM-V	**0.124**	0.965	**2.254**

5) **Video modality excels at tasks related to atoms and basic attributes**. In a comparison of a large number of non-pretrained methods, video-based method (ResNet18-V-R and IEM-V) achieve the best performance on MBANet_{atom} and MBANet_{attr} and geometry-based method (TFN) achieve the best performance on MBANet_{bond}. In particular, as shown in Table S25, we find that the video modality has obvious advantages over other modalities in learning simple MW (Molecular Weight), MR (Molar Refractivity), VE (Valence Electrons) and TPSA (Topological Polar Surface Area). See Appendix K.7 for a more detailed analysis.

6) **Almost all pre-training tasks can improve the performance of the model on atoms, bonds, and attributes.** We can observe that existing pre-training tasks can improve the performance of models on MBANet. For example, EdgePred, IEM-V improve the performance of their non-pretrained versions (GIN-R, and ResNet18-V-R) respectively.

7) **The performance of the same modality on MBANet is always similar.** We find that the performance fluctuations of models with the same modality are small. For example, the variance of the sequence-based model on MBANet_{atom} is only 0.011. This reflects the differences in feature extraction between different modalities and guides us in choosing the modality for specific tasks.

We further expand the data scale of MBANet and verify the validity of the findings in Appendix H.

5.4 Fine-tuning on StructNet

Findings. Table 6 shows the average performance of methods with different modalities on StructNet. We also report detailed results on in Appendix I and Tables S31±S32, S33±S34, S35±S36, S37±S38, S39±S40, S41±S42. We summarize the following findings:

Table 6: The average RMSE performance on AC (acyclic chain), A (acyclic), CC (complete chain), M (macro), MP (macrocyclic peptide) and R (reticular) of StructNet. We select the methods with the best average performance on 10 datasets from different modalities for presentation. Geom Image represents Geometry Image. The green background represents top-3 performance.

	AC	A	CC	M	MP	R
Fingerprint	10.619	12.508	**9.228**	18.290	9.436	2.478
Sequence	10.439	12.520	9.307	18.918	9.385	2.485
Graph	10.459	12.536	9.246	**18.109**	10.956	2.496
Geometry	**10.284**	**12.192**	9.249	18.596	9.424	2.471
Image	10.550	12.473	9.259	19.136	**9.321**	**2.454**
Geom Image	10.430	12.482	9.306	18.846	9.343	2.467
Video	10.441	12.444	9.352	18.923	9.339	2.469

8) **The geometry graph modality prefers acyclic (AC and A) molecules; The fingerprint and graph modalities prefer cyclic (CC and M) molecules; The visual-based modalities (Image, Geometry Image, and Video) prefer macrocyclic peptide (MP) and reticular (R) molecules.** Table 6 shows the experimental results of different modalities without pre-training. First, we find that geometry graph achieves the best performance on acyclic chain molecules and acyclic molecules, which indicates that it prefers acyclic molecules. Then, we find that fingerprint and graph modalities achieve good performance on complete chain molecules and macro molecules. We speculate that it may be suitable for cyclic molecular structures and further counted the number of rings in complete chain molecules and macro molecules. we find that more than 93.6% of the complete chain molecules and all macro molecules are cyclic, which indicates that graph prefers cyclic structures. Finally, we find that vision-based modalities can achieve the best performance on macrocyclic peptide molecules and reticular molecules. The top 3 in performance are image, video and geometry image modalities, which indicates that vision-based modalities prefer macrocyclic peptide molecules and reticular molecules. We also provide the details of significance test in Appendix K.8.
9) **Pre-training tasks may fail for certain types of molecules.** On MBANet benchmark, we conclude that almost all pre-training tasks are effective. However, in StructNet benchmark, many pre-training tasks may fail. For example, graph-based pre-training tasks achieve worse performance than unpre-trained GIN in complete chain molecules in Table S35 and image-based pre-training tasks achieve worse performance than unpre-trained ResNet18 in reticular molecules in Table S41. This suggests that in molecules with certain specific types, we need to design special pre-training for them to improve performance.

6 Conclusion

We first proposed a unified and flexible platform supporting different molecular modalities, called BenchMol, to promote reproducibility in the molecular representation learning (MRL) community, which provides the entire pipeline from raw data to final model evaluation and ensures fair and comprehensive benchmarking. Subsequently, we proposed two new benchmarks, MBANet and StructNet, to explore the performance and preferences of existing models on different modalities. Finally, we used BenchMol to train at least 57,060 models and provided many meaningful insights. BenchMol reviews and integrates mainstream MRL models with 7 different modalities, allowing researchers to easily understand these models and quickly iterate on them.

Limitations and Future Works: Currently, BenchMol does not support multi-modal fusion. In the future, we plan to continue maintaining BenchMol and upgrading it to a multi-modal fusion platform. This enhanced platform will be capable of further improving molecular representation and tackling interaction-based tasks such as drug-drug interaction (DDI), drug-target interaction (DTI), and protein-protein interaction (PPI). Furthermore, while BenchMol currently focuses on evaluating its performance in MRL, it is important to note that its applicability is not restricted to the molecular domain. We hope to leave the exploration of more fields to the community for verification.

7 REPRODUCIBILITY STATEMENT

For reproducibility, we open sourced BenchMol and made all data publicly accessible at Github.

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Appendix

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A BENCHMOL TOOLKIT USER GUIDE

A.1 OVERVIEW

Here, we will go through the usage instructions of BenchMol in detail to demonstrate the flexibility and user-friendliness of BenchMol. In addition to the two contributed benchmarks (MBANet and StructNet), user interfaces of BenchMol are divided into four modules: Modality Extractor, Model Initializer, Training Strategy, and Evaluation Metric. Since words alone cannot fully demonstrate the full functionality of BenchMol, we next will focus on describing the main features here.

A.2 MODALITY EXTRACTOR

Given a molecule, BenchMol can transform this molecule into different modalities, including fingerprint (can be viewed as a row-type or table-type modality), sequence, graph, geometry, image, and video. We describe several main modality extraction steps in detail below:

- **Fingerprint Modality.** Listing 1 shows the script for extracting 44 fingerprints. By changing the parameter fp_name, different fingerprint types can be extracted.
- **Sequence Modality.** BenchMol automatically extracts sequence modality from SMILES by using a predefined function *collate()*, which tokenizes a batch of molecular data.
- **Graph and Geometry Modalities.** Listing 2 shows the script for extracting geometry modality from molecular sdf file. By changing the parameter graph_feat_extractor, different geometry types can be extracted. For graph modality, the extraction process is similar to that of geometry modality, which defines two methods to generate graphs.
- **Image Modality.** BenchMol defines a function *loadSmilesAndSave(smiles, path)* which takes a smiles as input saves the molecule image to a path.
- **Geometry Image and Video Modalities.** BenchMol defines a method with parameter img_type to extract different types of visual modalities by changing the value of img_type.

```python
from benchmol.data_process.molecules import FPGeneration

fg = FPGeneration()
features = fg.get_fingerprints(
    df, fp_name="maccs", smiles_column_name="smiles"
)
```

Listing 1: Transforming process from molecules to fingerprint modality.

```python
if graph_feat_extractor == "ogb":
    x, edge_index, edge_attr, coords = mol_to_3d_graph_data_ogb(sdf_path)
elif graph_feat_extractor == "jure":
    x, edge_index, edge_attr, coords = mol_to_3d_graph_data_jure(sdf_path
    )
elif graph_feat_extractor == "geom3d":
    x, edge_index, edge_attr, coords = mol_to_graph_data_obj_simple_3D(
    sdf_path, pure_atomic_num=False)
elif graph_feat_extractor == "geom3d_pure_atomic_num":
    x, edge_index, edge_attr, coords = mol_to_graph_data_obj_simple_3D(
    sdf_path, pure_atomic_num=True)
elif graph_feat_extractor == "geom3d_full_edge":
    x, edge_index, edge_attr, coords =
    mol_to_graph_data_obj_simple_3D_full_edge(sdf_path, pure_atomic_num=
    False)
elif graph_feat_extractor == "geom3d_pure_atomic_num_full_edge":
    x, edge_index, edge_attr, coords =
    mol_to_graph_data_obj_simple_3D_full_edge(sdf_path, pure_atomic_num=
    True)
elif graph_feat_extractor == "unimol":
    atoms, coords, smi, scaffold = unimol_data(sdf_path)
else:
```

```
    raise Exception("graph_feat_extractor {} is undefined".format(graph_feat_extractor))
```

Listing 2: Transforming process from molecules to geometry modality.

A.3 Model Initializer

As shown in Listing 3, BenchMol define 4 factories to initialize models for handling different modalities, including SmilesModelFactory, GraphModelFactory, GeometryModelFactory, and ImageModelFactory. Through these factory classes, we can easily initialize various models by giving the model name and necessary parameters (such as the configuration of the neck model related to the task).

```
# SmilesModelFactory for sequence modality
class SmilesModelFactory(torch.nn.Module):
    def __init__(self, model_name, head_arch, num_tasks, vocab_path, d_dropout=0, head_arch_params=None, pretrain_path=None, device="cpu", **kwargs):
        ...
    def forward(self, batch):
        ...
    def get_model(self):
        ...

# GraphModelFactory for graph modality
class GraphModelFactory(torch.nn.Module):
    def __init__(self, model_name, head_arch, num_tasks, head_arch_params=None, pretrain_gnn_path=None, model_key=None, num_layer=5, emb_dim=300, JK="last", dropout=0.5, graph_pooling="mean", gnn_type="gin", update_predictor=True, **kwargs):
        ...
    def forward(self, batch):
        ...
    def get_model(self, update_predictor=True):
        ...

# GeometryModelFactory for geometry modality
class GeometryModelFactory(torch.nn.Module):
    def __init__(self, model_name, head_arch, num_tasks, head_arch_params=None, pretrain_gnn_path=None, model_key=None, emb_dim=300, args=None, **kwargs):
        ...
    def forward(self, batch):
        ...
    def get_model(self, args, num_tasks, node_class, edge_class):
        ...

# ImageModelFactory for image/geometry-image/video modality
class ImageModelFactory(torch.nn.Module):
    def __init__(self, model_name, head_arch, num_tasks, pretrained=False, head_arch_params=None, **kwargs):
        ...
    def forward(self, x):
        ...
    def get_model(self):
        ...
```

Listing 3: The model factories of different modalities.

A.4 Training Strategy

BenchMol provides two training strategies, linear probing and fine-tuning.

Linear Probing. To improve the efficiency of linear probing, BenchMol designs 6 extractors to extract feature representations from molecules of different modalities. A core idea of these extractors is that given a pre-trained model and a molecule, the feature extractor will return features of a specific modality. As shown in 4, BenchMol defines an interface FeatureExtractor, which is an abstract class with two abstract methods: extract_features() and return_features(). Subsequently, we defined an implementation class for each modality to extract features from the corresponding modality. The specific details class is as follows:

- **FingerprintExtractor.** The features of fingerprint can be extracted by giving the fingerprint name and SMILES sequence.
- **SmilesFeatureExtractor.** The sequence features are generated from molecular SMILES sequences.
- **GraphFeatureExtractor.** The graph features can be extracted from graph modality given a pre-trained graph model.
- **ImageFeatureExtractor.** The image feature can be extracted from image modality given a pre-trained image model.
- **MCImageFeatureExtractor.** The features of geometry image and video can be extracted given a corresponding pre-trained model.
- **MVImageFeatureExtractor.** The extractor is used to extract features from molecular geometry images or videos with multiple conformers.

```
@dataclasses.dataclass
class FeatureExtractor(abc.ABC):

    @abc.abstractmethod
    def extract_features(self):
        pass

    @abc.abstractmethod
    def return_features(self):
        pass
```

Listing 4: General interface of modality extractor.

Fine-Tuning. Fine-tuning refers to training the pre-trained model and the network related to the downstream task at the same time. With BenchMol, fine-tuning of different modes can be achieved very easily. Appendix A.6 shows fine-tuning on image modality. Similar to the image modality, we can also easily implement fine-tuning of other modalities.

A.5 Evaluation Metric

BenchMol provides a large number of metrics for classification and regression tasks and users can switch between different evaluation metrics at will by specifying the eval_metric parameter. The following shows the evaluation indicators supported by different tasks:

- **Classification task:** accuracy, ROC-AUC, F1-score, AUPR, Precision, Recall, Kappa coefficient, Matthews coefficient.
- **Regression task:** MAE, MSE, RMSE, Ppearman coefficient, Pearson coefficient, R^2 coefficient.

A.6 Use Case for Image Modality

We assume that there is a requirement: there are a batch of molecular images and corresponding binary labels of active or inactive. We first need to divide these data into training set, validation set and test set. Then, we need to use a ViT network to train on the training set and use the ROC-AUC metric to evaluate on the validation set. Finally, we need to obtain the results of the test set based on the best performance on the validation set. We can easily achieve this with BenchMol. Listing 5 shows instructions for using BenchMol in image modality, which shows the flexibility and ease of use of BenchMol. We can see that only 4 lines of effective code are needed to complete the training of the model, which are lines #10, #15, #23, and #28.

```python
from benchmol.dataloader.image_dataset import TrainValTestFromCSVFactory
from benchmol.trainer import Trainer
from benchmol.model_pools import ImageModelFactory

# Take the image modality and classification task as an example
modality="image"
task_type = "classification"

# define Model with backbone as ViT and neck as arch4 for n tasks
model = ImageModelFactory(
    model_name="vit_small_patch16_224", head_arch="arch4",
    head_arch_params={"inner_dim": 128, "dropout": 0.2, "activation_fn":
    "gelu"}, num_tasks=2
)

# define Dataset
factory = TrainValTestFromCSVFactory(
    dataroot, csv_path, data_type="image", image_dir_name="image",
    task_type=task_type, batch_size=16, num_workers=8
)
train_loader = factory.get_dataloader(split="train")
valid_loader = factory.get_dataloader(split="valid")
test_loader = factory.get_dataloader(split="test")

# define Trainer
trainer = Trainer(
    model, modality, train_loader, valid_loader, test_loader, task_type,
    criterion=nn.BCEWithLogitsLoss(reduction="none"), optimizer=Adam(
    model.parameters(), lr=0.001, weight_decay=1e-5), device="cuda:0"
)

# training and evaluation
results = trainer.train(num_epochs=100, eval_metric="ROCAUC",
    valid_select="max", min_value=-np.inf, save_finetune_ckpt=True,
    save_dir="./experiments/")

# Output model results
print("results: {}\n".format(results))
```

Listing 5: The use case on image modality with classification task and ROC-AUC metric.

B Visual Rendering of Molecule

We describe in detail how to render molecules as images, geometry images, and videos:

- **Rendering of image:** Following CGIP (Xiang et al., 2023), the molecualr image is generated by the *MolsToGridImage()* method of RDKit. This method takes the SMILES sequence of a molecule as input and generates an image of length 224, width 224, and 3 channels.
- **Rendering of geometry image:** Molecular images represent the 2D planar structure of molecules. However, molecules have three-dimensional conformational information. Following IEM (Xiang et al., 2024a), we use PyMOL to render the 3D structure of the molecule. Since the 3D structure is easily obscured when displayed on a single image, 4 viewing angles are used to render the molecule from different angles. Therefore, through multi-view rendering with PyMOL, we can obtain 4 geometric images with different vies for a molecule, which can be formulated as a matrix of $4 \times 224 \times 224 \times 3$.
- **Rendering of video:** VideoMol (Xiang et al., 2024b) represents the 3D image of a molecule as a video. Specifically, a molecular video is constructed by rotating a molecule along the x-axis, y-axis, and z-axis. During the rotation process, VideoMol captures 60 frames at equal intervals to represent the molecular video. Therefore, we use PyMOL to render the molecule with 3D information and generate a 60-frame video, which can be formulated as a matrix of $60 \times 224 \times 224 \times 3$.

C Discussion

C.1 Motivation and Potential Impact of BenchMol

Motivation. The main motivations for BenchMol are to address the following research gaps in the field of chemical machine learning: (1) There is a lack of fair and comprehensive evaluation of methods across different modalities; (2) The strengths and differences of different modalities are still unknown, which limits the development of multi-modal fusion; and (3) There is a lack of a unified and easy-to-use platform to integrate methods across different modalities. With BenchMol, researchers can easily use and compare various molecular representation learning methods.

Potential Impact. The two benchmarks MBANet and StructNet proposed in this paper have important impacts on promoting certain chemical problems, which are summarized as follows:

- MBANet aims to evaluate the ability of different methods to capture basic molecular information that is critical for many chemical problems. For example, the distribution of atoms and bonds is crucial for understanding the three-dimensional structure of a molecule [1] and aids in molecular dynamics simulations [2]. Predicting the basic attributes of a molecule helps design molecules with specific properties [3]. Attributes such as molecular weight, MolMR (molecular refractive index), and NumHDonors (number of hydrogen bond donors) can be used to infer the biological activity of a molecule and its interaction with its target [4].
- StructNet aims to explore the preferences of various methods for molecules of different structural types and it is of great significance for certain specific targets. For example, molecules targeting KRAS targets are often macromolecules, and the model needs to learn and predict in the sample space of macromolecules [5]; a class of antiviral and antimalarial drugs usually have chain-like molecular structures, while molecules targeting fibroblast activation protein (FAP) usually exhibit non-cyclic structures. These chemical preferences indicate that it is important to select appropriate molecular modalities to work more effectively in different scenarios.

C.2 Motivation and Practicality of Benchmark Datasets

MBANet. The motivation and practicality of MBANet are as follows:

- **Motivation.** Currently, a large number of benchmarks focus on mapping molecules to complex properties or biological activities. However, it is a complex process for models to learn to map molecules to complex properties or biological activities, which may be related to the regulatory network of molecules from a microscopic perspective. This complex process is not conducive to describing the model's understanding of the basic properties of molecules. In this paper, we hope to clarify the understanding of the most basic attributes of molecules by different modalities. This basic attribute reflects the properties directly related to the molecule and has nothing to do with the complex regulatory network. This has always been a research gap but is also crucial for the model to understand molecules. Therefore, we design MBANet to evaluate the model's ability to understand the basic attributes of molecules.
- **Practicality.** From a practical point of view, the basic attributes evaluated by MBANet are closely related to complex properties or biological activities. For example, LogP is an important indicator of BBBP and it and TPSA are two key physicochemical parameters in drug design (Prasanna & Doerksen, 2009), affecting drug absorption, distribution, metabolism, excretion and toxicity.

We hope that this simple, decoupled basic task can provide reference and more thinking for modality selection on related active tasks. In addition, we will consider providing more complex prediction tasks for MBANet in future work to make it more comprehensive. However, in this paper, we prefer to study this unique aspect.

StructNet. We describe the motivation and practicality of classifying molecules into different types based on their 2D structural patterns (e.g., acyclic, acyclic chain, cyclic chain, macrocyclic peptide, macromolecule, and reticular molecule). Molecular structure is intimately linked to molecular properties, so that certain drug targets exhibit preferences for specific molecular structures, and some therapeutic applications correspond to molecules with particular structures. For instance:

- **Macrocyclic peptides** are increasingly being recognized for their therapeutic potential in targeting aberrant protein-protein interactions (PPIs), with several macrocyclic peptide-based oncology drugs already approved by the U.S. Food and Drug Administration (FDA) for clinical application (Yang et al., 2022).
- Due to the mechanism of action of the KRAS target involving covalent binding with a cysteine generated by the mutation of glycine at the 12th position in proteins, the designed molecules tend to have a larger molecular weight, resulting in a relatively larger binding area, more stable binding, and stronger specificity. Therefore, molecules targeting KRAS targets tend to be **macromolecules** (Cox & Der, 2024).
- **Acyclic single-chain** fragment variable (scFv) molecules target fibroblast activation protein (FAP) (Baum et al., 2007) and have applications in CAR-T cell therapy (Niu et al., 2024; Loureiro et al., 2023).
- As structural analogues to the **chain molecules** defined herein, acyclonucleoside phosphonates (ANPs) exhibit a distinctive acyclic structure (Bessières et al., 2024), which constitute a significant class of compounds with antiviral and anticancer properties (Holỳ, 2006), and they harbor substantial potential as candidates for antimalarial drug development (Cheviet et al., 2020).

Thus, we have introduced StructNet, which categorizes based on 2D structural patterns, with the aim of offering insights and suggestions on model selection for drug design targeting specific structures and the future optimization directions for various models.

C.3 The Importance of Molecular Visual Modalities

Currently, molecular visual modalities consist of image, geometry image, video. Compared with previous molecular representations (such as SMILES or graph), the importance of molecular visual modalities is reflected in the following four aspects:

- **Direct representation of structural and geometric information**: Compared with previous SMILES or graph representations, molecular visual modalities naturally retain structural and geometric properties such as atom type, chemical bond type, bond angle, spatial conformation, etc. through pixel information (color, texture, etc.). Molecular videos can further describe the dynamic information of molecules, which is particularly important for tasks involving molecular geometry or dynamic behavior;
- **Stronger interpretability**: Molecular visual modalities show the molecular structure and its dynamic behavior in an intuitive form, which is convenient for humans to understand and analyze the features learned by the model. For example, through technologies such as GradCAM (Selvaraju et al., 2017), researchers can intuitively understand how the model makes decisions;
- **Enriching molecular representation technology**: Molecular visual modalities can use another technology stack (computer vision) to extract potential features from molecular images, geometry images or videos, enriching existing molecular representation technology;
- **Independence of the number of atoms and bonds**: Molecular visual modalities are pixel-based representations, which are independent of the number of atoms and bonds. At present, drug discovery is increasingly biased towards large molecules. Significantly different from SMILES and graph, visual modalities have the unique advantage that the computational cost does not increase with the number of atoms and bonds, which will be of great benefit in drug development of large molecules.

Molecular visual modalities are particularly suitable for the following tasks, which rely on the spatial configuration or dynamic characteristics of molecules:

- **General molecular representation**: Image-, geometry-image- and video-based methods have achieved excellent performance in various drug discovery tasks, just like other molecular representation methods (as shown in Table S20 and Table S21, etc.). Therefore, they can be used for general representation of drug discovery tasks;
- **Multi-modal fusion**: Obviously, visual modalties are different from previous molecular representation learning methods. Through multi-modal fusion or cross-modal contrastive learning tasks, more diversity will be provided to further improve the performance of drug discovery. As shown in Appendix K.1, Table S46 and Table S47, video has the largest differences with other modalities;

- **Tasks related to atomic molecular distribution and basic properties**: As shown in Table 5, video modality can achieve the best performance on atom-level and attr-level tasks of MBANet. Therefore, the video modality makes up for the lack of understanding of atomic distribution and basic properties of other modal representations;
- **Virtual screening of macrocyclic or reticular structures**: As shown in Table 6, images and videos can achieve the best performance on MP and R tasks on StructNet. Therefore, images or videos make up for the lack of understanding of macrocyclic peptide (MP) and reticular (R) structures by other modalities, such as virtual screening of PPI targets (Cox & Der, 2024).

In general, molecules exist in the physical world. Currently, due to the limitations of molecular imaging technology, molecular images/geometry images/videos are obtained by economical image rendering technology. However, with the continuous advancement of molecular imaging technology, it is promising to directly represent molecules and inference about them in a visual way.

D BENCHMARKS

Here, we present the details of the two proposed sets of benchmarks: MBANet and StructNet.

D.1 MBANET

MBANet is used to evaluate the performance of deep learning models in understanding the basic information of molecules, aiming to measure whether the model can effectively capture the low-level core features related to molecules. In the fields of cheminformatics and drug development, the basic information of molecules (such as atomic information, bond information, molecular weight, TPSA and other basic attributes) is crucial for predicting molecular activity, drug-target interactions, and chemical reactivity. Therefore, MBANet attempts to examine the model's capabilities at these basic levels and explore whether the model can accurately capture these basic features by learning molecules.

MBANet has a total of 10,000 molecules and includes three groups of prediction tasks, namely prediction of atoms MBANet_{atom}, bonds MBANet_{bond} and basic attributes MBANet_{attr}. We check MBANet for duplication according to canonical SMILES and find that only 25 molecules are duplicated in canonical SMILES. Therefore, the impact on the evaluation is negligible. In the MBANet benchmark, the model needs to predict the atomic distribution, bond distribution, and basic attributes given a molecule. The Figure S1, Figure S2, and Figure S3 show the distribution information of MBANet_{atom}, MBANet_{bond} and MBANet_{attr} respectively. In particular, we describe the meaning of each attribute in MBANet_{attr} in detail as follows:

- **Molecular Weight:** The sum of the relative atomic masses of all atoms in a molecule, usually expressed in daltons (Da) or grams per mole (g/mol). It is a fundamental property that affects a compound's physical and chemical behavior.
- **MolLogP (LogP):** The logarithmic value of the distribution coefficient ratio (P) of a compound in n-octanol (oil) and water. It indicates the hydrophobicity or lipophilicity of the molecule, with higher values suggesting greater affinity for lipid environments.
- **MolMR (Molecular Refractivity):** A calculated property that reflects the volume and polarizability of a molecule. It is used to estimate the interactions of the molecule with its environment, including its ability to penetrate biological membranes.
- **BalabanJ (Balaban's J Index):** A topological index that measures the complexity of a molecular structure. It is used in quantitative structure-activity relationship (QSAR) studies to correlate molecular structure with biological activity.
- **NumValenceElectrons (Number of Valence Electrons):** The total number of valence electrons present in the atoms of a molecule. This property is important for understanding the chemical reactivity and bonding behavior of the compound.
- **TPSA (Topological Polar Surface Area):** A measure of the polar surface area of a molecule, calculated based on its structure. TPSA is often used to predict a compound's absorption, permeability, and bioavailability.

- **NumHAcceptors (Number of Hydrogen Bond Acceptors):** The count of atoms in a molecule that can accept hydrogen bonds, typically involving oxygen and nitrogen atoms. This property is crucial for evaluating molecular interactions in biological systems.
- **NumHDonors (Number of Hydrogen Bond Donors):** The count of atoms that can donate hydrogen bonds, usually hydrogen atoms attached to electronegative atoms like nitrogen or oxygen. This property influences a molecule's interaction with biological targets.

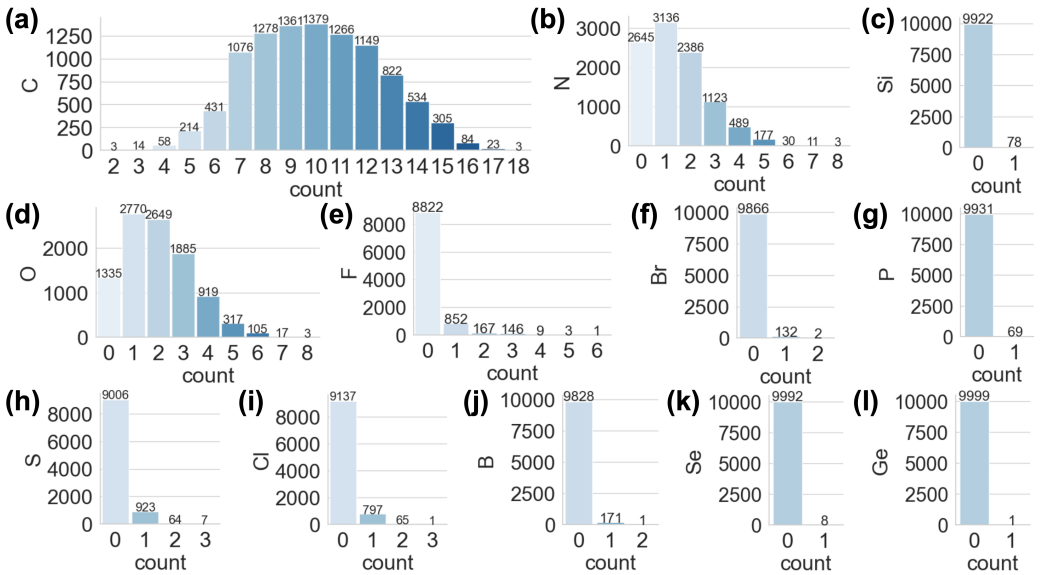

Figure S1: The distribution figures of MBANet$_{atom}$. (a)-(i) represent the distribution information of C, N, Si, O, F, Br, P, S, Cl, B, Se, Ge respectively.

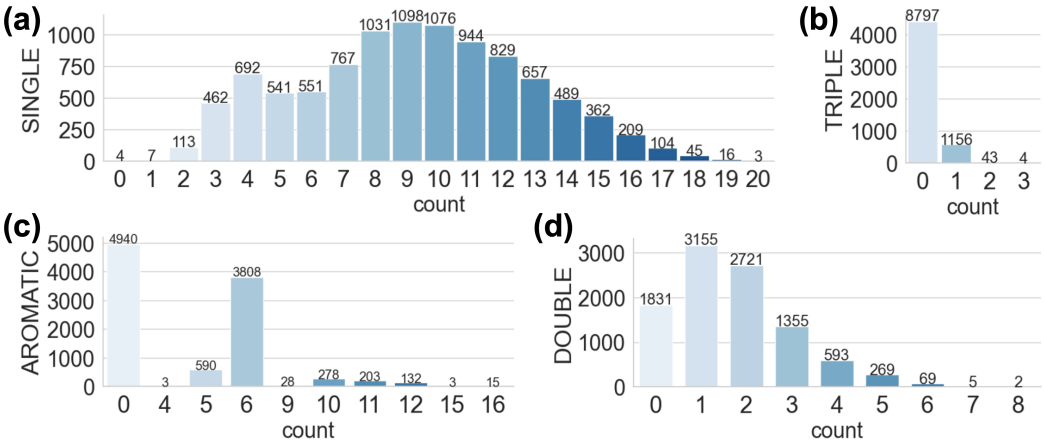

Figure S2: The distribution figures of MBANet$_{bond}$. (a)-(d) represent the distribution information of single, triple, aromatic, double bonds respectively.

MBANet Task Settings and Limitation Analysis. The reasons why we incorporate atoms with skewed distribution histograms into MBANet and design MBANet as a regression task and the evaluation metric of RMSE are as follows:

- **Reasons for including atoms (Si, Br, P, S, Cl, B, Se, Ge) and bonds (TRIPLE) with skewed distribution histograms**. This consideration is mainly to reflect the wide applicability of the evaluation. Even if the data distribution of some atoms and bonds is skewed, they reflect the actual chemical distribution. For the special case of Ge, it is a good choice to remove the task of

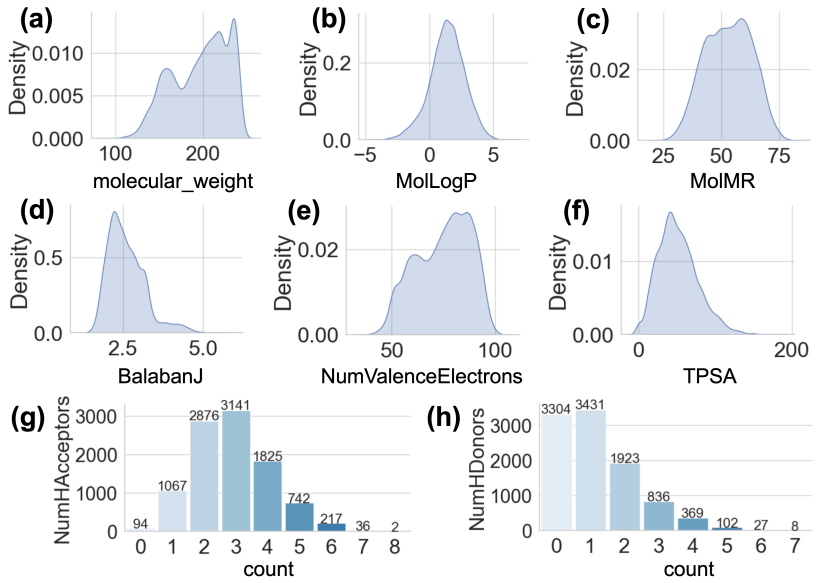

Figure S3: The distribution figures of MBANet$_{attr}$. (a)-(h) represent the distribution information of molecular weight, MolLogP, MolMR, BalabanJ, NumValenceElectrons, TPSA, NumHAcceptors, NumHDonors respectively.

this atom. But we still prefer to keep it. In detail, in our training process, the absence of Ge atoms is also a supervision label, that is, the number of Ge atoms is 0. Therefore, when training, the model needs to predict that there is no Ge in a molecule, which also reflects the learning ability of the model in this case. In addition, we can also see from Table S25 that Ge has little effect on the results, which will not affect the main conclusions given in the paper. Therefore, we tend to keep these atoms and bonds with skewed distributions;

- **Reasons for choosing regression tasks**. Given that molecules have a wide range of chemical space, we cannot exhaustively enumerate the number of atoms and bonds because they may continue to increase. In addition, if the model has the ability to count the distribution of atoms and bonds, it should be able to get answers from molecules with any number of atoms and bonds. If a classification task is used, the model will not be able to expand the distribution. For example, we set the distribution of C to 10 classification tasks from 0 to 9. Then for a molecule with more than 10 C, the model will not be able to make accurate predictions. Therefore, we design this task as a regression task rather than a classification task.
- **Reasons for using RMSE evaluation**. The reason why we choose to use RMSE (Root Mean Squared Error) instead of Spearman or Pearson correlation coefficient in regression tasks is mainly because the correlation coefficient only reflects the relative ranking or linear correlation between the predicted value and the true value and cannot directly quantify the actual error of the prediction. In the task of predicting the distribution of atoms and bonds, we need the model to accurately predict the number of certain atoms or bonds in a molecule. Therefore, we prefer to use RMSE to evaluate the performance of the model.

Although atoms with skewed distribution histograms have little impact on the final conclusion, we have to acknowledge that MBANet may be affected by atoms with extremely skewed count histograms, which may introduce noise to the final metrics.

D.2 StructNet

StructNet has a total of 6 different molecular types, including reticular (R)-, acyclic (A)-, complete chain (CC)-, acyclic chain (AC)-, macrocyclic peptide (MP)-, macro (M)-molecules. The scenarios corresponding to these six different types of molecules are of great significance in drug discovery. For example, molecules targeting KRAS targets tend to be large molecules and the model needs to

learn and predict in the sample space of large molecules (Cox & Der, 2024). Here, we first give the specific 6 screening rules for each type of molecule, as follows:

- **R rule for reticular molecules.** Reticular molecules exhibit better structural stability, drug loading capacity, and controllable release due to their complex cross-linked pore structure and larger specific surface area, making them suitable for multiple fields such as drug carriers, tissue engineering, and drug controlled release systems. Because reticular molecules are highly cross-linked structures composed of multiple interwoven molecular chains, similar to a network mesh. The more cross-linking (chemical bonds) between atoms, the higher the average degree. So we define molecules with average degree greater than 2.33 as reticular molecules.
- **A rule for acyclic molecules.** Acyclic molecules usually have higher reactivity and spatial adaptability, and can better bind to active sites that cyclic structures cannot reach smoothly due to steric hindrance in cyclic structures. Thus, We retain molecules with ring number equal to 0.
- **CC rule for complete chain molecules.** Compared to other structural molecules, chain molecules serve as drug molecules or carriers, based on their better flexibility and adaptability, high specificity and affinity, targeting specific biological targets to achieve precise treatment or drug delivery. Chain molecule is a long chain structure composed of a main molecular chain with few branches, where most of the atoms have a degree of 2. Therefore, We retain molecules where the ratio of branched atoms (atoms with degree greater than 2) to the total number of atoms is less than 0.2.
- **AC rule for acyclic chain molecules.** Strictly speaking, chain molecules do not have cyclic structures. Therefore, based on custom chain molecules, we retain molecules where the ratio of branched atoms (atoms with degree greater than 2) to the total number of atoms is less than 0.255 and the number of rings is equal to 0 as acyclic chain molecules.
- **MP rule for macrocyclic peptide molecules.** As one of the hot topics in the field of drug development today, macrocyclic peptides have high targeting, excellent pharmacokinetic properties, and low immunogenicity. Some macrocyclic peptides even have strong penetration ability. These advantages make macrocyclic peptides have broad application prospects in drug development and treatment. Therefore, we chose macrocyclic peptides as a class of structural molecules to study. We retain molecules with a maximum number of rings greater than 12 and a number of peptide bonds greater than 0 according to the definition of macrocyclic peptide, which refer to compounds connected by peptide bond (amide bond) and possessing a large cyclic structure.
- **M rule for macro molecules.** Large molecule targeted drugs, such as monoclonal antibodies, generally act on targets on the cell surface and have strong specificity. Moreover, compared to traditional small molecule targeted drugs, large molecule drugs have a longer half-life, which greatly reduces the frequency of medication. Therefore, We keep molecules with molecular weight greater than 900 as macro molecules.

After filtering according to the above rules, for each molecular type, the top 10 assay with the largest number of molecules are selected to further construct the StructNet, which means that each molecular type has 10 datasets. In particular, for multiple trials on the same SMILES, we only keep one and use the mean of multiple trials as the label. We show the statistics of the datasets with different molecular types in Tables S1, S2, S3, S4, S5 and S6.

Table S1: The 10 datasets composed of **reticular** molecules in StructNet, called R, with scaffold split.

Assay ChEMBL ID	Train/Valid/Test	Standard Type
CHEMBL4888485	512/64/65	Inhibition(%)
CHEMBL1614458	391/49/49	Potency(nM)
CHEMBL1614459	277/35/35	Potency(nM)
CHEMBL1613914	241/30/31	Potency(nM)
CHEMBL1614421	161/54/54	Potency(nM)
CHEMBL1614087	155/52/52	Potency(nM)
CHEMBL1614249	148/49/50	Potency(nM)
CHEMBL1614236	139/46/47	Potency(nM)
CHEMBL1614544	130/44/44	Potency(nM)
CHEMBL1614038	127/43/43	Potency(nM)

Table S2: The 10 datasets composed of **acyclic** molecules in StructNet, called A, with random split.

Assay ChEMBL ID	Train/Valid/Test	Standard Type
CHEMBL4513082	271/34/34	Inhibition(%)
CHEMBL4495582	271/34/34	Inhibition(%)
CHEMBL1614458	246/31/31	Potency(nM)
CHEMBL4303805	164/56/56	Inhibition(%)
CHEMBL4808149	164/56/56	Inhibition(%)
CHEMBL4296187	164/56/56	Inhibition(%)
CHEMBL4808150	163/55/55	Inhibition(%)
CHEMBL4296188	153/52/52	Inhibition(%)
CHEMBL4649955	153/51/51	Percent Effect(%)
CHEMBL4649949	153/51/51	Percent Effect(%)

Table S3: The 10 datasets composed of **complete chain** molecules in StructNet, called CC, with scaffold split.

Assay ChEMBL ID	Train/Valid/Test	Standard Type
CHEMBL4649949	1410/176/177	Percent Effect(%)
CHEMBL4649948	1410/176/177	Percent Effect(%)
CHEMBL4649955	1393/174/175	Percent Effect(%)
CHEMBL4888485	1336/167/167	Inhibition(%)
CHEMBL4296187	1005/126/126	Inhibition(%)
CHEMBL4296188	956/120/120	Inhibition(%)
CHEMBL4296802	907/113/114	Inhibition(%)
CHEMBL1614459	852/106/107	Potency(nM)
CHEMBL1614458	689/86/87	Potency(nM)
CHEMBL1614530	540/68/68	Potency(nM)

Table S4: The 10 datasets composed of **acyclic chain** molecules in StructNet, called AC, with random split.

Assay ChEMBL ID	Train/Valid/Test	Standard Type
CHEMBL1614458	145/49/49	Potency(nM)
CHEMBL4513082	137/47/47	Inhibition(%)
CHEMBL4495582	137/47/47	Inhibition(%)
CHEMBL4296187	124/42/42	Inhibition(%)
CHEMBL4296188	115/39/39	Inhibition(%)
CHEMBL1614361	110/38/38	Potency(nM)
CHEMBL4303805	108/36/36	Inhibition(%)
CHEMBL4649955	107/36/36	Percent Effect(%)
CHEMBL4649949	107/36/36	Percent Effect(%)
CHEMBL4649948	107/36/36	Percent Effect(%)

Next, we describe the details of the standard type. The standard type in the ChEMBL database refers to the type of biological or chemical measurement that is being recorded in the database for a particular bioactivity or assay, which defines the specific biological endpoint or property that has been measured for a compound. The standard types included in StructNet benchmarks are as follows:

Table S5: The 10 datasets composed of **macrocyclic peptide** (MP) molecules in StructNet, with scaffold split.

Assay ChEMBL ID	Train/Valid/Test	Standard Type
CHEMBL4888485	384/48/48	Inhibition(%)
CHEMBL2354301	336/42/42	AC50(nM)
CHEMBL3880198	168/56/57	Ki(nM)
CHEMBL4420271	150/50/51	Inhibition(%)
CHEMBL4419595	150/50/51	Inhibition(%)
CHEMBL4420282	136/45/46	IC50(nM)
CHEMBL3214979	129/44/43	AC50(nM)
CHEMBL4420277	124/42/42	Inhibition(%)
CHEMBL4419601	124/42/42	Inhibition(%)
CHEMBL4419606	124/42/42	IC50(nM)

Table S6: The 10 datasets composed of **macro** molecules in StructNet, called M, with scaffold split.

Assay ChEMBL ID	Train/Valid/Test	Standard Type
CHEMBL4420282	1125/141/141	IC50(nM)
CHEMBL4419606	985/123/124	IC50(nM)
CHEMBL4420281	580/72/73	Inhibition(%)
CHEMBL3881498	569/71/72	Inhibition(%)
CHEMBL4419605	568/71/72	Inhibition(%)
CHEMBL4420271	555/69/70	Inhibition(%)
CHEMBL4419595	555/69/70	Inhibition(%)
CHEMBL3881499	548/69/69	IC50(nM)
CHEMBL4420273	418/52/53	Inhibition(%)
CHEMBL4419597	418/52/53	Inhibition(%)

- **Inhibition** means the inhibition rate under certain conditions, and its unit is %.
- **Potency** means the dosage at which a drug achieves a certain pharmacological effect, and its unit is nM.
- **Percent Effect** means the percentage of physiological effects caused by a drug, measuring the efficacy of the drug, and its unit is %.
- **AC50** means 50% the maximum active concentration. The concentration at which a drug reaches 50% of its maximum effect under specific conditions. its unit is nM.
- **Ki** means inhibition constant, which is the concentration of the free inhibitor corresponding to the binding of 50% of the enzyme to the inhibitor. its unit is nM.
- **IC50** means 50% inhibitory concentration, which is the concentration of the inhibitor required to achieve a 50% inhibitory effect. its unit is nM.

In StructNet, an Assay ChEMBL ID corresponds to an assay. For example, CHEMBL1614458 represents a biological assay: qHTS Assay for Inhibitors of Aldehyde Dehydrogenase 1 (ALDH1A1). CHEMBL1614459 is qHTS Assay for Lipid Storage Modulators in Drosophila S3 Cells. CHEMBL1613914 is qHTS Assay for Inhibitors of Human Jumonji Domain Containing 2E (JMJD2E). You can easily find the description of all assays in StructNet by following the link https://www.ebi.ac.uk/chembl/assay_report_card/{using your Assay ChEMBL ID in here}/.

It is worth noting that when constructing StructNet from ChEMBL, considering that the isomerism phenomenon of drug molecules may significantly affect the measurement results of its biological activity, we only use SMILES as the merging condition of multiple experiments instead of canonical SMILES. We filtered through RDKit's isomericSmiles condition and find that all SMILES are isomers, so we retain the differences between SMILES. We also present the results of repeatability detection using canonical SMILES. As shown in Tables S7, S8, S9, S10, S11, S12, we find that in

most datasets, the duplication of canonical SMILES is not obvious. Notably, we perform an isomer check for SMILES using RDKit on all molecules where canonical SMILES was repeated. The results show that these SMILES repeated in canonical formats sequences are isomers.

Table S7: Statistics after deduplication using canonical SMILES in **reticular** molecules from StructNet, called R. #Mol represents the total number of molecules. #Unique represents the number of molecules after removing duplicates. Ratio represents $1 - \frac{\#\text{Unique}}{\#\text{Mol}} \times 100$.

Assay ChEMBL ID	#Mol / #Unique / #Ratio
CHEMBL1613914	302 / 300 / 0.7%
CHEMBL1614038	213 / 213 / 0%
CHEMBL1614087	259 / 257 / 0.8%
CHEMBL1614236	232 / 232 / 0%
CHEMBL1614249	247 / 246 / 0.4%
CHEMBL1614421	269 / 264 / 1.9%
CHEMBL1614458	489 / 486 / 0.6%
CHEMBL1614459	347 / 346 / 0.3%
CHEMBL1614544	218 / 215 / 1.4%
CHEMBL4888485	641 / 637 / 0.6%

Table S8: Statistics after deduplication using canonical SMILES in **acyclic** molecules from StructNet, called A. #Mol represents the total number of molecules. #Unique represents the number of molecules after removing duplicates. Ratio represents $1 - \frac{\#\text{Unique}}{\#\text{Mol}} \times 100$.

Assay ChEMBL ID	#Mol / #Unique / #Ratio
CHEMBL1614458	308 / 302 / 1.9%
CHEMBL4296187	274 / 274 / 0%
CHEMBL4296188	257 / 257 / 0%
CHEMBL4303805	276 / 273 / 1.1%
CHEMBL4495582	339 / 331 / 2.4%
CHEMBL4513082	339 / 331 / 2.4%
CHEMBL4649949	255 / 255 / 0%
CHEMBL4649955	255 / 255 / 0%
CHEMBL4808149	274 / 269 / 1.8%
CHEMBL4808150	273 / 268 / 1.8%

Table S9: Statistics after deduplication using canonical SMILES in **complete chain** molecules from StructNet, called CC.

Assay ChEMBL ID	#Mol / #Unique / #Ratio
CHEMBL1614458	862 / 855 / 0.8%
CHEMBL1614459	1065 / 1064 / 0.1%
CHEMBL1614530	676 / 676 / 0%
CHEMBL4296187	1257 / 1257 / 0%
CHEMBL4296188	1196 / 1196 / 0%
CHEMBL4296802	1134 / 1134 / 0%
CHEMBL4649948	1763 / 1762 / 0.1%
CHEMBL4649949	1763 / 1762 / 0.1%
CHEMBL4649955	1742 / 1741 / 0.1%
CHEMBL4888485	1670 / 1669 / 0.1%

Table S10: Statistics after deduplication using canonical SMILES in **acyclic chain** molecules from StructNet, called AC.

Assay ChEMBL ID	#Mol / #Unique / #Ratio
CHEMBL1614361	186 / 184 / 1.1%
CHEMBL1614458	243 / 240 / 1.2%
CHEMBL4296187	208 / 208 / 0%
CHEMBL4296188	193 / 193 / 0%
CHEMBL4303805	180 / 179 / 0.6%
CHEMBL4495582	231 / 228 / 1.3%
CHEMBL4513082	231 / 228 / 1.3%
CHEMBL4649948	179 / 179 / 0%
CHEMBL4649949	179 / 179 / 0%
CHEMBL4649955	179 / 179 / 0%

Table S11: Statistics after deduplication using canonical SMILES in **macrocyclic peptide** molecules from StructNet, called MP.

Assay ChEMBL ID	#Mol / #Unique / #Ratio
CHEMBL2354301	420 / 131 / 68.8%
CHEMBL3214979	216 / 87 / 59.7%
CHEMBL3880198	281 / 243 / 13.5%
CHEMBL4419595	251 / 221 / 12%
CHEMBL4419601	208 / 180 / 13.5%
CHEMBL4419606	207 / 181 / 12.6%
CHEMBL4420271	251 / 221 / 12%
CHEMBL4420277	208 / 180 / 13.5%
CHEMBL4420282	227 / 198 / 12.8%
CHEMBL4888485	480 / 480 / 0%

Table S12: Statistics after deduplication using canonical SMILES in **macro** molecules from StructNet, called M.

Assay ChEMBL ID	#Mol / #Unique / #Ratio
CHEMBL3881498	712 / 696 / 2.2%
CHEMBL3881499	686 / 666 / 2.9%
CHEMBL4419595	694 / 657 / 5.3%
CHEMBL4419597	523 / 507 / 3.1%
CHEMBL4419605	711 / 693 / 2.5%
CHEMBL4419606	1232 / 1187 / 3.7%
CHEMBL4420271	694 / 657 / 5.3%
CHEMBL4420273	523 / 507 / 3.1%
CHEMBL4420281	725 / 707 / 2.5%
CHEMBL4420282	1407 / 1358 / 3.5%

E EXPERIMENT SETTINGS

E.1 BASELINES

To comprehensively evaluate the performance of different modality-based methods, we selected a large number of common baselines for each modality. However, since BenchMol supports many baseline methods, it is difficult for us to cover all baseline methods. Therefore, we select methods based on whether they are representative and leave the remaining methods to the community for verification and exploration. Even so, we have selected a large number of baselines and the conclusions we draw are also statistically significant.

Table S13: The 6 common fingerprints for benchmarking evaluation.

No.	Baselines	Descriptions
1	mcfp4_2048	MCFP4 (Morgan Connectivity Fingerprints) with dimension 2048 and radius 2, which is a variant of ECFP4 based on the Morgan algorithm and is a classical approach for generating circular fingerprints that focus on capturing connectivity patterns in molecules
2	ecfp4_2048	ECFP4 (Extended Connectivity Fingerprints) with dimension 2048 and radius 2, which is a circular topological fingerprint that encodes information about a molecule's atomic connectivity and its local environment
3	maccs	166-dimensional MACCS (Molecular ACCess System) keys are a type of molecular fingerprint that is widely used in cheminformatics to represent molecular structures
4	physchem	physchem (physicochemical) fingerprints is based on molecular physicochemical properties
5	atompair_2048	2048-dimensional AtomPair fingerprints, which represents molecules based on pairs of atoms and the shortest path (in bonds) between them, encoding both the atom types and their relative positions within a molecule
6	rdkDes	rdkDsc (RDKit Descriptors) are a set of predefined molecular descriptors calculated using the RDKit cheminformatics toolkit

Table S14: The sequence-based baselines for benchmarking evaluation.

No.	Baselines	Descriptions
1	BERT-6L	6-layer BERT model, which comes from Chem-BERT with random initialization
2	BERT-8L/BERT-8L-R	8-layer BERT model, which comes from Chem-BERT with random initialization
3	RoBERTa-12L	12-layer RoBERTa model, which comes from Chem-RoBERTa with random initialization
4	molformer-R/MolFormer-R	MolFormer with random initialization
5	Chem-BERT-6L	6-layer Chem-BERT
6	Chem-BERT-8L	8-layer Chem-BERT
7	CHEM-RoBERTa-12L	12-layer CHEM-RoBERTa
8	Molformer/MolFormer	MolFormer

We describe the selected baselines for different modalities below:

- **Fingerprint.** Table S13 shows 6 commonly used molecular fingerprinting methods.
- **Sequence.** The baselines here use molecular sequences as input. As shown in Table S14, we use 8 common sequence-based models for benchmarking evaluation.
- **Graph.** The baselines here use molecular graphs as input. The Table S15 shows 9 graph-based baselines for benchmarking evaluation.
- **Geometry Graph.** The baselines here use molecular geometry graph as input. In the paper, the word "geometry" alone refers to the geometry graph. The Table S16 shows 9 geometry-based baselines for benchmarking evaluation.
- **Image.** The baselines here use molecular images as input. The Table S17 shows 5 image-based baselines for benchmarking evaluation.
- **Geometry Image.** The baselines here use molecular geometry images as input. The Table S18 shows 6 geometry image-based baselines for benchmarking evaluation.
- **Video.** The baselines here use molecular videos as input. The Table S19 shows 4 video-based baselines for benchmarking evaluation.

E.2 Hyperparameters Search

In molecular property prediction, we use a linear probing strategy to train all models. To be fair, the hyperparameter search ranges for all models are the same. We set the batch size to 32 and perform grid search on learning rates of 0.001, 0.005, 0.01, and 0.05. In the MBANet task, all models use the same hyperparameters (batch size of 8 and learning rate of 0.005) to train the models. In StructNet, we set the batch size to 8 and perform grid search on learning rates of 0.001, 0.005.

To eliminate the influence of randomness, we used a uniform set of 10 random seeds ranging from 0 to 9 to calculate the mean and variance in all experiments in this paper.

Table S15: The 9 graph-based baselines for benchmarking evaluation.

No.	Baselines	Descriptions
1	GIN-R	GIN model with random initialization
2	EdgePred	-
3	ContextPred	-
4	infomax	-
5	masking	-
6	MolCLR	-
7	MoleBERT	-
8	GraphMVP	-
9	CGIP-Graph	-

Table S16: The 9 geometry-based baselines for benchmarking evaluation.

No.	Baselines	Descriptions
1	Uni-Mol-R (1 conf)	Uni-Mol with random initialization, which uses 1 conformation during fine-tuning
2	Uni-Mol (1 conf)	pre-trained Uni-Mol, which uses 1 conformation during fine-tuning
3	Uni-Mol-R (10 conf)	Uni-Mol with random initialization, which uses 10 conformations during fine-tuning
4	Uni-Mol (10 conf)	pre-trained Uni-Mol, which uses 10 conformations during fine-tuning
5	SchNet	-
6	EGNN	-
7	TFN	-
8	SE3_Transformer	-
9	PaiNN	-

E.3 Training Loss

In MoleculeNet benchmark, there are 8 classification tasks and 4 regression tasks. For classification tasks, we use cross entropy loss. For regression tasks, we use MSE loss. In MBANet and StructNet benchmarks, we use MSE loss because they are regression tasks.

F More Results on MoleculeNet

F.1 Detailed Results

Table S20 and Table S21 show the ROC-AUC performance on 8 classification tasks and RMSE performance on 4 regression tasks from MoleculeNet, respectively. In classification tasks, as shown in Table S20, we find that fingerprint-based atompair_2048 achieves the best performance on Sider and BACE, which shows that fingerprinting is a simple and effective method for property prediction. The sequence-based MolFormer-R achieves state-of-the-art performance on BBBP and ClinTox without any pre-training, which indicates that sequence have a strong inductive bias for molecular property prediction. The graph-based MoleBERT, geometry-based Uni-Mol (10 conf), and geometry image-based IEM-G (10 conf) achieved the best performance on Tox21, ToxCast and MUV, HIV, respectively. These findings suggest that different modalities have certain preferences for different tasks, which can further establish guiding ideas for multi-modal learning of molecules.

In addition, in Table S21, we find the superiority of fingerprints on the regression task of property prediction, which achieves the best performance on 3 (Lipo, Malaria and CEP) out of 4 datasets.

F.2 Results of Geometry Image and Video with RGB Format

Images are available in RGB and BGR formats, and it is meaningful to study the difference between RGB and BGR for images. Here, we report the results using RGB format images as input. Table S22 and Table S23 show the results of 6 vision-based methods on 8 classification tasks and 4 regression tasks from MoleculeNet using RGB-format images. We find that using RGB or BGR as the input of the visual modality has little impact on performance.

Table S17: The 5 image-based baselines for benchmarking evaluation.

No.	Baselines	Descriptions
1	ResNet18-R	image-based ResNet18 without pre-training
2	ImageMol	
3	MaskMol	
4	CGIP-Image	-
5	IEM -I	

Table S18: The 6 geometry image-based baselines for benchmarking evaluation.

No.	Baselines	Descriptions
1	ResNet18-G-R	geometry image-based ResNet18 without pre-training
2	IEM-G	IEM using geometry images as input
3	ResNet18-G-R (10 conf)	geometry image-based ResNet18 without pre-training, which uses 10 conformations during fine-tuning
4	IEM-G (10 conf)	IEM using geometry images as input, which uses 10 conformations during fine-tuning
5	ViT-G-R	ViT without pre-training, which is the bachbone of VideoMol and uses geometry image as input
6	VideoMol-G	pre-trained VideoMol, which uses geometry image as input

F.3 Results of Fine-tuning

Fine-tuning is a common strategy to maximize the performance of pre-trained models on downstream tasks. To compare the performance difference between linear probing and fine-tuning, we fine-tune Uni-Mol, Molformer, ImageMol and VideoMol using their public source code. Specifically, we use the officially released no-H pre-trained weight and the corresponding optimal hyperparameters to fine-tune Uni-Mol [1].

The Table S24 shows the fine-tuning performance of Uni-Mol, Molformer, ImageMol, and VideoMol on 6 classification tasks (Tox21, ToxCast, Sider, HIV, BBBP and BACE) from MoleculeNet. Overall, except for Molformer, fine-tuning on other methods helps improve the performance compared to linear probing. In particular, we find that after fine-tuning, the image modality improves performance by 14.98% compared to linear probing. The significant performance improvement indicates that the image modality currently relies on detailed fine-tuning to further improve performance.

G More Results on MBANet

G.1 Atom

Table S25 shows the average RMSE performance of a large number of baselines on MBANet$_{atom}$ for 12 atom distribution prediction tasks (C, N, O, F, S, Cl, Br, P, Si, B, Se and Ge) with 10 seeds. Table S26 shows the corresponding standard deviation. We find that IEM-V based on video modality achieves the best performance on half of the atom prediction tasks, which shows the advantages of video-based modality. At the same time, we also find that models based on images and geometry images also achieve good performance, such as CGIP-Image and IEM-G (1 conf), indicating that the model can accurately count the number of atoms from the image. Furthermore, we observe that sequence- and graph-based models perform poorly on MBANet$_{atom}$, indicating that the global representations extracted by sequence- and graph-based models are not conducive to atomic-level prediction tasks.

G.2 Bond

Table S27 shows the average RMSE performance of a large number of baselines on MBANet$_{atom}$ for 4 bond distribution prediction tasks (single bond, aromatic bond, double bond, and triple bond) with 10 seeds. We find that the geometry-based TFN model has strong predictive power at the bond level, which suggests that utilizing both molecular geometry and bond information can effectively improve the model's understanding of bond distribution compared to graph models that only use

[1] https://github.com/deepmodeling/Uni-Mol/tree/main/unimol#molecular-property-prediction

Table S19: The 4 video-based baselines for benchmarking evaluation.

No.	Baselines	Descriptions
1	ResNet18-V-R	video-based ResNet18 without pre-training
2	IEM-V	IEM using video as input
3	VideoMol-R	VideoMol without pre-training
4	VideoMol	pre-trained VideoMol, which uses video as input

Table S20: The ROC-AUC (%) performance on 8 classification tasks with linear probing using BenchMol. The modality types from top to bottom are fingerprint, sequence, graph, geometry graph, image, geometry image, and video. -R means no pre-training, L means the number of layers, -I, -G, and -V mean the modalities are image, geometry image, and video, respectively. Note that the geometry images and videos use the BGR format.

Model	BBBP	Tox21	ToxCast	Sider	ClinTox	MUV	HIV	BACE	Avg
mcfp4_2048	64.9±0.3	73.5±0.1	61.9±0.0	65.0±0.1	83.0±0.2	73.4±0.3	73.8±0.3	77.8±1.0	71.66
ecfp4_2048	63.9±0.1	71.2±0.1	59.7±0.1	64.2±0.0	76.9±0.1	70.1±0.1	72.0±0.1	80.5±0.6	69.81
maccs	66.6±0.3	70.1±0.0	61.7±0.0	63.8±0.1	85.1±0.2	71.9±0.5	70.7±1.5	74.4±1.2	70.54
physchem	61.8±0.1	62.6±0.1	59.4±0.1	57.7±0.9	66.4±5.7	69.6±0.1	66.7±0.0	61.1±2.5	63.16
atompair_2048	65.5±0.3	73.1±0.3	64.8±0.1	**66.2±0.0**	63.9±0.4	70.4±0.2	74.8±0.2	**83.5±1.3**	70.28
rdkDes	59.3±0.5	64.1±0.2	60.5±0.2	54.0±0.7	56.2±0.0	66.1±1.1	68.8±0.0	76.5±0.0	63.19
BERT-8L-R	68.7±0.9	72.0±0.3	62.1±0.2	56.6±0.5	82.6±1.4	64.0±1.8	71.1±1.6	74.4±1.5	68.94
Chem-BERT-8L	69.2±0.2	75.5±0.1	62.6±0.1	62.6±0.1	83.3±0.6	77.6±1.2	78.2±0.2	78.3±0.3	73.41
MolFormer-R	**74.6±0.5**	71.6±0.3	61.5±0.3	55.9±0.3	**86.2±0.3**	67.2±1.6	71.2±0.5	75.0±1.6	70.40
MolFormer	63.3±0.2	72.1±0.1	61.4±0.1	63.4±0.2	68.2±3.3	75.4±0.8	74.5±0.4	78.3±1.8	69.58
GIN-R	57.3±0.3	69.3±0.3	58.3±0.4	56.2±0.8	64.9±0.4	69.6±1.1	66.9±1.2	63.2±0.3	63.21
EdgePred	52.1±1.0	67.1±0.3	56.4±0.0	54.5±0.6	55.0±2.7	65.8±0.2	67.6±0.5	67.3±2.8	60.73
ContextPred	57.6±0.3	70.6±0.1	60.7±0.0	60.8±1.0	58.6±3.2	76.4±0.3	72.4±1.3	78.1±0.2	66.90
infomax	62.4±0.0	68.6±0.2	59.2±0.1	58.5±0.7	60.1±2.2	76.4±0.3	71.9±0.3	73.7±1.0	66.35
masking	57.9±0.4	68.9±0.2	58.2±0.0	58.8±0.2	52.2±0.4	70.7±1.4	65.5±0.5	68.0±0.2	62.53
MolCLR	63.5±0.1	69.0±0.0	61.4±0.1	58.6±0.3	64.2±1.0	65.0±1.4	72.1±0.3	70.2±0.2	65.50
MoleBERT	66.3±0.1	**77.1±0.1**	65.0±0.1	63.9±0.1	74.8±3.4	79.7±0.2	76.2±0.1	75.2±0.9	72.28
GraphMVP	64.2±0.4	69.5±0.1	60.6±0.1	58.6±0.1	56.7±1.5	68.6±0.4	71.6±0.4	76.4±0.1	65.78
CGIP-Graph	66.4±0.9	71.5±0.3	58.6±0.0	57.5±0.1	70.3±0.2	72.5±3.4	73.6±0.8	66.0±3.7	67.05
Uni-Mol-R	66.3±1.0	67.6±0.1	62.4±0.1	59.2±0.1	59.2±0.4	59.5±1.6	74.7±0.2	77.5±0.7	65.80
Uni-Mol	69.8±0.1	74.5±0.1	65.6±0.4	60.2±0.4	84.3±0.4	78.5±0.2	78.4±0.1	78.7±0.5	73.75
Uni-Mol-R (10 conf)	64.6±0.4	67.7±0.1	63.1±0.1	59.3±1.2	58.6±0.2	56.5±1.8	77.2±0.1	77.2±0.1	65.53
Uni-Mol (10 conf)	69.3±0.6	75.2±0.1	**65.8±0.5**	61.6±0.4	85.1±4.4	**80.3±0.7**	77.5±0.1	78.2±0.2	**74.13**
ResNet18-R	52.4±0.1	53.8±0.1	54.7±0.0	55.8±0.1	65.3±0.2	49.8±1.7	52.8±0.2	58.1±2.2	55.34
ImageMol	60.5±0.5	66.4±0.3	59.0±0.3	58.2±0.2	64.5±2.6	61.3±1.3	70.8±0.9	60.3±1.0	62.63
MaskMol	62.3±1.1	65.9±0.1	60.1±0.1	59.1±0.7	56.4±0.7	58.8±3.2	74.4±0.6	67.2±1.2	63.03
CGIP-Image	56.2±0.3	66.0±0.0	55.6±0.0	57.2±0.3	68.0±0.3	63.1±0.0	69.5±0.1	59.9±0.3	61.94
IEM -I	59.7±0.1	65.8±0.2	57.1±0.1	56.8±0.9	57.8±3.6	56.8±1.0	72.2±0.5	61.4±0.6	60.95
ResNet18-G-R	55.2±0.0	58.2±0.2	57.4±0.2	55.7±0.1	60.0±0.4	54.3±3.7	71.5±0.4	57.5±1.9	58.73
IEM-G	64.1±0.2	69.0±0.3	60.9±0.2	56.0±1.0	55.8±0.8	60.8±2.0	75.5±0.7	75.2±0.6	64.66
ResNet18-G-R (10 conf)	60.1±0.0	62.3±0.1	59.8±0.1	57.1±0.4	67.5±0.1	57.0±0.3	74.3±0.0	71.3±1.3	63.68
IEM-G (10 conf)	68.1±0.4	71.9±0.2	63.9±0.1	61.5±0.8	68.6±1.0	68.7±1.1	**80.0±0.4**	79.6±1.2	70.29
VideoMol-R	59.7±0.2	61.3±0.1	59.2±0.4	55.2±1.4	60.0±0.1	53.1±1.5	75.4±0.4	65.4±2.1	61.16
VideoMol	63.4±0.7	73.5±0.1	63.9±0.2	60.3±0.3	63.8±2.9	75.3±0.9	76.8±1.2	75.2±0.4	69.03

bond information. In addition, compared with SE3-Transformer and Uni-Mol, TFN is more suitable for capturing local bond information.

G.3 Basic Attributes

The Table S28 shows the average RMSE performance of a large number of baselines on MBANet$_{attr}$ for 8 basic attribute prediction tasks (MW, LogP, MR, BalabanJ, #HA, #HD, #VE, TPSA) with 10 seeds. The Table S29 shows the corresponding standard deviation. We find that graph modality achieves the best performance on LogP, BalabanJ, HA and HD and video modality achieves the best performance on MW, MR, VE and TPSA. In particular, IEM-V equipped with video modality achieves the best average performance, which is 54.2% higher than IEM-G using geometry image modality (from 4.916 to 2.254), indicating the superiority of combining IEM with video modality.

Table S21: The RMSE performance on 4 regression tasks with linear probing using BenchMol. Note that the geometry images and videos use the BGR format.

Model	ESOL	Lipo	Malaria	CEP	Avg
mcfp4_2048	1.510±0.017	0.833±0.001	1.096±0.001	**1.582±0.001**	1.255
ecfp4_2048	1.658±0.019	0.902±0.001	1.088±0.001	1.551±0.000	1.300
maccs	1.339±0.014	0.980±0.002	1.128±0.001	1.759±0.002	1.302
physchem	1.713±0.020	0.994±0.003	1.155±0.001	1.953±0.001	1.454
atompair_2048	1.220±0.006	**0.817±0.002**	**1.087±0.000**	1.632±0.003	1.189
rdkDes	1.830±0.001	1.067±0.003	1.166±0.002	2.529±0.001	1.648
BERT-8L-R	1.102±0.041	1.005±0.005	1.161±0.005	1.786±0.008	1.264
Chem-BERT-8L	**0.858±0.013**	0.823±0.003	1.106±0.002	1.584±0.006	**1.093**
MolFormer-R	1.278±0.007	0.994±0.003	1.157±0.005	1.845±0.001	1.319
MolFormer	1.350±0.016	0.936±0.004	1.123±0.004	1.764±0.006	1.293
GIN-R	1.780±0.013	1.078±0.004	1.148±0.002	2.299±0.004	1.576
EdgePred	2.396±0.019	1.075±0.001	1.134±0.002	2.053±0.006	1.665
ContextPred	1.520±0.010	1.031±0.006	1.129±0.003	2.165±0.007	1.461
infomax	1.450±0.012	1.035±0.007	1.131±0.004	2.018±0.012	1.409
masking	1.696±0.011	1.065±0.002	1.130±0.006	2.070±0.004	1.490
MolCLR	1.506±0.013	0.931±0.006	1.114±0.003	1.925±0.005	1.369
MoleBERT	1.544±0.006	0.897±0.001	1.105±0.001	1.735±0.004	1.320
GraphMVP	1.623±0.008	0.959±0.012	1.143±0.002	1.879±0.009	1.401
CGIP-Graph	2.494±0.020	0.903±0.006	1.113±0.004	1.696±0.009	1.552
Uni-Mol-R	1.048±0.019	0.999±0.004	1.146±0.002	2.038±0.010	1.308
Uni-Mol	1.003±0.005	0.856±0.004	1.113±0.001	1.676±0.004	1.162
Uni-Mol-R (10 conf)	0.997±0.017	0.984±0.008	1.149±0.002	1.974±0.006	1.276
Uni-Mol (10 conf)	0.978±0.005	0.839±0.004	1.109±0.002	1.648±0.008	1.144
ResNet18-R	1.917±0.004	1.108±0.002	1.166±0.001	2.535±0.000	1.682
ImageMol	1.655±0.021	1.053±0.008	1.150±0.008	2.169±0.003	1.507
MaskMol	1.329±0.034	1.056±0.005	1.160±0.004	2.219±0.006	1.441
CGIP-Image	1.710±0.023	1.078±0.001	1.149±0.001	2.287±0.001	1.556
IEM -I	1.730±0.028	1.057±0.005	1.156±0.001	2.364±0.002	1.577
ResNet18-G-R	1.561±0.015	1.073±0.002	1.164±0.001	2.472±0.001	1.568
IEM-G	1.313±0.008	0.974±0.003	1.155±0.001	2.180±0.001	1.406
ResNet18-G-R (10 conf)	1.359±0.015	1.055±0.003	1.161±0.002	2.364±0.001	1.485
IEM-G (10 conf)	0.936±0.023	0.887±0.006	1.155±0.001	1.868±0.007	1.212
VideoMol-R	1.520±0.017	1.058±0.006	1.163±0.001	2.364±0.003	1.526
VideoMol	1.085±0.011	0.887±0.003	1.137±0.004	1.780±0.003	1.222

Table S22: The ROC-AUC (%) performance on 8 classification tasks from MoleculeNet with RGB format with linear probing using BenchMol. The first 4 are geometry image-based methods, and the last 2 are video-based methods.

Model	BBBP	Tox21	ToxCast	Sider	ClinTox	MUV	HIV	BACE	Avg
ResNet18-G-R	56.2±0.2	59.9±0.1	57.1±0.1	53.8±0.1	59.2±0.5	57.9±0.8	72.4±0.8	63.3±1.4	59.98
IEM-G	64.5±0.6	69.5±0.4	61.3±0.3	56.1±1.0	49.7±0.7	61.8±0.7	75.2±0.8	76.5±0.2	64.33
ResNet18-G-R (10 conf)	58.7±0.0	62.8±0.1	58.5±0.1	56.3±0.1	**68.0±0.1**	57.3±0.6	74.8±0.6	69.2±0.6	63.20
IEM-G (10 conf)	65.7±1.3	73.2±0.2	**63.8±0.2**	59.8±0.2	58.3±0.2	68.8±0.7	**78.5±0.0**	**80.3±0.1**	**68.55**
VideoMol-R	60.6±0.3	60.6±0.1	58.8±0.4	57.2±0.2	59.1±0.0	53.9±1.7	76.7±0.4	61.0±1.9	60.99
VideoMol	**66.5±0.2**	**73.7±0.2**	63.2±0.2	**61.8±0.2**	57.2±2.8	**74.0±0.5**	75.2±0.5	76.6±0.8	68.53

H EXPANDING MBANET TO 30,000 MOLECULES

In order to study the generalization of MBANet's conclusions, we further expand MBANet to 30,000 molecules, referred to as MBANet^{30K}. As shown in Table S30, we find that the results of MBANet^{30K} are not significantly different from the results on the original MBANet (see Table 5). For example, the video modality still achieves the best performance in tasks related to atoms and basic attributes. Therefore, the conclusion of MBANet is effective after further expanding the scale of MBANet to 30K.

Table S23: The RMSE performance on 4 regression tasks with RGB format with linear probing using BenchMol.

Model	ESOL	Lipo	Malaria	CEP	Avg
ResNet18-G-R	1.715±0.007	1.080±0.002	1.163±0.001	2.469±0.000	1.607
IEM-G	1.194±0.007	0.981±0.001	1.170±0.001	2.130±0.002	1.369
ResNet18-G-R (10 conf)	1.395±0.013	1.048±0.005	1.163±0.001	2.392±0.002	1.500
IEM-G (10 conf)	**0.968±0.017**	0.893±0.004	1.132±0.005	1.874±0.004	1.217
VideoMol-R	1.515±0.025	1.050±0.009	1.165±0.001	2.337±0.003	1.517
VideoMol	1.035±0.005	**0.867±0.005**	**1.099±0.004**	**1.760±0.006**	**1.190**

Table S24: The ROC-AUC (%) performance on 6 classification tasks from MoleculeNet with fine-tuning setting under 10 random seeds ranging from 0 to 9. We fine-tune these models using their public code. Specifically, we evaluate Uni-Mol using the officially released no-H pre-trained weight and the corresponding optimal hyperparameters. FT denotes the average performance of fine-tuning on 6 datasets. LP denotes the average result of linear probing, which is obtained by Table S20. δ denotes $(\frac{FT}{LP} - 1) * 100\%$. Note that IEM and VideoMol use the BGR format.

	Tox21	ToxCast	Sider	HIV	BBBP	BACE	FT	LP	δ
Uni-Mol (1 conf)	78.3 (0.4)	68.7 (0.5)	63.7 (1.3)	79.2 (1.0)	69.6 (2.0)	81.0 (3.9)	73.4	71.2	↑ 3.11%
Uni-Mol (10 conf)	78.8 (0.7)	69.0 (0.5)	63.6 (1.4)	79.2 (0.9)	69.9 (2.7)	81.7 (3.4)	73.7	71.3	↑ 3.41%
Molformer	47.4 (2.1)	56.2 (1.5)	61.1 (1.0)	74.6 (0.9)	69.5 (1.0)	80.9 (1.9)	65.0	**69.6**	↓ 6.65%
ImageMol	75.5 (1.0)	65.6 (0.9)	64.9 (1.3)	76.8 (1.3)	70.5 (1.3)	78.1 (3.5)	71.9	62.5	↑ **14.98%**
VideoMol	**78.8 (0.5)**	**66.7 (0.5)**	**66.3 (0.9)**	**79.4 (0.5)**	**70.7 (2.2)**	**82.4 (0.9)**	**74.1**	69.0	↑ 7.27%

I MORE RESULTS ON STRUCTNET

I.1 ACYCLIC CHAIN MOLECULES

Table S31 and Table S32 show the average RMSE performance and corresponding standard deviation of the baselines on 10 acyclic chain datasets, respectively. Overall, each modality has baselines that make it into the top 5 in terms of performance. From the average performance, we find the effectiveness of graph pre-training strategies on acyclic chain molecules because 4 (MoleBERT, infomax, MolCLR and masking) of the top 5 methods are graph-based pre-training methods. We also find that even without pre-training, SchNet can still achieve good performance on acyclic chain molecules, ranking second, which demonstrates the effectiveness of geometric methods on this type of molecules. Furthermore, we find that the non-pre-trained vision-based ResNet18-G-R and ResNet18-V-R also achieve top-5 performance, which indicates the effectiveness of these visual representations.

I.2 ACYCLIC MOLECULES

Table S33 and Table S34 show the average RMSE performance and corresponding standard deviation of the baselines on 10 acyclic datasets, respectively. Here, we find the effectiveness of graph pre-training methods because the top 5 methods on performance are all based on graph pre-training methods, such as MoleBERT, ContextPred, masking, CGIP-Graph and infomax. When no pre-training is performed, the geometry-based TFN model achieves the best performance, demonstrating the advantage of geometric methods on acyclic molecules.

I.3 COMPLETE CHAIN MOLECULES

Table S35 and Table S36 show the average RMSE performance and corresponding standard deviation of the baselines on 10 complete chain datasets, respectively. We find that molecular fingerprint-based maccs achieves the best average performance, surpassing a number of pre-training methods, demonstrating the advantages of maccs on complete chain molecules.

Notably, we observe that a large number of graph-based and image-based pre-training strategies fail on this type of molecules. Specifically, 6 out of 8 graph-based pre-training methods (EdgePred, ContextPred, infomax, masking, MoleBERT and CGIP-Graph) and all 4 image-based pre-training

Table S25: The average RMSE performance on MBANet$_{atom}$ with 10 seeds. The modality types from top to bottom are sequence, graph, geometry graph, image, geometry image, and video. -R means no pre-training, L means the number of layers, -I, -G, and -V mean the modalities are image, geometry image, and video, respectively. The green background represents top-5 performance.

Models	#C	#N	#O	#F	#S	#Cl	#Br	#P	#Si	#B	#Se	#Ge	Mean
BERT-6L	2.001	0.810	1.460	0.742	0.322	0.422	0.206	**0.014**	0.023	0.237	0.012	0.009	0.522
BERT-8L	2.011	0.807	1.479	0.727	0.321	0.420	0.210	0.027	0.022	0.238	0.014	0.015	0.524
RoBERTa-12L	2.309	0.806	1.463	0.722	0.319	0.426	0.196	0.022	0.035	0.233	0.010	0.012	0.546
molformer-R	2.288	0.819	1.514	0.702	0.321	0.423	0.204	0.025	0.036	0.241	0.019	0.021	0.551
Chem-BERT-6L	1.985	0.801	1.453	0.730	0.320	0.425	0.205	0.019	0.031	0.247	0.013	0.026	0.521
Chem-BERT-8L	2.002	0.805	1.491	0.722	0.323	0.422	0.197	0.024	0.022	0.239	0.023	0.013	0.524
CHEM-RoBERTa-12L	2.186	0.807	1.468	0.708	0.319	0.430	0.200	0.017	0.024	0.234	0.010	0.007	0.534
Molformer	2.121	0.798	1.448	0.702	0.322	0.423	0.201	0.022	0.034	0.238	0.035	0.023	0.531
GIN-R	1.445	0.520	0.376	0.392	0.459	0.366	0.195	0.035	0.042	0.234	0.009	0.002	0.340
EdgePred	1.396	0.586	0.354	0.442	0.197	0.300	0.189	0.036	0.045	0.234	0.011	0.002	0.316
ContextPred	1.353	0.532	0.366	0.406	0.283	0.315	0.193	0.058	0.031	0.234	0.031	0.006	0.317
infomax	1.427	0.593	0.372	0.418	0.211	0.296	0.188	0.036	0.027	0.235	0.012	0.002	0.318
masking	1.481	0.529	0.412	0.406	0.337	0.305	0.184	0.034	0.035	0.233	0.024	0.004	0.332
MolCLR	1.375	0.533	0.395	0.344	0.395	0.355	0.187	0.034	0.037	0.233	0.012	**0.001**	0.325
MoleBERT	1.552	0.487	0.378	0.380	0.197	0.260	0.191	0.045	0.044	0.233	0.019	0.016	0.317
CGIP-Graph	1.165	0.310	0.318	0.207	0.115	0.127	0.190	0.041	0.037	0.233	0.017	0.008	0.231
GraphMVP	1.324	0.519	0.390	0.428	0.452	0.325	0.192	0.030	0.031	0.236	0.008	0.004	0.328
Uni-Mol-R (1 conf)	1.436	0.713	0.741	0.352	0.143	0.161	0.168	0.061	0.044	0.235	0.015	0.002	0.339
Uni-Mol (1 conf)	1.466	0.784	0.756	0.362	0.144	0.156	0.156	0.043	0.029	0.225	0.020	0.005	0.346
TFN	0.602	0.337	0.290	0.207	0.117	0.149	**0.139**	0.047	0.037	0.176	0.015	0.010	0.177
SE3_Transformer	1.773	0.849	1.475	0.720	0.312	0.410	0.195	0.019	**0.020**	0.234	0.010	0.015	0.503
ResNet18-I-R	1.332	0.608	0.747	0.473	0.210	0.321	0.194	0.047	0.029	0.234	0.007	0.004	0.350
ImageMol	1.340	0.578	0.750	0.485	0.225	0.321	0.194	0.054	0.035	0.232	0.007	0.002	0.352
CGIP-Image	1.251	0.581	0.713	0.462	0.224	0.305	0.196	0.040	0.025	0.234	**0.005**	0.005	0.337
MaskMol	2.096	0.818	1.466	0.706	0.325	0.432	0.198	0.034	0.028	0.242	0.016	0.017	0.532
IEM-I	1.324	0.580	0.738	0.489	0.210	0.326	0.195	0.043	0.025	0.233	0.009	0.003	0.348
ResNet18-G-R	0.939	0.349	0.467	0.409	0.165	0.343	0.198	0.045	0.056	0.223	0.012	0.013	0.268
IEM-G (1 conf)	0.803	0.327	0.452	0.338	0.152	0.313	0.196	0.050	0.046	0.232	0.009	0.003	0.243
ResNet18-V-R	0.520	0.154	0.215	0.225	0.087	0.191	0.156	0.043	0.140	**0.129**	0.008	**0.001**	0.156
IEM-V	**0.354**	**0.130**	**0.186**	**0.148**	**0.074**	**0.119**	0.164	0.021	0.058	0.220	0.007	0.002	**0.124**

Table S26: The standard deviation on MBANet$_{atom}$ with 10 seeds. The modality types from top to bottom are sequence, graph, geometry graph, image, geometry image, and video. -R means no pre-training, L means the number of layers, -I, -G, and -V mean the modalities are image, geometry image, and video, respectively.

Models	#C	#N	#O	#F	#S	#Cl	#Br	#P	#Si	#B	#Se	#Ge
BERT-6L	0.216	0.035	0.082	0.058	0.007	0.015	0.026	**0.010**	0.012	0.009	0.011	0.006
BERT-8L	0.218	0.028	0.079	0.043	0.013	**0.007**	0.027	0.022	0.014	0.008	0.008	0.010
RoBERTa-12L	0.134	0.017	0.079	0.024	0.009	0.013	0.004	0.021	0.025	0.004	0.010	0.013
molformer-R	0.187	0.064	0.109	0.023	0.007	0.014	0.010	0.024	0.032	0.013	0.011	0.021
Chem-BERT-6L	0.213	**0.012**	0.079	0.051	0.008	0.017	0.017	0.014	0.024	0.014	0.008	0.017
Chem-BERT-8L	0.177	0.028	0.070	0.038	0.009	0.013	0.004	0.022	0.021	0.007	0.026	0.012
CHEM-RoBERTa-12L	0.188	0.018	0.091	0.028	0.011	0.018	0.004	0.016	0.022	0.004	0.012	0.006
Molformer	0.162	**0.012**	0.093	**0.016**	0.010	0.015	0.006	0.025	0.018	0.011	0.037	0.020
GIN-R	0.313	0.129	0.076	0.054	0.095	0.027	0.015	0.027	0.024	0.004	0.013	0.003
EdgePred	0.254	0.154	0.089	0.094	0.036	0.098	0.004	0.015	0.026	0.004	0.010	0.002
ContextPred	0.211	0.167	0.077	0.128	0.121	0.073	0.010	0.030	0.017	0.004	0.029	0.008
infomax	0.172	0.088	0.102	0.187	0.065	0.097	0.004	0.031	0.018	0.004	0.011	0.002
masking	0.362	0.167	0.099	0.139	0.139	0.078	0.006	0.022	0.023	0.004	0.018	0.006
MolCLR	0.196	0.143	0.076	0.085	0.079	0.032	0.006	0.015	0.014	0.004	0.014	**0.001**
MoleBERT	0.389	0.120	0.105	0.085	0.066	0.104	0.006	0.016	0.024	**0.003**	0.014	0.025
CGIP-Graph	0.175	0.055	0.046	0.040	0.035	0.036	0.008	0.020	0.016	0.009	0.010	0.005
GraphMVP	0.248	0.144	0.103	0.146	0.078	0.021	0.008	0.021	0.018	0.005	0.009	0.006
Uni-Mol-R (1 conf)	0.133	0.086	0.057	0.025	0.031	0.030	0.018	0.021	0.029	0.013	0.005	0.002
Uni-Mol (1 conf)	0.123	0.081	0.045	0.042	0.035	0.042	0.029	0.011	0.013	0.009	0.025	0.005
TFN	0.092	0.069	0.038	0.025	0.025	0.039	0.053	0.014	**0.008**	0.040	0.016	0.011
SE3_Transformer	0.179	0.081	0.231	0.033	**0.006**	0.037	0.003	0.018	0.025	0.006	0.016	0.030
ResNet18-I-R	0.095	0.040	0.062	0.037	0.013	0.034	**0.002**	0.031	0.015	0.004	**0.003**	0.006
ImageMol	0.116	0.037	0.052	0.029	0.026	0.046	0.003	0.027	0.017	**0.003**	0.005	0.002
CGIP-Image	0.095	0.027	0.044	0.028	0.028	0.027	**0.002**	0.013	0.012	**0.003**	**0.003**	0.007
MaskMol	0.136	0.023	0.115	0.017	0.010	0.015	0.004	0.021	0.022	0.010	0.013	0.013
IEM-I	0.072	0.032	0.038	0.037	0.013	0.038	0.003	0.019	0.012	0.005	0.008	0.004
ResNet18-G-R	0.171	0.030	0.067	0.069	0.045	0.173	0.005	0.016	0.024	0.011	0.009	0.030
IEM-G (1 conf)	0.063	0.025	0.043	0.084	0.036	0.069	0.006	0.033	0.026	0.006	0.005	0.004
ResNet18-V-R	0.179	0.023	**0.032**	0.048	0.014	0.044	0.041	0.031	0.036	0.030	0.006	**0.001**
IEM-V	**0.062**	0.041	0.038	0.052	0.020	0.042	0.047	0.015	0.065	0.022	0.005	0.003

Table S27: The average RMSE (standard deviation) performance on MBANet$_{bond}$ with 10 seeds. The modality types from top to bottom are sequence, graph, geometry graph, image, geometry image, and video. -R means no pre-training, L means the number of layers, -I, -G, and -V mean the modalities are image, geometry image, and video, respectively. The green background represents top-5 performance.

	Models	#SINGLE	#AROMATIC	#DOUBLE	#TRIPLE	Mean
SMILES	BERT-6L	5.092±0.271	3.626±0.176	1.240±0.113	0.606±0.047	2.641
	BERT-8L	5.254±0.275	3.543±0.218	1.180±0.107	0.616±0.044	2.648
	RoBERTa-12L	5.520±0.313	3.652±0.190	1.210±0.142	0.628±0.075	2.753
	molformer-R	5.623±0.247	3.624±0.302	1.315±0.114	0.592±0.049	2.788
	Chem-BERT-6L	5.211±0.251	3.602±0.163	1.271±0.105	0.582±0.024	2.666
	Chem-BERT-8L	4.937±0.686	3.758±0.436	1.237±0.159	0.597±0.038	2.632
	CHEM-RoBERTa-12L	5.548±0.246	3.645±0.197	1.289±0.230	0.621±0.041	2.776
	Molformer	5.367±0.266	3.621±0.231	1.215±0.059	0.611±0.032	2.703
Graph	GIN-R	0.988±0.099	1.002±0.124	0.273±0.060	0.145±0.032	0.602
	EdgePred	1.005±0.094	0.970±0.135	0.256±0.096	0.157±0.054	0.597
	ContextPred	1.001±0.236	0.965±0.161	**0.190±0.044**	0.155±0.045	0.578
	infomax	0.999±0.236	0.980±0.110	0.219±0.098	0.171±0.055	0.592
	masking	0.994±0.158	0.967±0.103	0.233±0.065	0.178±0.064	0.593
	MolCLR	1.072±0.136	1.004±0.137	0.242±0.059	0.206±0.113	0.631
	MoleBERT	1.039±0.210	0.910±0.133	0.196±0.043	0.151±0.050	0.574
	CGIP-Graph	0.851±0.227	1.075±0.159	0.237±0.045	0.190±0.085	0.588
	GraphMVP	1.157±0.196	0.969±0.120	0.229±0.053	0.138±0.044	0.623
Geometry Graph	Uni-Mol-R (1 conf)	4.156±0.212	3.140±0.203	1.151±0.160	0.606±0.053	2.263
	Uni-Mol (1 conf)	4.195±0.263	3.203±0.265	1.184±0.166	0.600±0.039	2.296
	TFN	**0.380±0.060**	**0.452±0.203**	0.280±0.063	**0.125±0.016**	**0.309**
	SE3_Transformer	2.580±0.939	3.432±1.017	1.079±0.428	0.478±0.217	1.892
Image	ResNet18-I-R	2.508±0.234	2.515±0.192	1.010±0.070	0.486±0.038	1.630
	ImageMol	2.535±0.183	2.358±0.173	1.012±0.071	0.499±0.049	1.601
	CGIP-Image	2.503±0.283	2.324±0.153	1.025±0.079	0.510±0.044	1.591
	MaskMol	5.066±0.329	3.720±0.356	1.237±0.187	0.601±0.036	2.656
	IEM-I	2.655±0.232	2.423±0.178	1.059±0.078	0.543±0.111	1.670
Geometry Image	ResNet18-G-R	2.167±0.205	2.519±0.189	1.079±0.160	0.578±0.036	1.586
	IEM-G (1 conf)	2.236±0.224	2.402±0.102	1.008±0.093	0.591±0.030	1.560
Video	ResNet18-V-R	1.310±0.150	1.704±0.122	0.740±0.138	0.437±0.167	1.048
	IEM-V	1.194±0.157	1.596±0.138	0.650±0.069	0.421±0.131	0.965

methods (ImageMol, CGIP-Image, MaskMol and IEM-I) produce negative transfer in performance. This shows that for complete chain molecules, we need to further design more suitable pre-training tasks to improve the performance of the model on this type of molecules.

I.4 Macro Molecules

Table S37 and Table S38 show the average RMSE performance and corresponding standard deviation of the baselines on 10 macro-molecule datasets, respectively. In general, the graph-based methods show great advantages on macromolecules because the green areas are concentrated in the graph-based methods. At the same time, we observe that all molecules in the dataset have rings. Therefore, this indicates that graph-based models are suitable for macro molecules with rings. Furthermore, we find that molecular fingerprints are the second best modality overall compared to the graph modality, indicating that fingerprints are a good alternative for macro molecules with rings. We find that half of the graph-based methods have the problem of negative transfer on macro molecules, such as EdgePred, infomax, masking, MoleBert, which deserves further study in the future.

I.5 Macrocyclic Peptide Molecules

Table S39 and Table S40 show the average RMSE performance and corresponding standard deviation of the baselines on 10 macrocyclic peptide datasets, respectively. Overall, image-based methods show great advantages as 4 (CGIP-Image, MaskMol, IEM-I, ResNet18-I-R) out of the top 5 baselines are vision-based methods, which shows the advantages of image modality on macrocyclic peptides. Additionally, we found the limitation of fingerprinting on macrocyclic peptides, with only 2 results out of 66 achieving top-5 performance. If we only observe the non-pretrained methods, we

Table S28: The average RMSE performance on MBANet$_{attr}$ with 10 seeds. The modality types from top to bottom are sequence, graph, geometry graph, image, geometry image, and video. -R means no pre-training, L means the number of layers, -I, -G, and -V mean the modalities are image, geometry image, and video, respectively. The green background represents top-5 performance.

Models	MW	LogP	MR	BalabanJ	#HA	#HD	#VE	TPSA	Mean
BERT-6L	39.046	1.087	9.068	0.572	1.261	0.997	15.733	23.823	11.448
BERT-8L	39.414	1.092	8.988	0.579	1.198	0.939	15.709	23.853	11.471
RoBERTa-12L	47.678	0.969	10.315	0.415	1.401	0.990	18.931	24.872	13.197
molformer-R	45.182	1.121	10.428	0.534	1.261	1.051	18.099	25.567	12.906
Chem-BERT-6L	38.875	1.030	8.749	0.541	1.240	0.968	15.677	23.714	11.349
Chem-BERT-8L	38.660	1.043	8.813	0.600	1.170	0.941	15.539	23.422	11.273
CHEM-RoBERTa-12L	47.740	0.962	10.214	0.539	1.395	0.954	18.540	24.814	13.145
Molformer	43.787	1.105	9.434	0.624	1.295	1.096	17.079	25.100	12.440
GIN-R	30.627	0.944	9.947	0.530	0.800	0.877	13.438	10.946	8.514
EdgePred	26.004	0.825	7.613	0.661	0.811	0.748	9.433	8.925	6.878
ContextPred	27.664	0.822	7.389	0.599	0.773	0.754	9.318	8.387	6.963
infomax	26.379	0.761	7.152	0.585	0.721	0.694	9.176	8.827	6.787
masking	25.193	0.713	6.516	0.502	0.694	0.846	9.081	8.740	6.536
MolCLR	28.482	0.803	9.801	0.603	0.675	0.685	11.910	10.982	7.993
MoleBERT	23.475	**0.650**	6.062	0.495	0.788	0.714	8.871	8.795	6.231
CGIP-Graph	16.019	0.674	4.833	**0.384**	**0.614**	**0.447**	6.075	4.991	4.255
GraphMVP	28.781	0.761	9.264	0.704	0.662	0.813	11.588	10.157	7.841
Uni-Mol-R (1 conf)	36.526	1.081	9.017	0.640	1.103	1.171	14.486	22.852	10.859
Uni-Mol (1 conf)	37.862	1.101	8.893	0.533	0.918	1.095	14.419	22.263	10.886
TFN	10.075	0.688	3.408	0.534	0.713	0.654	3.380	5.279	3.091
SE3_Transformer	33.948	0.888	11.641	0.568	1.126	0.920	19.913	24.265	11.659
ResNet18-I-R	23.070	0.921	6.251	0.601	0.935	0.746	8.721	14.364	6.951
ImageMol	21.691	0.908	5.686	0.629	0.807	0.811	7.971	13.566	6.509
CGIP-Image	23.269	0.955	6.382	0.494	0.851	0.801	8.844	14.549	7.018
MaskMol	40.568	1.073	9.277	0.553	1.149	0.993	16.432	24.063	11.764
IEM-I	22.673	0.865	5.906	0.544	0.871	0.784	8.365	14.144	6.769
ResNet18-G-R	15.357	0.813	4.709	0.574	0.794	0.655	6.930	8.951	4.848
IEM-G (1 conf)	15.694	0.745	4.834	0.468	0.713	0.717	6.960	9.193	4.916
ResNet18-V-R	8.121	0.750	2.661	0.582	0.711	0.656	3.870	**3.929**	2.660
IEM-V	**6.542**	0.703	**2.280**	0.417	0.726	0.707	**2.699**	3.961	**2.254**

find that the 3 vision-based modalities are the best because their performance is in the top 3. This suggests that we can make some further efforts in the future and propose some vision-based methods for the prediction of macrocyclic peptides.

I.6 Reticular Molecules

Table S41 and Table S42 show the average RMSE performance and corresponding standard deviation of the baselines on 10 reticular datasets, respectively. We find that image-based methods without any pre-training achieve the best average performance on reticular molecules compared to many pre-trained methods, which suggests that the image modality is suitable for processing reticular molecules. If we only look at the non-pretrained methods, the three vision-based modalities achieve the best top 3 performance. In addition, we find that all vision-based pre-training methods suffer from negative transfer problems on reticular molecules, which deserves to be further studied and explored in the future.

J Computational Efficiency

In virtual screening, computational efficiency of models is very important. Here, we analyze the number of parameters of different modal methods and their computational efficiency in training and inference. All evaluation are performed on 1 GeForce RTX 4090 GPU and with a batch size of 8. As shown in Table S43, we find that the video modality takes the most time. This is because a molecular video consists of 60 frames, which greatly increases the time cost. Secondly, we find that the image, SMILES and geometry graph modalities have larger parameter counts, such as MaskMol, CHEM-RoBERTa and Uni-Mol, which is due to the fact that they utilize the architecture of transformer and its variants.

Table S29: The standard deviation performance on MBANet$_{attr}$ with 10 seeds. The modality types from top to bottom are sequence, graph, geometry graph, image, geometry image, and video. -R means no pre-training, L means the number of layers, -I, -G, and -V mean the modalities are image, geometry image, and video, respectively.

Models	MW	LogP	MR	BalabanJ	#HA	#HD	#VE	TPSA
BERT-6L	1.476	0.253	0.866	0.209	0.434	0.176	0.769	1.356
BERT-8L	1.547	0.259	0.916	0.213	0.310	0.145	0.779	1.373
RoBERTa-12L	2.329	0.077	0.910	0.170	0.313	0.140	1.335	2.093
molformer-R	1.529	0.213	1.332	0.247	0.252	0.277	1.360	2.695
Chem-BERT-6L	1.452	0.158	1.199	0.277	0.299	0.182	0.810	1.246
Chem-BERT-8L	1.734	0.159	0.989	0.294	0.274	0.167	0.938	1.322
CHEM-RoBERTa-12L	2.195	0.058	0.938	0.262	0.306	0.117	1.239	2.150
Molformer	1.678	0.210	1.102	0.285	0.294	0.329	1.072	2.522
GIN-R	2.209	0.290	1.356	0.178	0.192	0.269	0.939	2.655
EdgePred	2.390	0.213	1.193	0.231	0.161	0.110	1.481	1.197
ContextPred	2.546	0.180	1.127	0.169	0.178	0.084	0.985	1.142
infomax	3.192	0.137	0.920	0.186	0.116	0.089	0.765	1.403
masking	3.185	0.148	0.774	0.116	0.070	0.195	0.873	1.323
MolCLR	3.530	0.254	1.126	0.220	0.100	0.104	1.155	2.636
MoleBERT	2.107	0.045	0.824	0.163	0.138	0.080	1.129	0.654
CGIP-Graph	1.406	0.188	0.698	0.171	0.210	0.108	0.719	0.667
GraphMVP	3.009	0.179	1.105	0.196	0.095	0.105	1.259	1.657
Uni-Mol-R (1 conf)	4.428	0.136	1.514	0.305	0.183	0.208	2.452	2.007
Uni-Mol (1 conf)	3.155	0.155	1.411	0.244	0.098	0.297	1.957	1.797
TFN	1.837	0.122	0.679	0.244	0.092	0.065	0.506	1.458
SE3_Transformer	**0.083**	**0.003**	**0.039**	**0.025**	**0.019**	**0.009**	**0.026**	**0.017**
ResNet18-I-R	0.718	0.073	0.692	0.223	0.185	0.090	0.922	1.735
ImageMol	1.206	0.105	0.342	0.319	0.089	0.134	0.871	0.527
CGIP-Image	0.931	0.176	0.308	0.155	0.133	0.114	0.781	1.046
MaskMol	1.477	0.247	0.773	0.244	0.246	0.127	1.449	1.322
IEM-I	1.057	0.065	0.380	0.238	0.106	0.078	0.974	0.717
ResNet18-G-R	1.190	0.156	0.484	0.162	0.132	0.085	0.616	0.499
IEM-G (1 conf)	1.812	0.043	0.898	0.151	0.063	0.129	1.103	0.664
ResNet18-V-R	1.083	0.102	0.363	0.274	0.224	0.126	0.704	0.408
IEM-V	0.845	0.085	0.107	0.136	0.205	0.185	0.496	0.539

Given the high temporal cost of the video modality, we further discuss the impact of different numbers of frames on the computational cost. As shown in Table S44, We find that adjusting the number of frames can effectively improve computational efficiency. Therefore, we can try to reduce the number of frames when computing resources are limited.

Next, we analyze how long it takes the model to perform virtual screening on 10,000 molecules. As shown in Table S45, we find that the video modality required more time for virtual screening, while the other modalities took comparable time.

K In-depth Analysis of Insights

K.1 Prediction Diversity Between Different Modalities

Here, we study the pairwise diversity of different modalities, including feature diversity and prediction diversity. We use mcfp_2048, Chem-BERT-8L, MoleBERT, Uni-Mol (10 conf), MaskMol, IEM-G, and VideoMol as representatives of each modality (fingerprint, sequence, geometry graph, image, geometry image, video) because they achieve excellent performance on 8 molecular property prediction tasks based on classification tasks. The ROC-AUC performances of mcfp_2048, Chem-BERT-8L, MoleBERT, Uni-Mol (10 conf), MaskMol, IEM-G (10 conf), and VideoMol on the HIV test set are 74.1%, 78.8%, 76.7%, 77.8%, 75.1%, 80.5% and 78.3% respectively.

As shown in Table S46 and Table S47, we find that different modalities have different degrees of differences in the logits of predicting HIV, including RMSE differences and Pearson differences, which provides evidence that fusing different molecular modalities can increase the diversity of information to improve performance. It is worth noting that we find that **video have the largest differences with other modalities in RMSE, which may provide a direction for future multi-modal fusion**.

Table S30: The RMSE performance on $\text{MBANet}^{30K}_{atom}$, $\text{MBANet}^{30K}_{bond}$, $\text{MBANet}^{30K}_{attr}$ with 3 seeds. *-R, TFN and SE3_Transformer represent non-pre-trained models, and the others are pre-trained models.

	Models	$\text{MBANet}^{30K}_{atom}$	$\text{MBANet}^{30K}_{bond}$	$\text{MBANet}^{30K}_{attr}$
SMILES-based	BERT-6L	0.647	2.020	10.961
	BERT-8L	0.645	2.014	10.936
	RoBERTa-12L	0.644	2.030	11.223
	molformer-R	0.646	2.042	11.159
	Chem-BERT-6L	0.647	2.019	10.960
	Chem-BERT-8L	0.647	2.019	10.960
	CHEM-RoBERTa-12L	0.644	2.042	11.249
	Molformer	0.646	2.046	11.031
Graph-based	GIN-R	0.339	0.627	6.711
	EdgePred	0.306	0.613	6.480
	ContextPred	0.312	0.605	6.508
	infomax	0.325	0.612	6.313
	masking	0.315	0.606	6.323
	MolCLR	0.329	0.627	6.873
	MoleBERT	0.309	0.607	6.259
	CGIP-Graph	0.243	0.574	5.710
	GraphMVP	0.317	0.621	6.868
Geometry-based	Uni-Mol-R (1 conf)	0.504	1.977	10.299
	Uni-Mol (1 conf)	0.510	1.979	10.247
	TFN	0.187	**0.239**	2.913
	SE3_Transformer	0.642	2.070	11.087
Image-based	ResNet18-I-R	0.359	1.176	6.263
	ImageMol	0.355	1.156	6.224
	CGIP-Image	0.358	1.163	6.400
	MaskMol	0.641	1.984	10.748
	IEM-I	0.358	1.198	6.433
Geometry-based	ResNet18-G-R	0.242	1.139	5.164
	IEM-G (1 conf)	0.245	1.136	5.315
Video-based	ResNet18-V-R	0.167	0.843	2.853
	IEM-V	**0.135**	0.829	**2.359**

Table S31: The average RMSE performance on 10 acyclic chain (AC) datasets from StructNet. AC#1, AC#2, AC#3, AC#4, AC#5, AC#6, AC#7, AC#8, AC#9 and AC#10 represent CHEMBL1614458_Potency, CHEMBL4513082_Inhibition, CHEMBL4495582_Inhibition, CHEMBL4296187_Inhibition, CHEMBL4296188_Inhibition, CHEMBL1614361_Potency, CHEMBL4303805_Inhibition, CHEMBL4649955_Potency, CHEMBL4649949_Potency and CHEMBL4649948_Potency, respectively. The green background represents top-5 performance.

Models	AC#1	AC#2	AC#3	AC#4	AC#5	AC#6	AC#7	AC#8	AC#9	AC#10	Mean
mcfp4_2048	0.932	0.143	17.976	19.208	7.337	1.226	29.222	14.469	7.497	9.194	10.720
ecfp4_2048	0.954	0.147	18.022	19.186	7.399	1.290	29.415	14.468	7.482	9.194	10.756
maccs	1.044	0.151	17.191	19.148	7.091	1.159	29.277	14.473	7.468	9.190	10.619
physchem	0.795	0.155	18.122	19.271	7.318	1.159	29.234	14.483	7.517	9.202	10.726
atompair_2048	1.098	0.152	17.962	**18.759**	7.241	1.420	29.572	14.474	**7.301**	9.192	10.717
rdkDes	1.049	**0.133**	17.167	19.349	6.984	1.317	29.046	14.470	7.497	9.186	10.620
BERT-6L	**0.757**	0.400	15.976	19.173	6.818	1.175	28.848	14.499	7.594	9.379	10.462
BERT-8L	**0.757**	0.392	15.977	19.179	6.822	1.167	28.892	14.502	7.585	9.350	10.462
RoBERTa-12L	0.758	0.370	15.959	19.219	6.817	1.177	28.730	14.500	7.603	9.367	10.450
molformer-R	0.758	0.389	15.952	19.168	6.822	1.176	28.682	14.496	7.595	9.353	10.439
Chem-BERT-6L	**0.757**	0.442	15.980	19.164	6.821	1.171	28.918	14.502	7.597	9.348	10.470
Chem-BERT-8L	**0.757**	0.426	15.994	19.169	6.822	1.177	28.997	14.499	7.597	9.365	10.480
CHEM-RoBERTa-12L	0.760	0.416	15.984	19.196	6.818	1.174	28.965	14.502	7.601	9.359	10.478
Molformer	0.758	0.414	15.962	19.132	6.820	1.171	28.681	14.498	7.590	9.342	10.437
GIN-R	0.774	0.261	14.889	19.617	7.059	1.187	28.948	14.524	7.767	9.564	10.459
EdgePred	0.968	0.152	16.519	19.331	6.899	1.175	29.197	14.382	7.451	9.184	10.526
ContextPred	0.780	0.183	15.162	19.280	7.100	1.240	29.121	14.463	7.364	**9.086**	10.378
infomax	0.795	0.238	14.463	19.223	6.971	1.301	29.171	14.412	7.676	9.202	10.345
masking	**0.757**	0.185	14.693	19.193	7.122	1.208	29.123	14.463	7.699	9.224	10.367
MolCLR	0.773	0.200	14.899	19.312	6.817	1.195	28.959	**14.308**	7.681	9.445	10.359
MoleBERT	0.823	0.172	14.211	19.087	**6.713**	1.309	29.213	14.481	7.462	9.178	**10.265**
CGIP-Graph	0.806	0.160	14.426	19.914	6.859	1.264	29.180	14.442	7.876	9.290	10.422
GraphMVP	0.820	0.231	13.950	19.430	7.132	1.244	29.649	14.449	7.860	9.191	10.396
SchNet	0.828	0.324	**13.176**	19.230	7.347	1.360	29.318	14.485	7.513	9.262	10.284
EGNN	0.759	0.217	15.898	19.134	6.818	1.212	29.331	14.446	7.534	9.318	10.467
TFN	0.883	0.469	15.922	18.951	7.101	1.448	28.928	14.382	7.608	9.275	10.497
SE3_Transformer	0.868	0.334	15.426	18.934	6.967	1.435	29.001	14.621	7.886	9.537	10.501
PaiNN	2.365	0.708	15.725	18.935	7.885	1.604	30.369	14.485	7.993	9.645	10.971
Uni-Mol-R (1 conf)	0.758	0.411	15.935	19.185	6.819	1.175	28.797	14.499	7.606	9.370	10.455
Uni-Mol (1 conf)	0.758	0.418	15.864	19.196	6.817	1.179	28.814	14.483	7.568	9.353	10.445
ResNet18-I-R	0.759	0.328	16.341	19.140	6.888	1.195	29.084	14.455	7.908	9.404	10.550
ImageMol	0.781	0.355	16.109	19.259	6.970	1.252	29.039	14.368	7.948	9.470	10.555
CGIP-Image	0.768	0.247	16.281	19.147	6.897	1.196	28.980	14.525	8.170	9.625	10.584
MaskMol	0.759	0.404	15.962	19.161	6.821	1.172	28.944	14.491	7.576	9.319	10.461
IEM-I	0.762	0.199	16.288	19.166	6.819	1.229	29.279	14.421	7.879	9.498	10.554
ResNet18-G-R	0.775	0.334	16.538	18.802	6.737	**1.139**	29.115	14.524	7.560	9.290	10.481
IEM-G (1 conf)	0.771	0.218	15.876	19.294	6.996	1.155	28.986	14.733	7.664	9.242	10.494
ViT-G-R	0.758	0.359	15.962	19.165	6.827	1.176	**28.662**	14.462	7.582	9.344	10.430
VideoMol-G	0.759	0.401	15.970	19.157	6.822	1.172	28.680	14.476	7.580	9.350	10.437
ResNet18-V-R	0.769	0.359	16.196	18.761	6.821	1.156	29.119	14.435	7.475	9.323	10.441
IEM-V	0.771	0.210	15.798	18.949	6.813	1.175	29.280	14.432	7.460	9.311	10.420

Table S32: The standard deviation on 10 acyclic chain (AC) datasets from StructNet. AC#1, AC#2, AC#3, AC#4, AC#5, AC#6, AC#7, AC#8, AC#9 and AC#10 represent CHEMBL1614458_Potency, CHEMBL4513082_Inhibition, CHEMBL4495582_Inhibition, CHEMBL4296187_Inhibition, CHEMBL4296188_Inhibition, CHEMBL1614361_Potency, CHEMBL4303805_Inhibition, CHEMBL4649955_Potency, CHEMBL4649949_Potency and CHEMBL4649948_Potency, respectively.

Models	AC#1	AC#2	AC#3	AC#4	AC#5	AC#6	AC#7	AC#8	AC#9	AC#10
mcfp4_2048	0.008	0.003	0.056	0.008	0.009	0.004	0.013	0.005	0.003	0.002
ecfp4_2048	0.008	0.004	0.053	0.008	0.008	0.004	0.021	0.005	0.004	0.002
maccs	0.011	0.006	0.045	0.006	0.009	0.004	0.025	0.006	0.009	0.006
physchem	0.013	0.028	0.050	0.012	0.026	0.008	0.020	0.012	0.006	0.013
atompair_2048	0.008	0.003	0.056	0.011	0.006	0.004	0.028	0.007	0.005	0.003
rdkDes	0.013	0.003	0.085	0.009	0.008	0.007	0.015	0.006	0.007	0.008
BERT-6L	0.000	0.062	0.060	0.050	0.005	0.013	0.253	0.007	0.019	0.034
BERT-8L	0.000	0.058	0.069	0.066	0.007	0.013	0.269	0.003	0.018	0.053
RoBERTa-12L	0.002	0.082	0.022	0.061	0.003	0.009	0.406	0.006	0.017	0.029
molformer-R	0.002	0.082	0.005	0.034	0.005	0.004	0.216	0.008	0.016	0.042
Chem-BERT-6L	0.001	0.077	0.044	0.041	0.015	0.009	0.310	0.005	0.027	0.038
Chem-BERT-8L	0.000	0.094	0.059	0.051	0.012	0.005	0.241	0.006	0.019	0.041
CHEM-RoBERTa-12L	0.003	0.081	0.038	0.049	0.004	0.008	0.315	0.009	0.022	0.030
Molformer	0.001	0.097	0.031	0.042	0.006	0.010	0.249	0.011	0.007	0.073
GIN-R	0.017	0.101	1.137	0.412	0.265	0.047	0.305	0.194	0.279	0.341
EdgePred	0.558	0.029	1.804	0.107	0.109	0.033	0.073	0.257	0.199	0.156
ContextPred	0.023	0.028	0.473	0.117	0.145	0.022	0.142	0.045	0.043	0.162
infomax	0.026	0.047	0.333	0.168	0.166	0.034	0.093	0.092	0.303	0.130
masking	0.014	0.020	1.020	0.080	0.328	0.016	0.237	0.030	0.177	0.102
MolCLR	0.028	0.049	0.570	0.215	0.269	0.024	0.430	0.176	0.145	0.271
MoleBERT	0.045	0.020	0.555	0.172	0.073	0.031	0.128	0.026	0.069	0.019
CGIP-Graph	0.018	0.032	1.051	0.721	0.208	0.055	0.374	0.042	0.131	0.142
GraphMVP	0.076	0.047	1.503	0.528	0.234	0.068	0.498	0.064	0.394	0.076
SchNet	0.030	0.124	1.182	0.634	0.283	0.076	0.395	0.080	0.096	0.172
EGNN	0.001	0.148	0.089	0.065	0.011	0.025	0.195	0.143	0.024	0.359
TFN	0.093	0.157	0.598	0.110	0.149	0.052	0.266	0.231	0.129	0.143
SE3_Transformer	0.045	0.098	0.721	0.182	0.182	0.058	0.472	0.267	0.303	0.231
PaiNN	0.305	0.240	0.915	0.366	0.443	0.138	0.562	0.244	0.280	0.365
Uni-Mol-R (1 conf)	0.001	0.068	0.064	0.091	0.005	0.006	0.154	0.008	0.022	0.034
Uni-Mol (1 conf)	0.001	0.089	0.161	0.067	0.026	0.015	0.178	0.071	0.050	0.049
ResNet18-I-R	0.010	0.113	0.217	0.174	0.095	0.038	0.399	0.119	0.326	0.181
ImageMol	0.023	0.184	0.421	0.398	0.267	0.057	0.694	0.161	0.531	0.539
CGIP-Image	0.013	0.099	0.663	0.602	0.248	0.036	0.337	0.111	0.419	0.650
MaskMol	0.003	0.074	0.069	0.053	0.005	0.015	0.262	0.020	0.061	0.039
IEM-I	0.009	0.038	0.772	0.178	0.317	0.059	0.464	0.183	0.417	0.571
ResNet18-G-R	0.010	0.076	0.368	0.290	0.159	0.020	0.354	0.175	0.205	0.097
IEM-G (1 conf)	0.026	0.110	0.268	0.607	0.231	0.021	0.381	0.634	0.189	0.172
ViT-G-R	0.001	0.078	0.141	0.080	0.014	0.023	0.165	0.120	0.026	0.046
VideoMol-G	0.002	0.120	0.225	0.056	0.007	0.023	0.070	0.030	0.031	0.043
ResNet18-V-R	0.009	0.081	0.264	0.118	0.030	0.018	0.141	0.063	0.102	0.088
IEM-V	0.022	0.088	0.284	0.224	0.191	0.028	0.278	0.147	0.181	0.207

Table S33: The average RMSE performance on 10 acyclic (A) datasets from StructNet. A#1, A#2, A#3, A#4, A#5, A#6, A#7, A#8, A#9, A#10 represent CHEMBL4513082_Inhibition, CHEMBL4495582_Inhibition, CHEMBL1614458_Potency, CHEMBL4303805_Inhibition, CHEMBL4808149_Inhibition, CHEMBL4296187_Inhibition, CHEMBL4808150_Inhibition, CHEMBL4296188_Inhibition, CHEMBL4649955_Potency, CHEMBL4649949_Potency, respectively. The green background represents top-5 performance.

Models	A#1	A#2	A#3	A#4	A#5	A#6	A#7	A#8	A#9	A#10	Mean
mcfp4_2048	0.134	17.750	0.666	15.248	26.727	17.738	14.628	6.490	13.084	13.108	12.557
ecfp4_2048	0.130	17.746	0.626	15.214	26.740	17.713	14.622	6.544	13.029	13.147	12.551
maccs	0.140	17.387	0.670	**15.129**	26.760	17.704	14.634	6.537	13.085	13.109	12.516
physchem	0.154	17.765	**0.531**	15.208	26.739	17.823	14.634	6.430	13.073	13.107	12.546
atompair_2048	0.135	17.996	0.782	15.394	26.721	17.548	14.626	6.431	13.104	12.989	12.573
rdkDes	0.139	17.284	0.697	15.379	26.736	17.843	14.629	6.369	**12.982**	13.019	12.508
BERT-6L	0.131	17.418	0.566	15.311	27.162	17.480	14.636	6.437	13.004	13.079	12.522
BERT-8L	0.130	17.410	0.566	15.306	27.158	17.480	14.637	6.425	13.006	13.081	12.520
RoBERTa-12L	0.133	17.415	0.567	15.370	27.225	17.477	14.636	6.401	13.001	13.052	12.528
molformer-R	0.130	17.423	0.566	15.369	27.206	17.479	14.650	6.386	13.005	13.100	12.531
Chem-BERT-6L	0.131	17.410	0.566	15.406	27.194	17.479	14.638	6.425	13.004	13.070	12.532
Chem-BERT-8L	0.132	17.417	0.567	15.302	27.178	17.479	14.639	6.425	13.001	13.066	12.521
CHEM-RoBERTa-12L	0.132	17.420	0.566	15.397	27.186	17.409	14.638	6.413	13.002	13.031	12.519
Molformer	0.131	17.421	0.566	15.277	27.167	17.476	14.637	6.380	13.003	13.045	12.510
GIN-R	0.191	15.578	0.590	15.626	27.423	18.694	14.554	6.319	13.169	13.217	12.536
EdgePred	0.147	49.805	0.578	15.623	26.868	17.992	13.821	6.215	13.253	13.072	15.737
ContextPred	0.185	12.660	0.585	15.560	26.788	17.876	13.928	5.881	13.180	12.875	11.952
infomax	0.258	14.447	0.545	15.551	26.784	17.795	14.156	6.176	13.288	**12.475**	12.147
masking	0.156	13.128	0.558	15.562	26.814	17.769	14.119	6.133	13.330	13.102	12.067
MolCLR	0.137	15.522	0.575	15.826	27.120	17.939	14.544	6.352	13.077	13.121	12.421
MoleBERT	0.216	12.496	0.552	15.343	26.824	17.704	**13.605**	**5.633**	13.124	12.936	**11.843**
CGIP-Graph	0.198	12.395	0.603	16.262	**26.659**	18.565	14.677	5.770	13.118	12.598	12.085
GraphMVP	0.142	**11.831**	0.599	16.323	26.701	19.174	14.528	6.195	13.233	12.997	12.172
SchNet	0.222	13.636	0.610	16.680	27.419	17.531	15.177	6.390	13.279	13.253	12.420
EGNN	0.134	16.746	0.576	16.158	27.181	17.527	14.781	6.403	13.072	13.110	12.569
TFN	0.177	14.336	0.641	15.314	27.021	17.453	14.530	6.396	13.071	12.981	12.192
SE3_Transformer	0.238	15.176	0.614	15.491	27.222	17.475	14.308	6.304	13.118	13.172	12.312
PaiNN	0.696	13.416	2.167	18.349	27.323	**17.384**	16.141	6.584	13.122	13.161	12.834
Uni-Mol-R (1 conf)	0.132	17.340	0.566	15.348	27.166	17.481	14.639	6.433	13.004	13.070	12.518
Uni-Mol (1 conf)	0.128	17.376	0.566	15.315	27.197	17.476	14.639	6.445	13.002	13.097	12.524
ResNet18-I-R	0.138	16.694	0.563	15.507	27.018	17.474	14.811	6.434	13.090	12.996	12.473
ImageMol	0.151	16.214	0.579	16.242	27.165	17.776	14.850	6.419	13.295	13.152	12.584
CGIP-Image	0.150	16.349	0.571	16.651	26.953	17.626	14.790	6.434	13.458	12.982	12.596
MaskMol	0.131	17.421	0.566	15.463	27.129	17.481	14.651	6.419	13.002	13.091	12.535
IEM-I	0.149	15.975	0.576	16.437	27.383	17.741	15.097	6.432	13.101	13.196	12.609
ResNet18-G-R	0.138	16.497	0.554	15.922	27.006	17.590	14.619	6.388	13.092	13.010	12.482
IEM-G (1 conf)	0.157	15.366	0.575	16.170	26.945	17.570	14.585	6.411	13.040	13.208	12.403
ViT-G-R	0.130	17.219	0.565	15.261	27.183	17.479	14.632	6.381	13.006	13.048	12.490
VideoMol-G	**0.127**	17.197	0.562	15.282	27.145	17.457	14.637	6.422	13.008	13.042	12.488
ResNet18-V-R	0.137	16.421	0.551	15.785	27.007	17.519	14.554	6.388	13.019	13.063	12.444
IEM-V	0.135	13.842	0.556	16.536	26.973	17.539	14.681	6.415	13.053	13.075	12.281

Table S34: The standard deviation on 10 acyclic (A) datasets from StructNet. A#1, A#2, A#3, A#4, A#5, A#6, A#7, A#8, A#9, A#10 represent CHEMBL4513082_Inhibition, CHEMBL4495582_Inhibition, CHEMBL1614458_Potency, CHEMBL4303805_Inhibition, CHEMBL4808149_Inhibition, CHEMBL4296187_Inhibition, CHEMBL4808150_Inhibition, CHEMBL4296188_Inhibition, CHEMBL4649955_Potency, CHEMBL4649949_Potency, respectively.

Models	A#1	A#2	A#3	A#4	A#5	A#6	A#7	A#8	A#9	A#10
mcfp4_2048	0.003	0.014	0.002	0.007	0.002	0.008	0.009	0.006	0.004	0.011
ecfp4_2048	0.003	0.012	0.002	0.008	0.002	0.006	0.020	0.007	0.011	0.027
maccs	0.008	0.021	0.006	0.006	0.008	0.005	0.006	0.017	0.015	0.016
physchem	0.017	0.017	0.002	0.008	0.011	0.014	0.003	0.017	0.015	0.005
atompair_2048	0.003	0.010	0.004	0.006	0.004	0.009	0.004	0.010	0.004	0.022
rdkDes	0.013	0.009	0.005	0.005	0.007	0.012	0.005	0.014	0.013	0.067
BERT-6L	0.005	0.018	0.001	0.097	0.111	0.002	0.011	0.034	0.007	0.037
BERT-8L	0.003	0.017	0.001	0.096	0.108	0.004	0.011	0.022	0.009	0.031
RoBERTa-12L	0.012	0.012	0.004	0.109	0.138	0.004	0.011	0.058	0.005	0.059
molformer-R	0.003	0.009	0.001	0.140	0.144	0.004	0.018	0.047	0.010	0.076
Chem-BERT-6L	0.004	0.011	0.000	0.246	0.161	0.001	0.013	0.023	0.008	0.035
Chem-BERT-8L	0.005	0.012	0.003	0.132	0.161	0.003	0.014	0.041	0.008	0.059
CHEM-RoBERTa-12L	0.005	0.007	0.002	0.096	0.068	0.218	0.015	0.071	0.005	0.067
Molformer	0.005	0.006	0.001	0.027	0.069	0.009	0.015	0.059	0.008	0.077
GIN-R	0.048	2.463	0.019	0.493	0.559	0.813	0.334	0.077	0.271	0.284
EdgePred	0.014	35.577	0.020	0.302	0.155	0.082	0.460	0.215	0.110	0.123
ContextPred	0.038	2.445	0.026	0.203	0.081	0.124	0.604	0.497	0.181	0.153
infomax	0.053	3.404	0.024	0.176	0.045	0.105	0.474	0.396	0.209	0.561
masking	0.016	2.483	0.015	0.234	0.062	0.147	0.430	0.472	0.251	0.024
MolCLR	0.014	2.662	0.018	0.549	0.317	0.412	0.265	0.183	0.204	0.300
MoleBERT	0.082	1.515	0.022	0.114	0.094	0.074	0.270	0.310	0.064	0.187
CGIP-Graph	0.049	1.801	0.041	0.548	0.037	0.552	0.314	0.199	0.164	0.210
GraphMVP	0.015	2.054	0.036	0.947	0.029	0.803	0.501	0.295	0.148	0.317
SchNet	0.053	2.075	0.037	1.866	0.644	0.347	0.735	0.144	0.337	0.604
EGNN	0.016	1.135	0.020	0.926	0.256	0.143	0.178	0.036	0.021	0.085
TFN	0.036	3.056	0.047	0.158	0.570	0.187	0.492	0.103	0.138	0.177
SE3_Transformer	0.035	1.728	0.037	0.228	0.259	0.059	0.173	0.120	0.108	0.158
PaiNN	0.333	2.170	0.279	1.693	0.536	0.193	0.558	0.200	0.134	0.258
Uni-Mol-R (1 conf)	0.015	0.134	0.001	0.148	0.165	0.004	0.013	0.023	0.012	0.079
Uni-Mol (1 conf)	0.006	0.075	0.017	0.161	0.190	0.012	0.013	0.069	0.011	0.082
ResNet18-I-R	0.011	0.972	0.026	0.181	0.089	0.106	0.170	0.072	0.078	0.178
ImageMol	0.022	1.743	0.035	0.605	0.519	0.317	0.326	0.121	0.319	0.187
CGIP-Image	0.019	1.598	0.030	1.923	0.278	0.307	0.107	0.202	0.753	0.175
MaskMol	0.003	0.008	0.002	0.350	0.175	0.004	0.016	0.015	0.011	0.062
IEM-I	0.014	1.901	0.023	1.000	0.652	0.513	0.498	0.121	0.069	0.308
ResNet18-G-R	0.011	0.509	0.013	0.504	0.290	0.073	0.037	0.051	0.118	0.092
IEM-G (1 conf)	0.025	2.382	0.020	0.934	0.404	0.147	0.309	0.083	0.219	0.274
ViT-G-R	0.006	0.112	0.006	0.133	0.104	0.015	0.009	0.029	0.024	0.083
VideoMol-G	0.005	0.083	0.010	0.158	0.156	0.071	0.013	0.045	0.020	0.090
ResNet18-V-R	0.011	0.382	0.009	0.313	0.246	0.116	0.050	0.043	0.053	0.039
IEM-V	0.012	1.533	0.015	1.862	0.596	0.402	0.073	0.176	0.174	0.071

Table S35: The average RMSE performance on 10 complete chain (CC) datasets from StructNet. CC#1, CC#2, CC#3, CC#4, CC#5, CC#6, CC#7, CC#8, CC#9, CC#10 represent CHEMBL4649949_Potency, CHEMBL4649948_Potency, CHEMBL4649955_Potency, CHEMBL4888485_Inhibition, CHEMBL4296187_Inhibition, CHEMBL4296188_Inhibition, CHEMBL4296802_Inhibition, CHEMBL1614459_Potency, CHEMBL1614458_Potency, CHEMBL1614530_Potency, respectively. The green background represents top-5 performance.

Models	CC#1	CC#2	CC#3	CC#4	CC#5	CC#6	CC#7	CC#8	CC#9	CC#10	Mean
mcfp4_2048	17.626	14.213	11.076	9.284	18.121	5.909	14.239	1.051	0.602	0.948	9.307
ecfp4_2048	17.678	14.256	11.091	9.291	18.067	5.858	14.282	1.064	0.625	0.964	9.318
maccs	**17.395**	14.265	11.041	9.247	18.068	5.537	14.311	0.995	0.537	0.885	**9.228**
physchem	17.742	14.407	11.040	9.257	18.084	5.905	14.344	0.948	0.451	0.841	9.302
atompair_2048	17.660	**14.087**	11.088	9.285	18.097	5.817	**14.034**	1.064	0.534	0.874	9.254
rdkDes	17.751	14.402	11.014	9.271	18.110	5.851	14.206	**0.912**	0.433	0.894	9.284
BERT-6L	17.709	14.406	11.045	9.275	18.169	5.879	14.406	0.934	0.435	0.824	9.308
BERT-8L	17.712	14.406	11.045	9.274	18.165	5.874	14.404	0.934	0.433	0.823	9.307
RoBERTa-12L	17.737	14.407	11.045	9.272	18.266	5.870	14.409	0.934	0.434	0.823	9.320
molformer-R	17.726	14.407	11.046	9.268	18.193	5.870	14.374	0.934	0.434	0.824	9.308
Chem-BERT-6L	17.705	14.406	11.046	9.272	18.180	5.872	14.396	0.934	0.434	0.824	9.307
Chem-BERT-8L	17.713	14.406	11.045	9.271	18.159	5.874	14.388	0.934	0.433	0.823	9.305
CHEM-RoBERTa-12L	17.733	14.406	11.044	9.280	18.303	5.868	14.455	0.934	0.434	0.823	9.328
Molformer	17.718	14.407	11.046	9.276	18.197	5.870	14.383	0.934	0.432	0.823	9.309
GIN-R	17.559	14.186	11.039	9.372	18.325	5.641	14.118	0.938	0.447	0.831	9.246
EdgePred	17.914	14.217	11.055	9.288	18.701	5.552	14.202	0.957	0.438	0.821	9.314
ContextPred	17.841	14.436	11.087	9.291	18.628	5.543	14.196	0.972	**0.429**	0.821	9.324
infomax	18.285	14.604	11.040	9.211	18.331	5.527	14.115	0.995	0.438	**0.814**	9.336
masking	18.085	14.417	11.029	9.287	18.432	5.604	14.120	0.973	0.444	0.822	9.321
MolCLR	17.590	14.171	10.998	9.299	18.305	5.625	14.098	0.946	0.437	0.825	9.229
MoleBERT	18.257	14.654	11.208	9.234	18.375	5.663	14.139	0.998	0.456	0.880	9.386
CGIP-Graph	17.800	14.195	10.993	11.150	18.333	5.657	14.238	0.980	**0.429**	0.840	9.462
GraphMVP	17.589	14.122	11.025	9.320	18.422	5.567	14.088	0.946	0.441	0.827	9.235
SchNet	17.605	14.351	11.030	9.314	18.156	5.638	14.338	0.951	0.438	0.826	9.265
EGNN	17.736	14.414	11.045	9.284	18.399	5.855	14.715	0.935	0.436	0.824	9.364
TFN	17.719	14.240	11.012	9.198	17.996	5.797	14.208	0.999	0.455	0.865	9.249
SE3_Transformer	17.699	14.283	11.047	9.337	**17.923**	5.872	14.104	1.000	0.468	0.879	9.261
PaiNN	-	14.604	11.048	9.470	18.300	**5.267**	14.461	1.650	0.659	1.592	-
Uni-Mol-R (1 conf)	17.703	14.406	11.045	9.297	18.126	5.681	14.355	0.937	0.434	0.824	9.281
Uni-Mol (1 conf)	17.690	14.405	11.037	9.329	18.129	5.621	14.353	0.938	0.435	0.824	9.276
ResNet18-I-R	17.658	14.169	11.066	9.215	18.126	5.809	14.346	0.936	0.435	0.831	9.259
ImageMol	17.655	14.236	11.068	**9.167**	18.191	5.781	14.471	0.939	0.437	0.839	9.278
CGIP-Image	17.713	14.285	11.023	9.246	18.172	5.695	14.264	0.939	0.432	0.835	9.260
MaskMol	17.702	14.406	11.046	9.274	18.224	5.876	14.478	0.935	0.434	0.823	9.320
IEM-I	17.659	14.279	11.029	9.270	18.099	5.757	14.319	0.941	0.439	0.826	9.262
ResNet18-G-R	17.744	14.659	10.991	9.284	18.198	5.802	14.534	0.935	0.435	0.823	9.340
IEM-G (1 conf)	17.664	14.552	11.079	9.221	18.244	5.743	14.632	0.937	0.436	0.835	9.334
ViT-G-R	17.722	14.410	11.043	9.253	18.166	5.873	14.397	0.934	0.434	0.824	9.306
VideoMol-G	17.726	14.404	11.046	9.260	18.201	5.853	14.394	0.934	0.433	0.824	9.308
ResNet18-V-R	17.852	14.613	**10.966**	9.309	18.343	5.709	14.537	0.939	0.431	0.823	9.352
IEM-V	17.768	14.611	11.010	9.282	18.352	5.627	14.474	0.939	0.434	0.825	9.332

Table S36: The standard deviation on 10 complete chain (CC) datasets from StructNet. CC#1, CC#2, CC#3, CC#4, CC#5, CC#6, CC#7, CC#8, CC#9, CC#10 represent CHEMBL4649949_Potency, CHEMBL4649948_Potency, CHEMBL4649955_Potency, CHEMBL4888485_Inhibition, CHEMBL4296187_Inhibition, CHEMBL4296188_Inhibition, CHEMBL4296802_Inhibition, CHEMBL1614459_Potency, CHEMBL1614458_Potency, CHEMBL1614530_Potency, respectively.

Models	CC#1	CC#2	CC#3	CC#4	CC#5	CC#6	CC#7	CC#8	CC#9	CC#10
mcfp4_2048	0.007	0.001	0.001	0.009	0.004	0.005	0.003	0.002	0.002	0.002
ecfp4_2048	0.011	0.001	0.002	0.016	0.006	0.006	0.008	0.002	0.003	0.003
maccs	0.005	0.004	0.004	0.020	0.005	0.031	0.004	0.002	0.008	0.006
physchem	0.005	0.001	0.001	0.016	0.001	0.006	0.001	0.003	0.002	0.002
atompair_2048	0.005	0.004	0.005	0.015	0.001	0.131	0.005	0.013	0.003	0.005
rdkDes	0.004	0.000	0.003	0.003	0.001	0.004	0.002	0.002	0.000	0.002
BERT-6L	0.016	0.000	0.002	0.009	0.073	0.008	0.113	0.000	0.003	0.000
BERT-8L	0.017	0.000	0.003	0.016	0.074	0.013	0.111	0.001	0.003	0.000
RoBERTa-12L	0.067	0.000	0.003	0.015	0.096	0.025	0.123	0.001	0.005	0.000
molformer-R	0.024	0.000	0.001	0.017	0.037	0.014	0.056	0.000	0.004	0.001
Chem-BERT-6L	0.013	0.000	0.002	0.012	0.080	0.008	0.106	0.000	0.002	0.000
Chem-BERT-8L	0.018	0.000	0.002	0.012	0.084	0.012	0.109	0.000	0.003	0.000
CHEM-RoBERTa-12L	0.069	0.000	0.005	0.009	0.094	0.018	0.112	0.001	0.004	0.000
Molformer	0.027	0.000	0.003	0.013	0.044	0.014	0.047	0.000	0.003	0.000
GIN-R	0.296	0.125	0.041	0.105	0.202	0.096	0.159	0.015	0.021	0.009
EdgePred	0.268	0.160	0.100	0.059	0.282	0.092	0.208	0.012	0.005	0.008
ContextPred	0.383	0.256	0.108	0.078	0.276	0.073	0.221	0.014	0.006	0.010
infomax	0.479	0.205	0.087	0.072	0.169	0.082	0.171	0.025	0.009	0.018
masking	0.306	0.263	0.090	0.047	0.177	0.113	0.247	0.009	0.012	0.018
MolCLR	0.230	0.176	0.046	0.074	0.139	0.091	0.161	0.013	0.020	0.005
MoleBERT	0.563	0.180	0.116	0.092	0.153	0.179	0.090	0.023	0.017	0.031
CGIP-Graph	0.282	0.187	0.065	2.305	0.183	0.153	0.338	0.020	0.007	0.037
GraphMVP	0.233	0.122	0.048	0.083	0.291	0.097	0.224	0.018	0.015	0.009
SchNet	0.144	0.165	0.041	0.106	0.117	0.227	0.278	0.011	0.011	0.009
EGNN	0.122	0.080	0.058	0.046	0.628	0.060	0.473	0.001	0.007	0.001
TFN	0.259	0.264	0.053	0.109	0.161	0.116	0.114	0.023	0.012	0.026
SE3_Transformer	0.069	0.112	0.041	0.131	0.083	0.088	0.056	0.026	0.021	0.035
PaiNN	-	0.265	0.092	0.098	0.096	0.120	0.168	0.156	0.106	0.195
Uni-Mol-R (1 conf)	0.029	0.001	0.003	0.040	0.033	0.188	0.054	0.003	0.003	0.000
Uni-Mol (1 conf)	0.041	0.004	0.014	0.103	0.036	0.146	0.058	0.005	0.003	0.000
ResNet18-I-R	0.296	0.139	0.051	0.130	0.120	0.075	0.198	0.007	0.012	0.008
ImageMol	0.232	0.297	0.092	0.136	0.271	0.078	0.323	0.009	0.005	0.019
CGIP-Image	0.217	0.162	0.025	0.119	0.122	0.100	0.143	0.009	0.005	0.016
MaskMol	0.006	0.001	0.005	0.013	0.057	0.008	0.193	0.001	0.002	0.001
IEM-I	0.249	0.290	0.081	0.083	0.231	0.076	0.254	0.006	0.009	0.008
ResNet18-G-R	0.135	0.195	0.041	0.057	0.109	0.041	0.200	0.009	0.006	0.005
IEM-G (1 conf)	0.145	0.121	0.113	0.056	0.114	0.094	0.391	0.009	0.007	0.014
ViT-G-R	0.032	0.010	0.006	0.045	0.083	0.019	0.116	0.000	0.001	0.001
VideoMol-G	0.037	0.012	0.003	0.046	0.124	0.023	0.080	0.001	0.003	0.001
ResNet18-V-R	0.152	0.126	0.023	0.051	0.098	0.051	0.249	0.004	0.005	0.002
IEM-V	0.115	0.107	0.027	0.093	0.207	0.156	0.171	0.006	0.010	0.009

Table S37: The average RMSE performance on 10 macro (M) datasets from StructNet. M#1, M#2, M#3, M#4, M#5, M#6, M#7, M#8, M#9, M#10 represent CHEMBL4420282_IC50, CHEMBL4419606_IC50, CHEMBL4420281_In, CHEMBL3881498_In, CHEMBL4419605_In, CHEMBL4420271_In, CHEMBL4419595_In, CHEMBL3881499_IC50, CHEMBL4420273_In, CHEMBL4419597_In, respectively. OOM represents the Exception of Out Of Memory (OOM) on single GPU of RTX 4090 Ti (24G). The green background represents top-5 performance.

Models	M#1	M#2	M#3	M#4	M#5	M#6	M#7	M#8	M#9	M#10	Mean
mcfp4_2048	0.842	0.709	36.331	35.451	36.827	26.782	26.682	0.564	10.862	10.851	18.590
ecfp4_2048	0.710	0.713	36.158	33.518	36.615	26.653	26.561	0.575	10.713	10.688	18.290
maccs	0.869	0.786	36.413	34.200	36.770	26.679	26.492	0.601	**10.637**	**10.616**	18.406
physchem	1.076	0.831	35.838	32.765	36.639	26.662	26.570	0.600	10.980	10.952	18.291
atompair_2048	**0.612**	**0.628**	35.734	33.803	35.983	26.669	26.585	**0.503**	11.601	11.564	18.368
rdkDes	1.041	0.832	40.833	36.886	41.905	29.449	29.207	0.606	11.388	11.377	20.352
BERT-6L	1.300	0.903	36.815	35.817	36.874	26.559	26.571	0.582	11.909	11.853	18.918
BERT-8L	1.306	0.898	36.823	35.839	36.876	26.580	26.568	0.583	11.872	11.878	18.922
RoBERTa-12L	1.329	0.906	37.413	36.289	37.288	26.644	26.636	0.583	11.894	11.878	19.086
molformer-R	1.337	0.906	37.330	36.212	37.327	26.557	26.612	0.583	11.919	11.867	19.065
Chem-BERT-6L	1.277	0.889	36.779	35.954	36.853	26.553	26.583	0.583	11.887	11.882	18.924
Chem-BERT-8L	1.290	0.887	36.737	35.725	36.862	26.594	26.552	0.582	11.894	11.883	18.901
CHEM-RoBERTa-12L	1.337	0.906	37.431	36.444	37.381	26.671	26.634	0.582	11.898	11.878	19.116
Molformer	1.304	0.898	37.262	36.103	37.371	26.536	26.597	0.583	11.893	11.862	19.041
GIN-R	0.779	0.721	35.136	32.007	35.215	26.727	26.231	0.552	11.798	11.928	18.109
EdgePred	0.748	0.723	35.367	31.874	35.741	26.849	26.630	0.543	11.657	11.320	18.145
ContextPred	0.746	0.721	35.374	32.078	35.355	27.409	26.911	0.543	11.047	10.866	18.105
infomax	0.703	0.719	35.799	33.396	35.739	27.274	27.130	0.540	10.738	10.743	18.278
masking	0.745	0.721	35.604	33.802	36.443	27.499	28.014	0.539	10.836	10.800	18.500
MolCLR	0.748	0.732	34.697	32.272	**35.083**	26.350	26.484	0.553	11.872	11.825	18.062
MoleBERT	0.720	0.715	35.893	34.593	37.110	27.696	27.373	0.549	11.054	11.034	18.674
CGIP-Graph	0.833	0.669	36.427	33.055	36.285	**24.291**	**24.490**	0.541	11.836	11.771	18.020
GraphMVP	0.653	0.666	**34.532**	**31.745**	35.121	26.295	26.539	0.544	11.773	12.108	**17.998**
SchNet	1.031	0.830	36.449	36.212	36.957	27.075	26.859	0.576	11.439	11.464	18.889
EGNN	1.239	0.878	36.260	35.416	37.008	26.572	26.741	0.583	11.923	11.885	18.851
TFN	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM
SE3_Transformer	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM
PaiNN	1.037	0.818	35.869	34.985	36.468	27.390	27.432	0.667	10.664	10.629	18.596
Uni-Mol-R (1 conf)	1.204	0.844	36.517	34.971	36.851	28.186	27.917	0.582	11.849	11.895	19.082
Uni-Mol (1 conf)	1.200	0.807	36.111	34.295	36.725	28.650	28.009	0.582	11.532	11.578	18.949
ResNet18-I-R	1.105	0.798	37.524	36.284	37.624	27.383	28.096	0.587	10.899	11.057	19.136
ImageMol	1.046	0.824	36.738	36.452	36.875	27.436	26.879	0.589	11.131	11.037	18.901
CGIP-Image	1.026	0.806	37.396	35.472	36.816	27.184	26.762	0.575	11.020	10.913	18.797
MaskMol	1.239	0.822	36.689	35.752	36.899	26.572	26.585	0.583	11.670	11.807	18.862
IEM-I	1.059	0.813	38.558	36.142	37.955	27.290	26.979	0.587	10.958	11.065	19.141
ResNet18-G-R	1.149	0.879	37.156	36.673	37.427	27.010	27.011	0.587	12.009	12.243	19.214
IEM-G (1 conf)	1.131	0.838	36.978	36.031	37.440	27.364	27.154	0.590	11.901	11.810	19.124
ViT-G-R	1.301	0.896	35.611	35.991	37.137	26.564	26.600	0.583	11.900	11.877	18.846
VideoMol-G	1.302	0.903	36.874	35.920	37.085	26.564	26.588	0.583	11.899	11.868	18.959
ResNet18-V-R	1.053	0.796	36.607	36.057	37.164	27.163	26.784	0.582	11.709	11.310	18.923
IEM-V	0.994	0.793	36.746	36.019	36.957	28.258	28.066	0.583	11.485	11.987	19.189

Table S38: The standard deviation on 10 macro (M) datasets from StructNet. M#1, M#2, M#3, M#4, M#5, M#6, M#7, M#8, M#9, M#10 represent CHEMBL4420282_IC50, CHEMBL4419606_IC50, CHEMBL4420281_In, CHEMBL3881498_In, CHEMBL4419605_In, CHEMBL4420271_In, CHEMBL4419595_In, CHEMBL3881499_IC50, CHEMBL4420273_In, CHEMBL4419597_In, respectively. OOM represents the Exception of Out Of Memory (OOM) on single GPU of RTX 4090 Ti (24G).

Models	M#1	M#2	M#3	M#4	M#5	M#6	M#7	M#8	M#9	M#10
mcfp4_2048	0.006	0.010	0.019	0.009	0.017	0.008	0.010	0.008	0.007	0.009
ecfp4_2048	0.010	0.007	0.028	0.008	0.018	0.015	0.011	0.016	0.003	0.004
maccs	0.004	0.002	0.025	0.013	0.014	0.015	0.032	0.007	0.003	0.004
physchem	0.001	0.002	0.017	0.008	0.020	0.010	0.010	0.002	0.013	0.012
atompair_2048	0.004	0.013	0.007	0.016	0.006	0.047	0.005	0.003	0.016	0.022
rdkDes	0.003	0.000	0.026	0.018	0.025	0.017	0.015	0.000	0.011	0.015
BERT-6L	0.056	0.025	0.514	0.331	0.363	0.068	0.056	0.002	0.020	0.064
BERT-8L	0.045	0.025	0.496	0.320	0.352	0.146	0.053	0.003	0.046	0.031
RoBERTa-12L	0.015	0.009	0.599	0.752	0.569	0.207	0.099	0.002	0.025	0.018
molformer-R	0.019	0.009	0.513	0.820	0.624	0.057	0.093	0.002	0.016	0.020
Chem-BERT-6L	0.061	0.017	0.425	0.337	0.339	0.065	0.060	0.002	0.027	0.025
Chem-BERT-8L	0.058	0.013	0.364	0.346	0.320	0.124	0.044	0.002	0.038	0.031
CHEM-RoBERTa-12L	0.022	0.008	0.642	0.650	0.550	0.284	0.090	0.002	0.037	0.018
Molformer	0.064	0.018	0.501	0.567	0.512	0.043	0.076	0.003	0.051	0.036
GIN-R	0.054	0.019	1.238	1.170	1.317	0.685	1.049	0.020	0.439	0.491
EdgePred	0.051	0.037	1.405	1.179	1.225	0.446	0.774	0.013	0.859	0.390
ContextPred	0.030	0.032	1.071	1.146	0.721	1.136	0.778	0.007	0.206	0.118
infomax	0.023	0.020	1.333	2.169	0.763	0.955	0.588	0.012	0.118	0.146
masking	0.035	0.020	1.377	1.143	0.858	1.004	1.364	0.008	0.250	0.236
MolCLR	0.031	0.030	1.148	1.466	1.021	0.608	1.087	0.009	0.634	0.387
MoleBERT	0.024	0.036	0.785	2.217	0.208	0.983	0.999	0.012	0.372	0.306
CGIP-Graph	0.030	0.018	0.971	2.610	0.898	0.328	0.684	0.015	0.624	0.789
GraphMVP	0.010	0.024	1.335	0.634	0.642	1.095	0.554	0.011	0.485	0.773
SchNet	0.049	0.024	0.318	0.924	0.237	0.319	0.412	0.003	0.336	0.412
EGNN	0.055	0.046	0.311	0.984	0.490	0.071	0.521	0.001	0.061	0.093
TFN	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM
SE3_Transformer	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM	OOM
PaiNN	0.137	0.074	0.462	0.869	0.328	0.346	0.158	0.039	0.174	0.253
Uni-Mol-R (1 conf)	0.034	0.023	0.264	0.943	0.434	0.840	1.233	0.001	0.097	0.029
Uni-Mol (1 conf)	0.082	0.021	0.616	0.706	0.429	0.499	1.006	0.002	0.397	0.315
ResNet18-I-R	0.103	0.033	1.086	1.646	0.763	0.727	0.773	0.012	0.316	0.467
ImageMol	0.100	0.043	1.841	1.757	1.742	1.004	1.180	0.030	0.373	0.696
CGIP-Image	0.062	0.055	1.532	0.893	1.185	0.890	0.833	0.020	0.594	0.591
MaskMol	0.069	0.038	0.462	0.285	0.304	0.071	0.056	0.002	0.302	0.200
IEM-I	0.084	0.040	0.810	2.070	0.928	1.225	0.823	0.016	0.374	0.470
ResNet18-G-R	0.092	0.037	0.565	1.196	0.565	0.444	0.648	0.009	0.638	0.483
IEM-G (1 conf)	0.073	0.029	1.173	1.450	0.720	0.948	0.665	0.020	0.595	0.336
ViT-G-R	0.061	0.017	0.082	0.379	0.344	0.054	0.047	0.003	0.019	0.022
VideoMol-G	0.053	0.024	0.541	0.467	0.437	0.054	0.068	0.002	0.050	0.032
ResNet18-V-R	0.051	0.022	0.567	0.846	0.585	0.523	0.373	0.005	0.372	0.477
IEM-V	0.039	0.033	0.820	1.774	0.970	0.814	0.891	0.010	0.524	0.647

Table S39: The average RMSE performance on 10 macrocyclic peptide (MP) datasets from StructNet. MP#1, MP#2, MP#3, MP#4, MP#5, MP#6, MP#7, MP#8, MP#9, MP#10 represent CHEMBL4888485_Inhibition, CHEMBL2354301_AC50, CHEMBL3880198_Ki, CHEMBL4420271_Inhibition, CHEMBL4419595_Inhibition, CHEMBL4420282_IC50, CHEMBL3214979_AC50, CHEMBL4420277_Inhibition, CHEMBL4419601_Inhibition, CHEMBL4419606_IC50, respectively. OOM represents the Exception of Out Of Memory (OOM) on single GPU of RTX 4090 Ti (24G). The green background represents top-5 performance.

Models	MP#1	MP#2	MP#3	MP#4	MP#5	MP#6	MP#7	MP#8	MP#9	MP#10	Mean
mcfp4_2048	10.134	0.405	1.012	25.835	25.835	0.838	0.293	15.057	15.057	0.970	9.544
ecfp4_2048	10.227	0.624	1.055	26.031	26.031	0.874	0.467	15.113	15.113	1.020	9.656
maccs	10.172	0.374	0.923	25.612	25.612	0.904	0.274	14.935	14.935	1.001	9.474
physchem	10.157	0.374	1.037	29.923	29.923	0.964	0.564	16.133	16.133	0.998	10.621
atompair_2048	10.034	0.422	0.921	25.458	25.458	0.911	0.278	14.918	14.918	1.041	9.436
rdkDes	10.169	0.381	1.044	32.574	32.574	1.050	0.933	17.745	17.745	1.085	11.530
BERT-6L	10.129	0.371	0.866	25.152	25.152	0.963	0.271	15.017	15.017	0.976	9.391
BERT-8L	10.124	0.371	0.866	25.132	25.132	0.963	0.272	15.005	15.005	0.980	9.385
RoBERTa-12L	10.124	0.371	0.873	24.697	24.697	0.963	0.270	15.522	15.522	0.942	9.398
molformer-R	10.130	0.371	0.866	24.772	24.772	0.963	0.274	15.571	15.571	0.955	9.424
Chem-BERT-6L	10.130	0.371	0.863	25.203	25.203	0.963	0.272	15.009	15.009	0.985	9.401
Chem-BERT-8L	10.128	0.371	0.872	25.183	25.183	0.963	0.273	15.050	15.050	0.957	9.403
CHEM-RoBERTa-12L	10.120	0.371	0.872	25.312	25.312	0.963	0.267	15.500	15.500	0.934	9.515
Molformer	10.130	0.371	0.865	25.667	25.667	0.963	0.268	15.038	15.038	0.948	9.496
GIN-R	10.526	0.385	0.771	27.702	27.007	1.191	**0.261**	19.136	21.544	1.034	10.956
EdgePred	10.410	0.359	0.774	25.692	25.676	0.886	0.265	14.951	14.917	0.956	9.489
ContextPred	10.156	0.366	0.791	25.773	25.807	0.814	0.273	14.912	14.923	0.955	9.477
infomax	10.070	**0.340**	0.823	24.658	24.503	0.817	0.286	14.716	**14.612**	0.946	**9.177**
masking	10.241	0.359	0.793	25.620	25.734	0.841	0.276	14.906	14.905	0.933	9.461
MolCLR	10.407	0.347	0.768	25.422	25.268	0.854	**0.261**	15.241	15.083	0.954	9.461
MoleBERT	10.059	0.352	0.863	25.321	25.316	**0.806**	0.284	14.893	14.895	0.937	9.373
CGIP-Graph	10.297	0.400	0.802	25.667	25.939	0.867	0.274	**14.617**	14.627	0.890	9.438
GraphMVP	10.454	0.385	0.789	25.686	25.381	0.864	0.264	15.423	15.122	0.990	9.536
SchNet	10.414	0.399	0.881	25.031	25.274	0.989	0.302	15.002	14.996	0.955	9.424
EGNN	10.117	0.371	0.877	26.386	26.386	0.963	0.272	15.245	15.245	0.929	9.679
TFN	10.106	0.364	0.868	OOM	OOM	OOM	0.363	OOM	OOM	OOM	-
SE3_Transformer	10.370	0.341	**0.742**	OOM	OOM	OOM	0.320	OOM	OOM	OOM	-
PaiNN	10.626	0.434	0.909	25.160	25.260	1.234	0.495	14.880	14.897	1.178	9.507
Uni-Mol-R (1 conf)	10.133	0.371	0.857	25.031	25.031	0.963	0.272	15.572	15.572	0.958	9.476
Uni-Mol (1 conf)	10.134	0.369	0.864	24.991	24.991	0.963	0.273	15.457	15.457	0.955	9.445
ResNet18-I-R	10.024	0.377	0.898	24.988	24.988	0.963	0.286	14.862	14.862	0.964	9.321
ImageMol	10.373	0.378	0.846	24.654	24.654	0.993	0.286	15.471	15.471	0.917	9.404
CGIP-Image	10.222	0.370	0.877	24.718	24.718	0.962	0.284	15.036	15.036	**0.857**	9.308
MaskMol	10.124	0.371	0.860	24.771	24.771	0.963	0.271	15.005	15.005	0.979	9.312
IEM-I	10.155	0.373	0.864	**24.413**	**24.413**	0.961	0.278	15.411	15.411	0.906	9.319
ResNet18-G-R	10.075	0.375	0.882	25.039	24.956	0.971	0.275	14.958	14.939	0.959	9.343
IEM-G (1 conf)	10.139	0.377	0.910	25.727	25.148	0.997	0.278	15.290	15.137	0.969	9.497
ViT-G-R	10.120	0.371	0.869	25.615	25.637	0.963	0.268	15.121	15.151	0.961	9.508
VideoMol-G	10.133	0.372	0.870	24.677	24.677	0.962	0.273	15.649	15.607	0.968	9.419
ResNet18-V-R	**9.968**	0.373	0.871	24.936	25.080	0.965	0.274	14.978	14.974	0.966	9.339
IEM-V	10.086	0.380	0.946	25.360	25.621	1.007	0.282	15.320	15.463	0.995	9.546

Table S40: The standard deviation on 10 macrocyclic peptide (MP) datasets from StructNet. MP#1, MP#2, MP#3, MP#4, MP#5, MP#6, MP#7, MP#8, MP#9, MP#10 represent CHEMBL4888485_Inhibition, CHEMBL2354301_AC50, CHEMBL3880198_Ki, CHEMBL4420271_Inhibition, CHEMBL4419595_Inhibition, CHEMBL4420282_IC50, CHEMBL3214979_AC50, CHEMBL4420277_Inhibition, CHEMBL4419601_Inhibition, CHEMBL4419606_IC50, respectively. OOM represents the Exception of Out Of Memory (OOM) on single GPU of RTX 4090 Ti (24G).

Models	MP#1	MP#2	MP#3	MP#4	MP#5	MP#6	MP#7	MP#8	MP#9	MP#10
mcfp4_2048	0.015	0.005	0.014	0.057	0.057	0.002	0.002	0.003	0.003	0.008
ecfp4_2048	0.012	0.026	0.018	0.008	0.008	0.001	0.004	0.005	0.005	0.003
maccs	0.008	0.007	0.014	0.042	0.042	0.004	0.004	0.006	0.006	0.017
physchem	0.006	0.001	0.005	0.050	0.050	0.000	0.043	0.035	0.035	0.012
atompair_2048	0.037	0.010	0.006	0.012	0.012	0.001	0.009	0.007	0.007	0.008
rdkDes	0.002	0.000	0.004	0.026	0.026	0.011	0.033	0.022	0.022	0.018
BERT-6L	0.006	0.000	0.012	0.189	0.189	0.000	0.003	0.110	0.110	0.038
BERT-8L	0.014	0.000	0.018	0.198	0.198	0.000	0.004	0.136	0.136	0.035
RoBERTa-12L	0.006	0.001	0.022	0.020	0.020	0.000	0.017	0.357	0.357	0.033
molformer-R	0.007	0.001	0.011	0.038	0.038	0.000	0.011	0.368	0.368	0.033
Chem-BERT-6L	0.007	0.001	0.013	0.170	0.170	0.000	0.003	0.106	0.106	0.019
Chem-BERT-8L	0.003	0.000	0.013	0.241	0.241	0.000	0.005	0.129	0.129	0.040
CHEM-RoBERTa-12L	0.010	0.001	0.011	1.380	1.380	0.000	0.015	0.471	0.471	0.051
Molformer	0.005	0.001	0.011	0.490	0.490	0.000	0.008	0.115	0.115	0.023
GIN-R	0.366	0.030	0.077	1.631	1.546	0.153	0.014	3.520	2.601	0.039
EdgePred	0.604	0.010	0.027	0.178	0.163	0.113	0.013	0.049	0.030	0.060
ContextPred	0.094	0.014	0.032	0.082	0.040	0.021	0.008	0.026	0.026	0.062
infomax	0.085	0.010	0.067	0.364	0.707	0.038	0.011	0.329	0.289	0.061
masking	0.458	0.013	0.084	0.929	0.170	0.043	0.014	0.025	0.020	0.038
MolCLR	0.218	0.016	0.070	0.542	0.314	0.062	0.005	0.559	0.429	0.038
MoleBERT	0.111	0.014	0.062	0.238	0.245	0.010	0.011	0.017	0.018	0.027
CGIP-Graph	0.429	0.027	0.087	1.038	0.644	0.038	0.013	0.335	0.151	0.067
GraphMVP	0.345	0.012	0.067	0.957	0.812	0.057	0.016	0.924	0.550	0.057
SchNet	0.309	0.006	0.083	0.635	0.814	0.025	0.008	0.184	0.228	0.028
EGNN	0.141	0.001	0.047	1.582	1.582	0.000	0.004	0.430	0.430	0.046
TFN	0.161	0.023	0.128	OOM	OOM	OOM	0.032	OOM	OOM	OOM
SE3_Transformer	0.188	0.018	0.054	OOM	OOM	OOM	0.013	OOM	OOM	OOM
PaiNN	0.225	0.018	0.063	0.413	0.467	0.311	0.079	0.224	0.206	0.315
Uni-Mol-R (1 conf)	0.010	0.001	0.028	0.054	0.054	0.000	0.003	0.136	0.136	0.033
Uni-Mol (1 conf)	0.018	0.003	0.039	0.063	0.063	0.000	0.005	0.262	0.262	0.040
ResNet18-I-R	0.272	0.009	0.034	0.310	0.310	0.012	0.017	0.083	0.083	0.033
ImageMol	0.489	0.020	0.066	1.230	1.230	0.075	0.014	0.655	0.655	0.092
CGIP-Image	0.278	0.007	0.060	0.831	0.831	0.035	0.020	0.760	0.760	0.022
MaskMol	0.018	0.002	0.016	0.098	0.098	0.001	0.004	0.119	0.119	0.034
IEM-I	0.288	0.010	0.055	1.126	1.126	0.032	0.010	0.564	0.564	0.048
ResNet18-G-R	0.073	0.008	0.036	0.235	0.296	0.011	0.010	0.112	0.150	0.035
IEM-G (1 conf)	0.199	0.011	0.064	0.973	0.447	0.032	0.023	0.518	0.425	0.049
ViT-G-R	0.055	0.002	0.015	0.412	0.514	0.000	0.006	0.080	0.099	0.039
VideoMol-G	0.034	0.002	0.018	0.007	0.007	0.002	0.007	0.219	0.238	0.038
ResNet18-V-R	0.325	0.007	0.022	0.397	0.527	0.006	0.009	0.121	0.136	0.041
IEM-V	0.320	0.015	0.094	0.420	0.566	0.032	0.014	0.457	1.033	0.050

Table S41: The average RMSE performance on 10 reticular (R) datasets from StructNet. R#1, R#2, R#3, R#4, R#5, R#6, R#7, R#8, R#9, R#10 represent CHEMBL4888485_In, CHEMBL1614458_P, CHEMBL1614459_P, CHEMBL1613914_P, CHEMBL1614421_P, CHEMBL1614087_P, CHEMBL1614249_P, CHEMBL1614236_P, CHEMBL1614544_P, CHEMBL1614038_P, respectively. The green background represents top-5 performance.

Models	R#1	R#2	R#3	R#4	R#5	R#6	R#7	R#8	R#9	R#10	Mean
mcfp4_2048	19.359	0.539	0.903	0.585	0.644	0.880	0.597	0.525	0.947	0.578	2.556
ecfp4_2048	19.335	0.606	0.965	0.615	0.719	0.924	0.651	0.641	**0.939**	0.730	2.613
maccs	19.325	0.443	0.723	0.472	0.637	0.857	0.569	**0.438**	0.981	0.536	2.498
physchem	19.352	0.440	0.695	0.409	0.601	**0.807**	0.433	0.496	0.956	0.589	2.478
atompair_2048	19.358	0.557	0.907	0.734	0.675	0.961	0.649	0.574	1.039	0.559	2.601
rdkDes	19.338	0.511	0.692	**0.386**	0.670	0.823	0.652	0.664	1.004	0.895	2.564
BERT-6L	19.700	0.438	0.664	0.396	0.590	**0.807**	0.388	0.451	0.999	0.429	2.486
BERT-8L	19.702	0.439	0.664	0.396	0.590	**0.807**	**0.387**	0.451	1.000	0.429	2.487
RoBERTa-12L	19.696	0.430	0.664	0.396	0.589	**0.807**	**0.387**	0.452	0.999	0.433	2.485
molformer-R	19.691	0.434	0.664	0.397	0.590	**0.807**	0.388	0.450	0.999	0.428	2.485
Chem-BERT-6L	19.665	0.438	0.664	0.395	0.591	**0.807**	**0.387**	0.451	1.001	0.429	2.483
Chem-BERT-8L	19.697	0.437	0.664	0.396	0.589	**0.807**	**0.387**	0.451	0.999	0.429	2.486
CHEM-RoBERTa-12L	19.701	0.425	0.664	0.397	0.590	**0.807**	0.392	0.452	0.997	0.432	2.486
Molformer	19.688	0.438	0.664	0.396	0.589	**0.807**	0.388	0.450	1.001	0.429	2.485
GIN-R	19.678	0.437	0.672	0.432	0.595	0.829	0.397	0.462	1.008	0.449	2.496
EdgePred	19.410	0.426	0.641	0.395	0.587	0.829	0.416	0.457	1.011	0.452	2.462
ContextPred	19.442	0.458	0.667	0.407	0.581	0.841	0.394	0.449	1.035	**0.407**	2.468
infomax	19.379	0.450	0.684	0.427	0.593	0.871	0.435	0.496	1.064	0.447	2.485
masking	19.478	0.450	0.712	0.393	0.572	0.855	0.411	0.468	1.058	0.452	2.485
MolCLR	19.542	0.437	0.656	0.416	0.591	0.822	0.397	0.465	1.014	0.425	2.477
MoleBERT	19.360	0.470	0.716	0.416	0.572	0.831	0.462	0.466	1.032	0.458	2.478
CGIP-Graph	**19.240**	0.493	**0.633**	0.410	0.585	0.841	0.445	0.479	1.051	0.443	2.462
GraphMVP	19.271	0.472	0.666	0.422	0.586	0.839	0.406	0.476	1.026	0.439	2.460
SchNet	19.639	0.459	0.719	0.432	0.575	0.874	0.400	0.478	0.987	0.425	2.499
EGNN	19.549	0.432	0.665	0.397	0.590	**0.807**	**0.387**	0.451	0.999	0.432	2.471
TFN	19.576	0.511	0.716	0.477	0.613	0.911	0.476	0.552	1.097	0.519	2.545
SE3_Transformer	19.644	0.457	0.691	0.536	**0.547**	0.866	0.480	0.535	1.069	0.491	2.532
PaiNN	19.317	1.385	1.393	0.978	1.160	1.377	0.962	1.326	1.594	0.888	3.038
Uni-Mol-R (1 conf)	19.565	0.431	0.664	0.396	0.590	**0.807**	0.388	0.450	1.001	0.428	2.472
Uni-Mol (1 conf)	19.529	0.426	0.665	0.398	0.591	0.808	0.395	0.451	1.001	0.428	2.469
ResNet18-I-R	19.346	0.429	0.672	0.397	0.585	0.817	0.394	0.456	1.010	0.430	**2.454**
ImageMol	19.618	0.447	0.751	0.423	0.605	0.835	0.431	0.487	1.052	0.459	2.511
CGIP-Image	19.459	0.425	0.679	0.408	0.596	0.818	0.393	0.471	0.999	0.437	2.468
MaskMol	19.660	0.430	0.664	0.396	0.588	**0.807**	0.388	0.450	1.000	0.426	2.481
IEM-I	19.474	0.440	0.688	0.401	0.591	0.816	0.394	0.455	1.005	0.432	2.470
ResNet18-G-R	19.500	0.430	0.663	0.403	0.589	0.824	0.393	0.458	0.987	0.426	2.467
IEM-G (1 conf)	19.796	0.433	0.676	0.413	0.590	0.843	0.397	0.467	1.010	0.445	2.507
ViT-G-R	19.654	0.436	0.664	0.397	0.585	**0.807**	**0.387**	0.450	0.998	0.422	2.480
VideoMol-G	19.667	0.434	0.665	0.397	0.589	0.809	0.390	0.451	0.999	0.428	2.483
ResNet18-V-R	19.529	**0.424**	0.676	0.395	0.585	0.822	0.390	0.457	0.984	0.430	2.469
IEM-V	19.726	0.429	0.672	0.414	0.581	0.839	0.401	0.471	1.017	0.464	2.501

Table S42: The standard deviation on 10 reticular (R) datasets from StructNet. R#1, R#2, R#3, R#4, R#5, R#6, R#7, R#8, R#9, R#10 represent CHEMBL4888485_In, CHEMBL1614458_P, CHEMBL1614459_P, CHEMBL1613914_P, CHEMBL1614421_P, CHEMBL1614087_P, CHEMBL1614249_P, CHEMBL1614236_P, CHEMBL1614544_P, CHEMBL1614038_P, respectively.

Models	R#1	R#2	R#3	R#4	R#5	R#6	R#7	R#8	R#9	R#10
mcfp4_2048	0.002	0.002	0.003	0.004	0.005	0.002	0.003	0.003	0.002	0.002
ecfp4_2048	0.002	0.004	0.005	0.005	0.005	0.002	0.003	0.003	0.004	0.002
maccs	0.034	0.004	0.006	0.003	0.003	0.005	0.005	0.002	0.004	0.003
physchem	0.008	0.005	0.001	0.002	0.005	0.001	0.017	0.015	0.001	0.037
atompair_2048	0.006	0.003	0.004	0.007	0.002	0.003	0.003	0.003	0.003	0.002
rdkDes	0.004	0.004	0.001	0.000	0.011	0.005	0.022	0.020	0.009	0.031
BERT-6L	0.093	0.002	0.001	0.002	0.001	0.002	0.001	0.002	0.003	0.003
BERT-8L	0.065	0.004	0.002	0.003	0.002	0.002	0.001	0.001	0.005	0.003
RoBERTa-12L	0.030	0.015	0.001	0.004	0.003	0.002	0.003	0.005	0.010	0.007
molformer-R	0.040	0.006	0.001	0.004	0.003	0.004	0.006	0.003	0.010	0.006
Chem-BERT-6L	0.058	0.005	0.001	0.003	0.002	0.001	0.002	0.002	0.003	0.002
Chem-BERT-8L	0.046	0.007	0.003	0.001	0.002	0.002	0.002	0.001	0.005	0.004
CHEM-RoBERTa-12L	0.020	0.008	0.002	0.004	0.004	0.001	0.014	0.005	0.006	0.007
Molformer	0.060	0.010	0.001	0.003	0.001	0.003	0.003	0.003	0.007	0.004
GIN-R	0.330	0.031	0.044	0.017	0.026	0.020	0.010	0.026	0.045	0.052
EdgePred	0.160	0.024	0.032	0.012	0.014	0.016	0.028	0.017	0.021	0.024
ContextPred	0.063	0.035	0.075	0.016	0.022	0.009	0.007	0.012	0.014	0.013
infomax	0.202	0.022	0.090	0.016	0.029	0.017	0.021	0.023	0.037	0.013
masking	0.079	0.030	0.100	0.006	0.007	0.011	0.008	0.018	0.024	0.010
MolCLR	0.123	0.060	0.018	0.010	0.016	0.007	0.015	0.021	0.030	0.021
MoleBERT	0.056	0.039	0.038	0.025	0.013	0.026	0.026	0.014	0.030	0.023
CGIP-Graph	0.200	0.053	0.024	0.023	0.026	0.021	0.025	0.015	0.049	0.029
GraphMVP	0.239	0.075	0.033	0.019	0.037	0.027	0.022	0.014	0.037	0.026
SchNet	0.191	0.024	0.057	0.037	0.006	0.039	0.009	0.012	0.024	0.008
EGNN	0.404	0.011	0.001	0.003	0.002	0.001	0.002	0.003	0.006	0.003
TFN	0.148	0.086	0.051	0.024	0.026	0.031	0.048	0.038	0.069	0.042
SE3_Transformer	0.194	0.040	0.053	0.060	0.025	0.028	0.036	0.049	0.069	0.038
PaiNN	0.197	0.124	0.141	0.104	0.050	0.131	0.047	0.113	0.115	0.042
Uni-Mol-R (1 conf)	0.121	0.007	0.001	0.002	0.002	0.003	0.002	0.002	0.002	0.003
Uni-Mol (1 conf)	0.088	0.010	0.004	0.009	0.004	0.001	0.006	0.004	0.007	0.004
ResNet18-I-R	0.173	0.013	0.021	0.014	0.015	0.012	0.010	0.012	0.027	0.018
ImageMol	0.533	0.036	0.094	0.021	0.041	0.022	0.032	0.047	0.052	0.028
CGIP-Image	0.126	0.012	0.017	0.018	0.011	0.012	0.013	0.016	0.031	0.015
MaskMol	0.066	0.007	0.004	0.005	0.003	0.002	0.003	0.003	0.006	0.003
IEM-I	0.137	0.033	0.030	0.017	0.009	0.017	0.019	0.010	0.035	0.019
ResNet18-G-R	0.128	0.021	0.016	0.012	0.008	0.009	0.013	0.012	0.023	0.011
IEM-G (1 conf)	0.255	0.020	0.023	0.016	0.024	0.036	0.012	0.021	0.035	0.024
ViT-G-R	0.046	0.009	0.001	0.004	0.008	0.005	0.002	0.003	0.013	0.008
VideoMol-G	0.065	0.019	0.002	0.005	0.010	0.004	0.003	0.005	0.008	0.011
ResNet18-V-R	0.171	0.016	0.007	0.006	0.005	0.008	0.006	0.009	0.016	0.014
IEM-V	0.241	0.023	0.012	0.019	0.012	0.016	0.017	0.017	0.033	0.031

Table S43: Parameters and computational costs of different models with a batch size of 8. "time cost (30 epochs)" represents the total time required to train, evaluate, and test the model for 30 epochs on 10,000 molecules. The number of molecules for training, evaluation, and testing are 8,000, 1,000, and 1,000, respectively. "params" represents the number of parameters of the model.

Modality	model	time cost (30 epochs)	params
SMILES	CHEM-BERT-no-pretrain	46.391 min	38.397M
	CHEM-BERT	47.918 min	38.397M
	CHEM-RoBERTa	98.599 min	85.495M
	CHEM-RoBERTa-no-pretrain	86.776 min	85.495M
	molformer-no-pretrain	41.306 min	25.511M
	CHEM-BERT-origin-no-pretrain	96.216 min	51.001M
	CHEM-BERT-origin	51.751 min	51.001M
	molformer	39.692 min	25.511M
Graph	GIN_RANDOM	20.487 min	1.862M
	EdgePred	20.780 min	1.862M
	ContextPred	20.277 min	1.862M
	infomax	20.425 min	1.862M
	masking	13.720 min	1.862M
	MolCLR	20.795 min	1.862M
	MoleBERT	20.827 min	1.862M
	CGIP-Graph	25.292 min	3.793M
	GraphMVP	20.698 min	1.862M
Geometry graph	Uni-Mol-R (1 conf)	35.763 min	47.600M
	Uni-Mol (1 conf)	73.444 min	47.600M
	TFN	273.258 min	8.663M
	SE3_Transformer	257.265 min	10.126M
Image	ResNet18-I-R	26.215 min	11.183M
	ImageMol	26.479 min	11.183M
	CGIP-Image	27.817 min	11.183M
	MaskMol	54.007 min	85.808M
	IEM-I	25.387 min	11.183M
Geometry Image	ResNet18-G-R	89.537 min	11.183M
	IEM-G (1 conf)	84.701 min	11.183M
Video	ResNet18-V-R	750.935 min	11.183M
	IEM-V	934.883 min	11.183M

Table S44: The time required to train the model for 1 epoch using different numbers of frames. The trained model is ResNet18-V-R. "time cost (1 epoch)" is the total time taken to train on 8,000 molecules and evaluate on 2,000 molecules.

#Frame	time cost (1 epoch)
1	2.024 min
3	2.786 min
5	4.169 min
10	5.934 min
30	13.836 min
60	25.849 min

Table S45: Time required for the model to virtually screen 10,000 molecules.

Modality	model	inference time (1,000 molecules)
SMILES	CHEM-BERT-no-pretrain	2.301 s
	CHEM-BERT	2.189 s
	CHEM-RoBERTa	3.367 s
	CHEM-RoBERTa-no-pretrain	3.315 s
	molformer-no-pretrain	3.489 s
	CHEM-BERT-origin-no-pretrain	2.652 s
	CHEM-BERT-origin	2.341 s
	molformer	3.441 s
Graph	GIN_RANDOM	2.013 s
	EdgePred	2.241 s
	ContextPred	2.097 s
	infomax	1.822 s
	masking	2.254 s
	MolCLR	1.633 s
	MoleBERT	2.401 s
	CGIP-Graph	1.884 s
	GraphMVP	1.727 s
Geometry graph	Uni-Mol-R (1 conf)	3.301 s
	Uni-Mol (1 conf)	2.924 s
	TFN	6.372 s
	SE3_Transformer	10.180 s
Image	ResNet18-I-R	4.151 s
	ImageMol	2.106 s
	CGIP-Image	2.111 s
	MaskMol	2.600 s
	IEM-I	2.147 s
Geometry Image	ResNet18-G-R	5.769 s
	IEM-G (1 conf)	5.618 s
Video	ResNet18-V-R	73.419 s
	IEM-V	73.163 s

Table S46: The RMSE in prediction results between different modalities on HIV dataset. The larger the RMSE, the greater the difference between the two modes. Green indicates the top three with the greatest differences, and bold indicates the greatest difference.

	fingerprint	sequence	graph	geometry_graph	image	geometry_image	video
fingerprint	0	0.093046	0.100329	0.067156	0.058782	0.102035	**0.177339**
sequence	0.093046	0	0.110071	0.089303	0.103196	0.102689	**0.157157**
graph	0.100329	0.110071	0	0.108321	0.117101	0.125993	**0.176838**
geometry_graph	0.067156	0.089303	0.108321	0	0.05556	0.091146	**0.166107**
image	0.058782	0.103196	0.117101	0.05556	0	0.098585	**0.182885**
geometry_image	0.102035	0.102689	0.125993	0.091146	0.098585	0	**0.153324**
video	0.177339	0.157157	0.176838	0.166107	**0.182885**	0.153324	0

Table S47: The Pearson correlation coefficient in prediction results between different modalities on HIV dataset. The smaller the Pearson correlation coefficient, the greater the difference between the two modes. Green indicates the top three with the greatest differences, and bold indicates the greatest difference.

	fingerprint	sequence	graph	geometry_graph	image	geometry_image	video
fingerprint	1	0.58403	0.606879	0.519876	0.448639	**0.387169**	0.410779
sequence	0.58403	1	0.576967	0.650448	**0.556317**	0.572682	0.579381
graph	0.606879	0.576967	1	0.533092	0.450626	**0.404815**	0.441554
geometry_graph	**0.519876**	0.650448	0.533092	1	0.660593	0.619167	0.58209
image	**0.448639**	0.556317	0.450626	0.660593	1	0.564617	0.516407
geometry_image	**0.387169**	0.572682	0.404815	0.619167	0.564617	1	0.601794
video	**0.410779**	0.579381	0.441554	0.58209	0.516407	0.601794	1

Given the differences in predicted logits between multi-modal molecules, we further conduct an exploratory experiment to find the upper limit of multi-modal fusion. We assume that the test set labels have been obtained. We generate the final prediction logits by determining the minimum difference between the prediction logits of different modalities and the true results. That is, for a certain molecule, the prediction logit of which modality is closest to its true result, we assign this prediction logit to this molecule. Finally, we obtained a ROC-AUC result of 99.7% on the HIV test set. This strong improvement means that **building a routing network or ranking algorithm for selecting results based on models of different molecular modalities is a promising direction**. We also show the contribution of different modalities in Figure S4, which suggests that incorporating video modality into multi-modal representation learning of molecules is promising in the future.

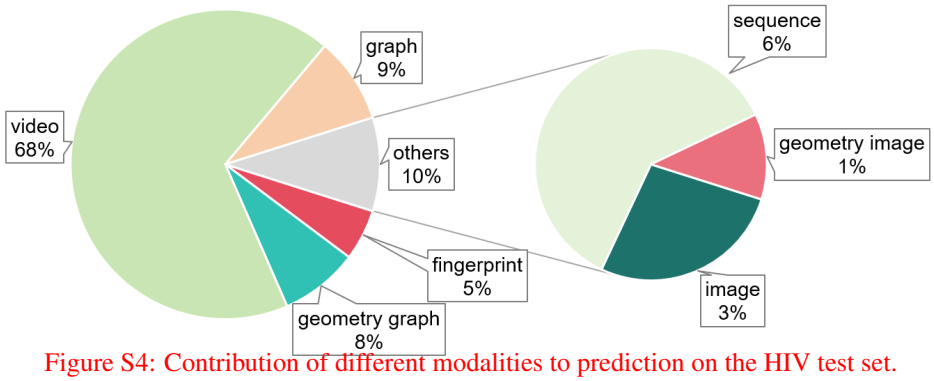

Figure S4: Contribution of different modalities to prediction on the HIV test set.

K.2 Why Do Uni-Mol Features Allow For The High Performance in MoleculeNet?

We speculate that the high performance of Uni-Mol benefits from its pre-training on 209 M molecular conformations. To verify this speculation, we pre-trained Uni-Mol using 50K, 200K, and 2M molecular conformations respectively and observed their performance on 8 classification datasets from MoleculeNet. In details, we use the official code and parameters provided by Uni-Mol [2], and pretrain with the first 50K, 200K and 2M conformations of pcqm4m-v2-train [3]. The experimental settings for linear probing on MoleculeNet are consistent with our paper. As shown in Table S48, we find that the amount of pre-training data is very important for Uni-Mol to achieve good performance on MoleculeNet.

Table S48: ROC-AUC performance of Uni-Mol pre-trained with different data sizes on 8 classification tasks from MoleculeNet with linear probing and 10 different run seeds. In linear probing, 10 conformations per molecule are used. #Conf indicates the amount of data used for pre-training Uni-Mol.

	#Conf	BBBP	Tox21	ToxCast	Sider	ClinTox	MUV	HIV	BACE	Avg
Uni-Mol	50K	68.0±0.2	71.3±0.1	63.1±0.4	60.4±0.2	68.0±3.2	71.3±0.5	78.0±0.4	80.7±0.7	70.1
	200K	65.0±0.9	73.9±0.2	64.0±0.8	62.6±0.3	62.6±1.5	72.4±1.0	76.2±1.0	79.2±0.7	69.5
	2M	67.3±0.2	73.2±0.2	63.7±0.7	60.6±0.3	64.8±1.3	73.9±0.7	77.1±0.3	77.9±0.3	69.8
	209M	69.3±0.6	75.2±0.1	65.8±0.5	61.6±0.4	85.1±4.4	80.3±0.7	77.5±0.1	78.2±0.2	74.1

K.3 Why Multiple Conformations Can Significantly Improve Geometry Image Performance?

Tables S20 and S21 show that multiple conformations can significantly improve the performance of the geometric image modality (such as IEM-G) with little gain for geometric graph modality (such as Uni-Mol). Here, we try to answer why this phenomenon occurs. We select BBBP, Tox21, ClinTox and ToxCast and analyze them in terms of the direction and scale of the features from different

[2] https://github.com/deepmodeling/Uni-Mol/blob/main/unimol/README.md
[3] https://ogb.stanford.edu/docs/lsc/pcqm4mv2/

conformations. We use IEM-G and Uni-Mol to extract features of 10 conformations from these 4 datasets.

- **Direction between features from different conformations.** Cosine similarity represents the similarity between features in direction. As shown in Figure S5, the The lower left corner and upper left corner show the difference between Uni-Mol and IEM-G in cosine similarity, respectively. Obviously, we find that the cosine similarity of IEM-G is lower than that of Uni-Mol, which means that IEM-G has more diversity of direction in extracting features of different conformations. The cosine similarities of Uni-Mol are all above 99%, which means that multiple conformations are difficult to provide additional information.
- **Scale between features from different conformations.** Euclidean distance represents the difference in feature scale. As shown in Figure S6, the lower left corner and upper left corner show the differences between Uni-Mol and IEM-G in Euclidean distance, respectively. It is easy to observe that IEM-G has a larger Euclidean distance than Uni-Mol, which means that IEM-G has a larger difference in feature scale than Uni-Mol. It may bring more degrees of freedom to IEM-G in feature space.

Furthermore, we plot the distribution of cosine similarity and Euclidean distance in Figure S7 . We can see that the distribution of IEM-G is much flatter than that of Uni-Mol. In general, Figures S5, S6, S7 show that **the multi-conformational features extracted by IEM-G have more directional diversity and scale diversity**.

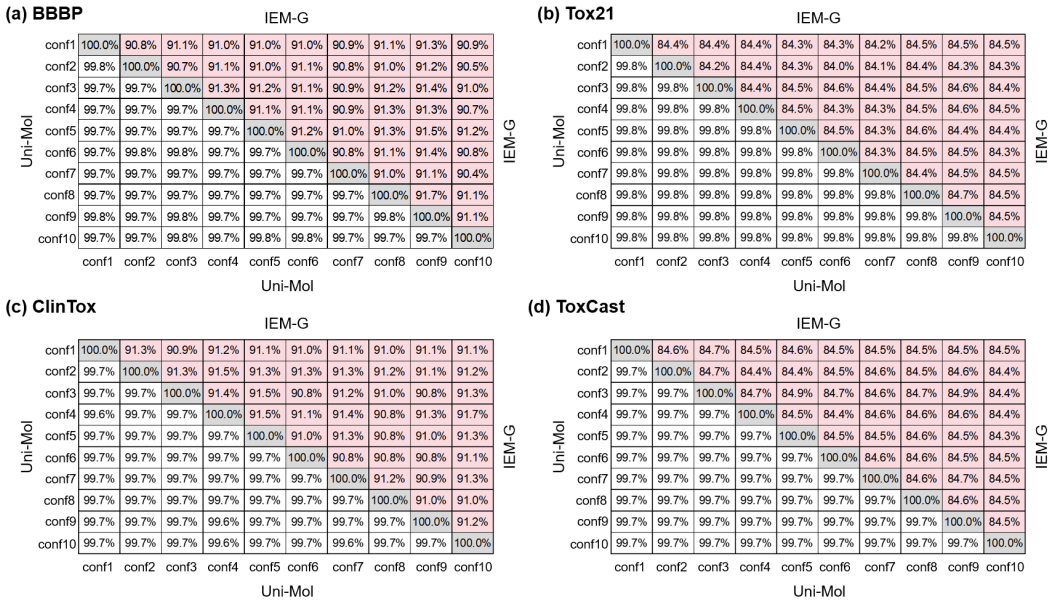

Figure S5: Cosine similarity between different conformations on BBBP, Tox21, ClinTox, and ToxCast. The white background in the lower left corner represents the features of different conformations extracted using Uni-Mol and the cosine similarity calculated, while the pink background in the upper right corner represents those of IEM-G.

K.4 Importance of Number of Layers for Sequence models on MoleculeNet

In the MoleculeNet experiment, we see that even without any pre-training, the features of sequence-based MolFormer-R can still achieve excellent performance on MoleculeNet. Therefore, in order to find out whether there is a correlation between the number of layers of MolFormer-R and its performance, we perform ablation on the number of layers of the MolFormer-R. In MolFormer-R, the default number of layers is 6. Here, we further set the number of layers to 1, 2, 4, and 8. As shown in Table S49, we find that as the number of layers increases, the average performance of the model increases from 69.91% of 1 layer to 71.55%, indicating that it is possible to further improve the performance by increasing the number of layers in the sequence model.

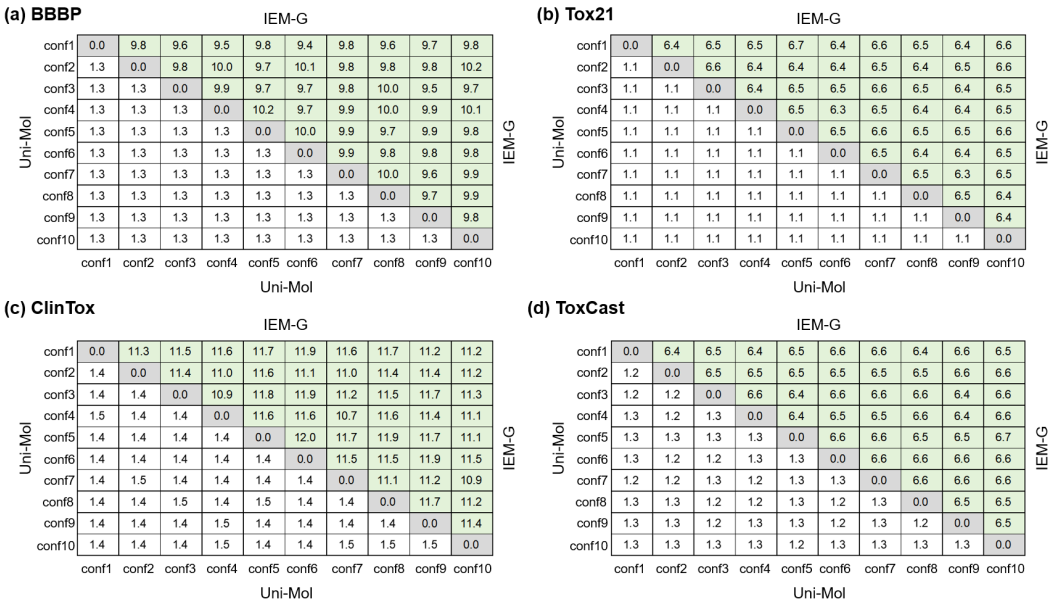

Figure S6: Euclidean distance between different conformations on BBBP, Tox21, ClinTox, and ToxCast. The white background in the lower left corner represents the features of different conformations extracted using Uni-Mol and the cosine similarity calculated, while the green background in the upper right corner represents those of IEM-G. "conf" 1 to "conf 10" refer to 10 different molecular conformations.

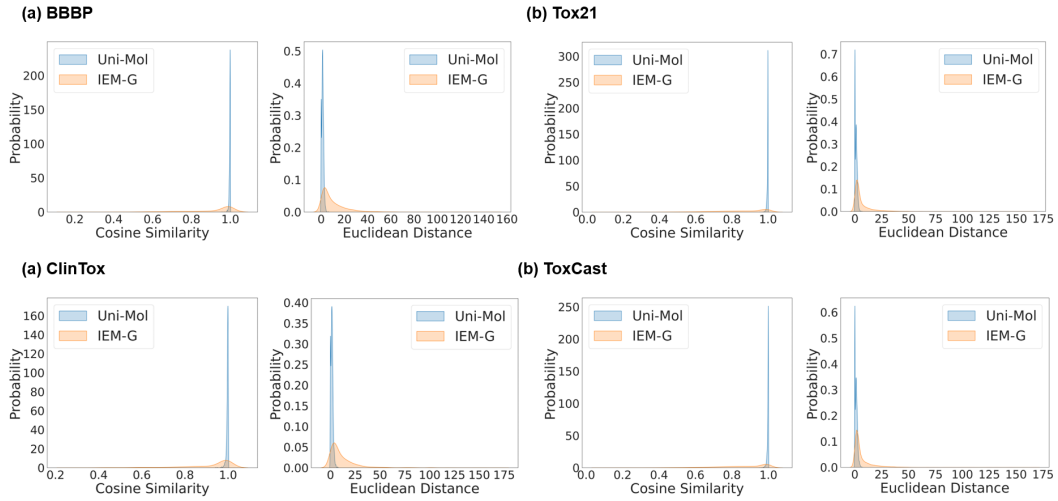

Figure S7: The distribution of cosine similarity and Euclidean distance between different conformations on BBBP, Tox21, ClinTox, and ToxCast datasets with Uni-Mol and IEM-G. The distribution here is calculated by randomly sampling 2 from 10 conformations.

K.5 Why Does The Some Modalities Have Strong Performance Even Without Training?

In the linear probing experiment of MoleculeNet, we find that some modalities (sequence and geometry) can still achieve good performance even without any training, especially the MolFormer-R based on the sequence modality, which achieves an average ROC-AUC of 70.40% on 8 classification tasks. We speculate that this is related to the inductive bias of the modality. Given the excellent performance of the sequence modality without training, we speculate that the model's ability to recognize molecular substructures is the key to its performance.

Table S49: Ablation study of the number of MolFormer-R layers using ROC-AUC metric and 10 runs. #layers indicates the number of layers in the setting.

	#layers	BBBP	Tox21	ToxCast	Sider	ClinTox	MUV	HIV	BACE	Avg
MolFormer-R	1	68.9±0.4	71.0±0.1	61.6±0.4	56.9±0.3	82.6±0.8	68.8±1.7	70.2±0.7	79.3±0.9	69.91
	2	71.5±0.9	71.1±0.1	61.8±0.4	57.2±0.5	81.4±0.6	68.1±1.5	70.8±0.7	78.5±1.3	70.05
	4	69.4±0.8	70.9±0.2	62.3±0.2	57.2±0.2	82.4±0.2	67.1±1.9	72.0±0.6	89.7±0.7	71.38
	6	74.6±0.5	71.6±0.3	61.5±0.3	55.9±0.3	86.2±0.3	67.2±1.6	71.2±0.5	75.0±1.6	70.40
	8	75.1±0.6	71.2±0.2	61.7±0.3	55.4±0.4	79.8±1.6	67.8±1.9	70.4±0.8	91.0±2.0	**71.55**

To verify this conjecture, we use 6 fingerprints (mcfp4_2048, atompair_2048, ecfp4_2048, maccs, physchem, rdkDes) and randomly initialized models from 6 different modalities (sequence, graph geometry graph, image, geometry image, video) for feature extraction on 8 classification datasets from MoleculeNet. In particular, the models from sequence, graph, geometry graph, image, geometry image and video are MolFormer-R, GIN-R, Uni-Mol-R, ResNet18-R, ResNet-G-R, VideoMol-R, respectively. Subsequently, we extract the scaffolds of the molecules and select the top 10 scaffolds with the highest counts as the substructures of the molecules. Finally, we sample 1,000 molecules from each of the selected substructures and use t-SNE for dimensionality reduction and clustering visualization. Note that all molecules will be sampled if the number of molecules containing a certain substructure is less than 1,000. To quantitatively analyze the model's ability to identify substructures, we calculate the Davis-Bouldin (DB) index (Davies & Bouldin, 1979) between the features of dimensionality reduction and the substructure labels, which is used to evaluate the clustering performance.

Table S50 shows the DB Index for different methods. We calculate the Pearson correlation coefficient using the results of DB index in Table S50 and the results of ROC-AUC in Table S20 . Taking BBBP as an example, we concatenate the results of BBBP on 12 methods in Table S50 into one vector as the vector of DB index. Then, we find the corresponding 12 methods in Table S20 and concatenate them into a ROC-AUC vector. Figure S8 shows the Pearson correlation coefficient calculated using the DB index vector and the ROC-AUC vector. Obviously, we can draw a conclusion: DB index is inversely proportional to ROC-AUC. This means that **the inductive bias of identifying substructures is important for predicting the properties of molecules**.

Table S50: The DB index of different models in 10 molecular substructures.

	BBBP	Sider	Tox21	ToxCast	HIV	MUV	ClinTox	BACE
mcfp4_2048	1.4224	1.5532	6.0667	5.0185	3.1764	10.3910	7.2231	0.4158
atompair_2048	1.8361	2.6594	2.9751	2.8674	4.8532	2.8937	3.0130	0.3366
ecfp4_2048	5.2635	1.1886	5.9853	3.3846	1.5788	1.9715	2.2368	0.3092
maccs	1.4097	2.9680	22.9463	7.9525	3.8023	3.4787	3.4251	0.5501
physchem	6.2917	4.3235	9.3082	10.4890	14.9467	20.1449	17.7473	7.9298
rdkDes	7.9706	20.7415	6.2466	7.1217	15.3582	24.6885	5.0376	4.8888
MolFormer-R	2.0754	1.8347	3.5817	4.1549	7.7935	17.2643	2.3998	1.1253
GIN-R	1.2580	1.5947	7.4661	4.2741	6.4118	5.4392	3.5611	1.2960
Uni-Mol-R	1.5010	2.7910	4.4361	5.2003	6.0031	3.7354	3.9110	1.1853
ResNet18-R	14.1199	13.5737	34.2620	25.3775	28.6812	36.1045	16.4683	16.3196
ResNet-G-R	8.9312	14.8282	12.5358	39.0406	13.5395	16.4114	9.5849	8.7852
VideoMol-R	12.6038	6.4966	11.0884	10.6541	10.0499	6.7280	11.6074	12.0046

Figure S9 shows t-SNE visualizations of different methods from 7 modalities on the HIV dataset. We find that fingerprint-based mcfp4_2024, sequence-based MolFormer, graph-based GIN-R, and Uni-Mol-R have a good inductive bias for identifying substructures, while the three vision-based modalities (image-based ResNet18-R, geometric image-based ResNet-G-R, and video-based VideoMol-R) need to rely on post-training. In particular, we find that MolFormer-R is consistent with mcfp4_2048 in the clustering distribution of some substructures, such as the red and cyan clusters, which may provide evidence for the similar performance of MolFormer-R and mcfp4_2048 on 8 classification tasks from MoleculeNet, with average ROC-AUC performance of 70.40% and 71.66%, respectively.

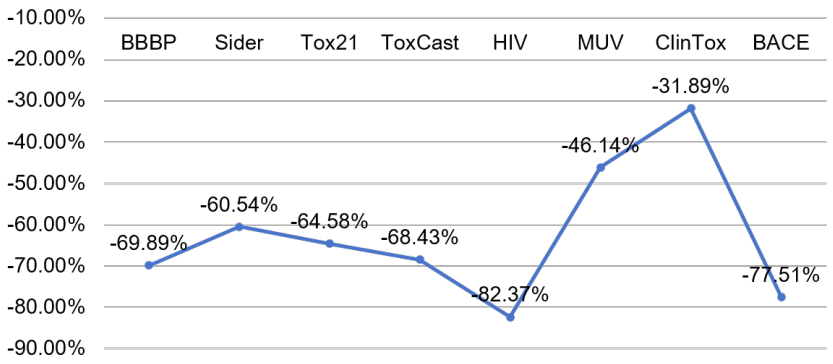

Figure S8: The Pearson correlation coefficient between the DB index vector and the ROC-AUC vector on 8 classification datasets from MoleculeNet.

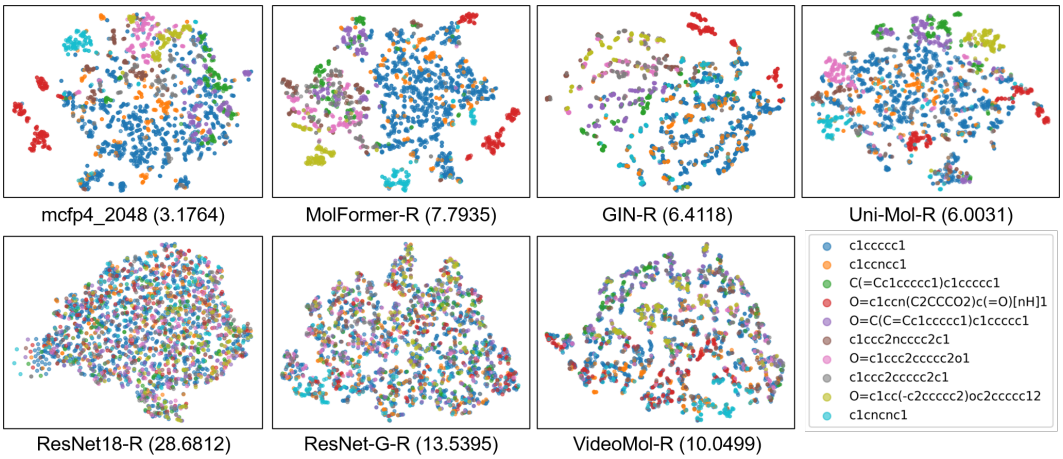

Figure S9: The t-SNE visualization of methods from 7 modalities on HIV dataset with 10 substructures. The value in brackets is the DB index.

K.6 Why Are Geometric Images And Videos Not Sensitive To RGB and BGR Formats?

A natural hypothesis about why geometric images and videos are insensitive to RGB and BGR formats is that the features extracted from RGB and BGR images have high similarity. To verify this hypothesis, we evaluate the cosine similarity between RGB features and BGR features extracted by IEM-G on 12 molecular property prediction datasets. As shown in Figure S10, we find that IEM-G has high similarity between RGB and BGR features ranging from 78.4% to 98.5%, which verifies the rationality of our hypothesis. Furthermore, we also find that RGB images and BGR images still have certain differences in features. Inspired by Appendix K.3, it is a **promising direction to use different formats of images to increase the diversity of molecules and further improve the performance of the model by fusing images of different formats**.

K.7 Why Is Video Modality So Good At Tasks Related To Atoms And Fundamental Properties?

Here we select representative models from 6 modalities, namely Chem-BERT-8L (sequence modality), MeleBERT (graph modality), TFN (geometry modality), ImageMol (image modality), IEM-G (geometry image modality), and IEM-V (video modality). We take the atomic distribution prediction task in MBANet as an example for analysis.

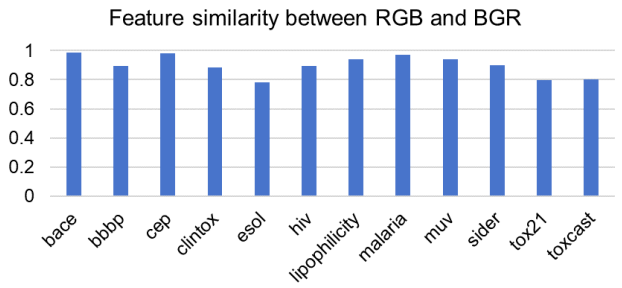

Figure S10: The cosine similarity between RGB and BGR features extracted by IEM-G on 12 molecular property prediction datasets.

In order to verify the robustness of the conclusion, we use coefficient of determination (R^2) (Di Bucchianico, 2008) and Kullback-Leibler Divergence (KLD) (Kullback & Leibler, 1951) to evaluate these methods on 6 common atomic distribution prediction tasks, including C, N, O, F, S, Cl. R^2 and KLD are used to evaluate the explanatory power of the predicted results for the true results (goodness of fit) and the difference between the predicted distribution and the true distribution, respectively. As shown in Table S51 and Table S52, we can further conclude that molecular video representation has advantages in atomic distribution prediction due to its superior performance on R^2 and KLD. Figure S11 also shows that the predicted results of the video modality (IEM-V) have very similar probability distributions to the true results.

Table S51: The R^2 performance (The higher the better) between predicted results and true results on 6 common atomic distribution prediction tasks from MBANet. The sequence, graph, geometry graph, image, geometry image, and video represent Chem-BERT-8L, MeleBERT, TFN, ImageMol, IEM-G, and IEM-V, respectively.

	C	N	O	F	S	Cl
sequence	-0.275562923	-0.00012514	-1.202954854	-0.388201064	-0.064102719	-0.003581481
graph	0.058803797	0.315124929	0.722590744	0.488687873	-0.459056973	0.774725318
geometry graph	0.763227582	0.819823027	0.887618303	0.900081873	0.818889618	0.876054287
image	0.099648419	0.446444922	0.586108756	0.443923142	0.423683317	0.376144971
geometry image	0.683815886	0.864677602	0.814648206	0.844222142	0.655123114	0.578081915
video	**0.957235135**	**0.89999567**	**0.941741096**	**0.977508473**	**0.949734432**	**0.959476785**

Table S52: The KLD performance (the smaller the better) between predicted results and true results on 6 common atomic distribution prediction tasks from MBANet. The sequence, graph, geometry graph, image, geometry image, and video represent Chem-BERT-8L, MeleBERT, TFN, ImageMol, IEM-G, and IEM-V, respectively.

	C	N	O	F	S	Cl
sequence	15.75108956	15.16975842	15.98924285	2.85108091	4.121494218	3.978459177
graph	0.811739339	0.965379503	0.486686054	0.290416465	0.305950864	1.514342345
geometry graph	0.184020462	0.976098698	1.227725127	0.430602355	1.102794542	0.850205376
image	0.185883083	0.819115708	0.68029777	2.182887729	1.368048899	0.474944866
geometry image	0.045753695	0.408585841	0.536781902	0.243507422	**0.114326531**	0.453135244
video	**0.011774227**	**0.119901919**	**0.457775393**	**0.0818768**	0.22734687	**0.178566473**

Next, we analyze the principle behind video to achieve better results. Since sequence-, graph-, and geometry-based methods have a similar learning paradigm, namely, message passing between tokens/nodes through attention, we choose the graph modality-based MoleBERT method as their representative.

We first analyze why MoleBERT performs poorly on the atom prediction task, which may be a chain reaction affecting the estimation of molecular weight in MBANet_{attr}. We conjecture that graph message passing is not conducive to the model learning the semantics of a single node because the representation of a node are determined by the representation of its neighbors. As shown in Table S53, there is no relationship between the same atoms, which means that the information

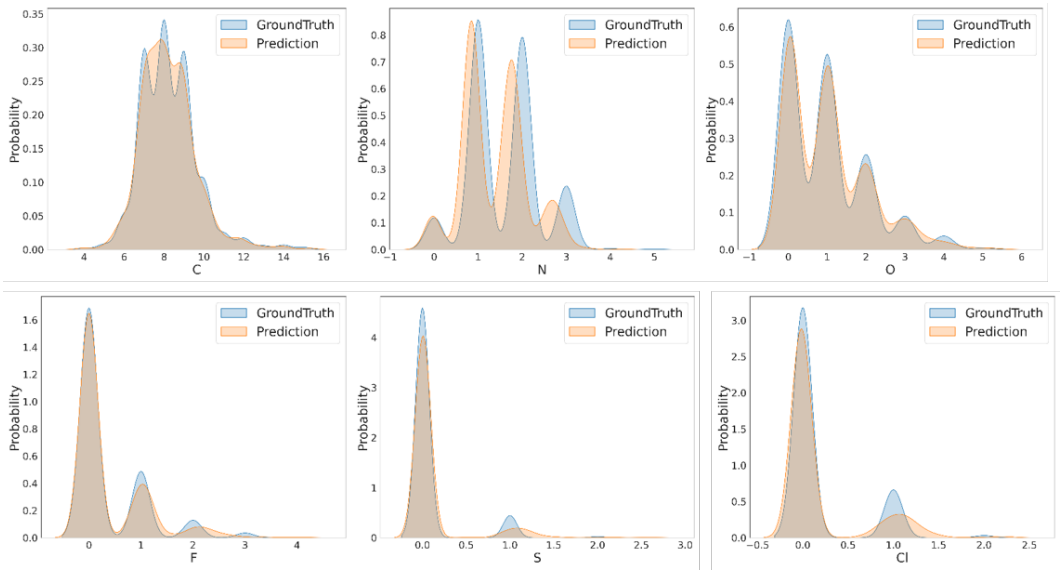

Figure S11: The probability distribution of 6 atoms (C, N, O, F, S, Cl) with video modality-based IEM-V. GroundTruth and Prediction represent the true results and the predicted results of IEM-V. The title of x-axis represents the name of atom.

represented by atoms is more affected by their neighbors than by themselves. For example, C#1 has 49% similarity with C#2 but only 10% similarity with C#8 and a higher 33% similarity with F#2 and N#4. Therefore, the inductive bias towards capturing structural information causes the graph modality to lose the ability to focus on the nodes themselves.

Table S53: Cosine similarity of pairwise atomic representations using MoleBERT on the molecule 'N#Cc1cccc2nnc(C(F)(F)F)n12'. # indicates the atom number.

	C#1	C#2	C#3	C#4	C#5	C#6	C#7	C#8	F#1	F#2	F#3	N#1	N#2	N#3	N#4
C#1	1.00	0.49	0.39	0.09	0.34	0.32	0.21	0.10	0.15	0.33	0.14	0.03	0.22	0.06	0.33
C#2	0.49	1.00	0.45	0.17	0.42	0.35	0.20	0.02	0.04	0.22	0.11	0.01	0.29	0.09	0.33
C#3	0.39	0.45	1.00	0.27	0.45	0.41	0.34	0.08	0.08	0.29	0.07	0.11	0.43	0.31	0.40
C#4	0.09	0.17	0.27	1.00	0.32	0.35	0.42	0.20	0.21	0.27	0.18	0.46	0.39	0.47	0.41
C#5	0.34	0.42	0.45	0.32	1.00	0.39	0.40	0.13	0.15	0.18	0.11	0.18	0.39	0.36	0.43
C#6	0.32	0.35	0.41	0.35	0.39	1.00	0.42	0.07	0.15	0.27	0.18	0.37	0.49	0.34	0.47
C#7	0.21	0.20	0.34	0.42	0.40	0.42	1.00	0.13	0.11	0.22	0.09	0.19	0.50	0.37	0.48
C#8	0.10	0.02	0.08	0.20	0.13	0.07	0.13	1.00	0.41	0.46	0.40	0.14	0.08	0.14	0.07
F#1	0.15	0.04	0.08	0.21	0.15	0.15	0.11	0.41	1.00	0.47	0.50	0.20	0.12	0.17	0.13
F#2	0.33	0.22	0.29	0.27	0.18	0.27	0.22	0.46	0.47	1.00	0.49	0.28	0.24	0.20	0.18
F#3	0.14	0.11	0.07	0.18	0.11	0.18	0.09	0.40	0.50	0.49	1.00	0.25	0.05	-0.03	0.11
N#1	0.03	0.01	0.11	0.46	0.18	0.37	0.19	0.14	0.20	0.28	0.25	1.00	0.39	0.34	0.30
N#2	0.22	0.29	0.43	0.39	0.39	0.49	0.50	0.08	0.12	0.24	0.05	0.39	1.00	0.49	0.46
N#3	0.06	0.09	0.31	0.47	0.36	0.34	0.37	0.14	0.17	0.20	-0.03	0.34	0.49	1.00	0.41
N#4	0.33	0.33	0.40	0.41	0.43	0.47	0.48	0.07	0.13	0.18	0.11	0.30	0.46	0.41	1.00

Different from graphs, the inductive bias of molecular videos focuses on learning local patterns. Figure S12 shows the GradCAM attention of IEM-V on MBANet$_{atom}$. We find that video-based IEM-V can obtain accurate atomic distribution information based on the local information. In particular, IEM-V accurately locates the positions of atoms with attention area in the video frame. Based on the located atoms, we can count the correct atomic distribution.

In general, **the advantage of molecular videos is that they can learn local information of molecules with high degrees of freedom, while graphs are limited by the message passing of neighbors, which weakens the extraction of local atomic-level information.**

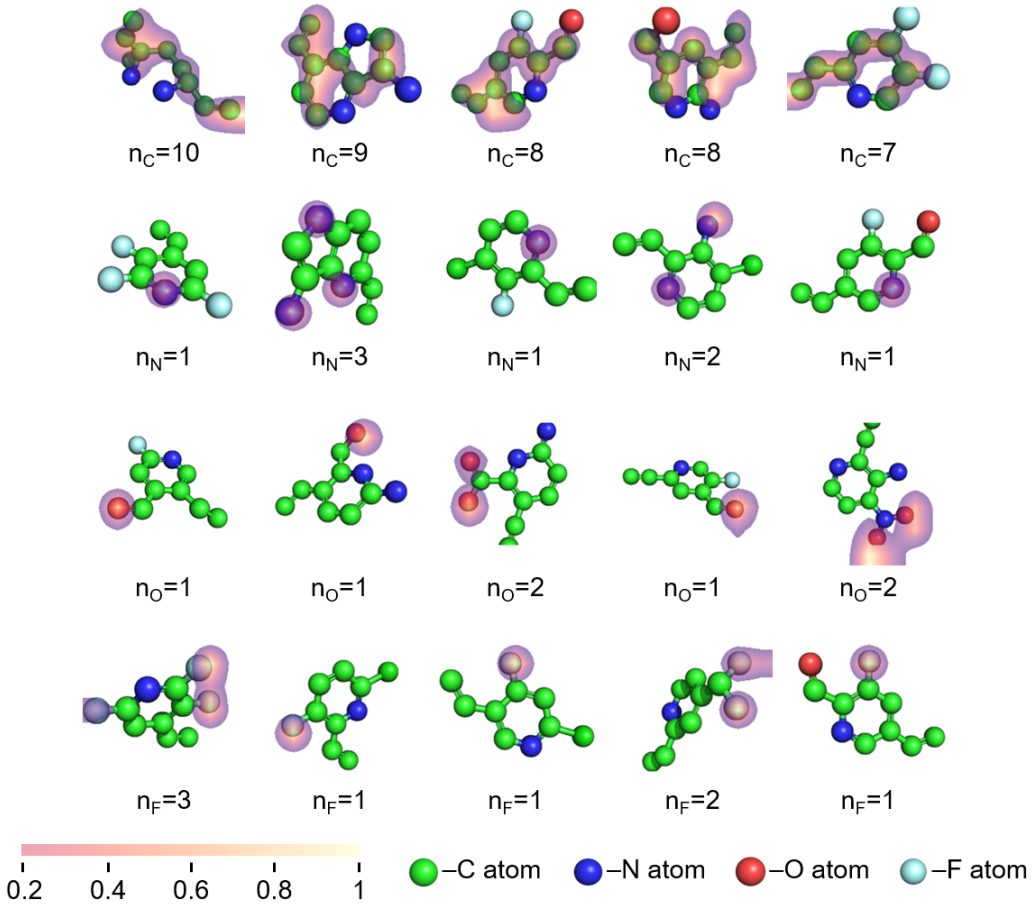

Figure S12: Grad-CAM visualization of IEM-V on frames of molecular videos. The first to fourth rows show the corresponding GradCAM visualizations of molecules when IEM-V predicts the distribution of C, N, O, and F atoms, respectively. We use 0.2 as the threshold for visualization, that is, set the importance lower than 0.2 to 0. n_C, n_N, n_O, n_F represent the number of C, N, O, F atoms, respectively.

K.8 Significance Test for Modal Preference in StructNet

In Table 6, we obtain the preferences of different modalities for molecular types. Since the means in the table are obtained from 100 results (10 runs on 10 data sets), the conclusions have good validity. Our conclusions on simplifying preferences are as follows:

- **Preference#1.** The geometry graph modality prefers acyclic (AC and A) molecules.
- **Preference#2.** The fingerprint and graph modalities prefer cyclic (CC and M) molecules.
- **Preference#3.** The visual-based modalities (image, geometry image, and video) prefer macrocyclic peptide (MP) and reticular (R) molecules.

In order to further test the robustness of the conclusions, we conduct significance tests on comparisons between different modalities. Specifically, we use a two-sided Mann-Whitney U test (Mann & Whitney, 1947) to evaluate whether the results between modalities are significantly different.

Table S54, Table S55 and Table S56 show the results of the significance test of Preference#1, Preference#2 and Preference#3 respectively. In Preference#1, we find that geometric graph prefer AC with significant differences compared with geometric images and video modalities. In Preference#2, We find that fingerprint and graph preferences are significantly different in M compared to all other

modalities. In Preference#3, modalities of image, geometry image and video prefer MP and R showing significant differences in most cases.

Overall, the average results reported in Table 6 are statistically significant and in most cases the results are significantly different.

Table S54: Results of two-sided Mann-Whitney U test between the graph modality and other modalities (fingerprint, sequence, graph, image, geometry image, video) with a significance level $p < 0.05$ on acyclic chain molecules (AC) and acyclic molecules (A) of StructNet. Green background indicates significant differences in results.

Modality	Molecular type	Fingerprint	Sequence	Graph	Image	Geometry Image	Video
Geometry Graph	AC	0.709404	0.072866	0.314039	0.176196	0.036353	0.000247
	A	0.086259	0.464252	0.089914	0.760945	0.906625	0.720347

Table S55: Results of two-sided Mann-Whitney U test between the fingerprint, graph modalities and other modalities (sequence, geometry graph, image, geometry image, video) with a significance level $p < 0.05$ on acyclic cyclic chain molecules (CC) and macro molecules (M) of StructNet. Green background indicates significant differences in results.

Molecular type	Modality	Sequence	Geometry Graph	Image	Geometry Image	Video
CC	Fingerprint	0.020995	0.036789	0.009321	0.460525	0.444346
	Graph	0.269363	0.030572	0.757225	0.242288	0.283926
M	Fingerprint	0	0	0	0	0
	Graph	0	0.00003	0	0	0

Table S56: Results of two-sided Mann-Whitney U test between the image, geometry image, video modalities and other modalities (fingerprint, sequence, graph, geometry graph) with a significance level $p < 0.05$ on macrocyclic peptide molecules (MP) and acyclic reticular molecules (R) of StructNet. Green background indicates significant differences in results.

Molecular type	Modality	Fingerprint	Sequence	Graph	Geometry Graph
MP	Image	0.000044	0.322914	0	0.000122
	Geometry Image	0.001465	0.255839	0	0.00024
	Video	0.000897	0.202102	0	0.000185
R	Image	0.000001	0.724008	0.005462	0.295615
	Geometry Image	0.000187	0.493057	0.007428	0.17973
	Video	0.000007	0.672471	0.001746	0.342439