Protein Language Model Predicts Mutation Pathogenicity and Clinical Prognosis

Xiangling Liu*
Northeastern University
liuxiangling@stumail.neu.edu.cn

Xinyu Yang*
University of Glasgow
x.yang.2@research.gla.ac.uk

Linkun Ouyang
Peking University
oylk@stu.pku.edu.cn

Guibing Guo
Northeastern University
guogb@swc.neu.edu.cn

Jin Su
Westlake University
sujin@westlake.edu.cn

Ruibin Xi†
Peking University
ruibinxi@math.pku.edu.cn

Ke Yuan†
University of Glasgow
Ke.Yuan@glasgow.ac.uk

Fajie Yuan†
Westlake University
yuanfajie@westlake.edu.cn

Abstract

Accurately predicting the effects of mutations in cancer has the potential to improve existing treatments and identify novel therapeutic targets. In this paper, we evidence for the first time that the large-scale pre-trained protein language models (PPLMs) are zero-shot predictors for two *clinically* relevant tasks: identifying disease-causing mutations and predicting patient survival rate. Then we benchmark a series of state-of-the-art (SOTA) PPLMs on 2279 protein variants across 20 cancer-related genes. Our empirical results show that the PPLMs outperform the SOTA baseline, EVE [1], trained on multiple sequence alignment (MSA) data. We also demonstrate that the evolutionary index score, generated from the PPLM's softmax layer, is good indicator for both mutation pathogenicity and patient survival rate. Our paper has taken a key step toward the clinical utility of large-scale PPLMs.

1 Introduction

Whether a mutation causes disease, known as the pathogenicity of a mutation, is a fundamental question in modern genomics. Traditional methods of predicting pathogenicity rely on mutation frequency and clinical evidence in the literature [2]. However, only less than 2% of mutations currently have known functional interpretation [3, 4, 5], leaving most variants have yet to be identified with clinical consequences. Conventional supervised learning models have been tested for this task [6, 7, 8, 9, 10, 11, 12], but the accuracy of these methods remains limited [13].

Recent advances in large-scale language models, such as GPT [14] and BERT, [15] have motivated powerful protein language models by leveraging the similarity between protein sequences and

*Equal Contribution. Work done when Xiangling was a visiting student at Westlake University.
†Corresponding Author.

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sentences. Among them, the BERT-based pre-trained protein language model (PPLM), a.k.a. ESM-1b [16], trained on 250 million protein sequences in UniRef50 [17], can capture the physical properties of amino acids. The newer version, ESMFold [18], equipped with a much larger PPLM, called ESM2, can predict protein structures with comparable accuracy with AlphaFold [19]. On the other side, a recent study showed [20] that the Evoformer-based PPLM module in AlphaFold2 couldn't predict protein mutation effects well, perhaps because AlphaFold2 were trained with less fewer protein sequences than ESM1b. A bidirectional LSTM model has been shown to predict whether viruses can escape attacks from hosts' immune systems [21]. Another similar model is the GPT2-based ProGen2 [22]. Those models can make predictions using a single protein sequence.

There is another direction of research that leverages sequences of homologous proteins. Examples include BERT-based ESM-MSA [23] and VAE-based DeepSequence [24] model. While these models have been tested for predicting mutation effects acrosss a variety of deep mutational scanning experiments, focused assessments in cancer-related mutations are still lacking. More importantly, whether these PPLMs can predict clinically relevant properties (i.e. prognosis) of the cancer patients carrying these mutations remains unknown.

In this paper, we present a systematic benchmark study of state-of-the-art protein language models in predicting pathogenic mutations in cancer diver genes. The models include alignment-based methods, i.e. EVE [1] and ESM-MSA [23], and alignment free approaches, including ESM-1, ESM-1b [16], ESM-1v [25], ESM-2 [18], and ProGen2 [22]. We examine whether these models can learn functional changes caused by mutations in amino acid sequences and identify high-risk mutant cancer proteins. We further test representations learned from these PPLM in a cox-regression framework for progression-free survival prediction. Evaluated on 10,248 patients from The Cancer Genome Atlas (TCGA), the winning model, ESM-1b, can achieve statically significant separation of high and low risk patients in six cancer types, while traditional methods are known to struggle.

2 Method and Analysis

2.1 Zero-shot Prediction of Pathogenic Mutations using Pre-Trained Protein Language Models

Here, we lay out a zero-shot pathogenic mutation prediction task for pre-trained protein language models. As illustrated in Figure 1, for each each mutation, the model will take the entire protein sequence as input. The output is the probability of sequence being pathogenic or benign. These probabilities are assignment probabilities obtained from fitting a two-component Gaussian mixture to a measurement called evolutionary index. Evolutionary index (EI) is the negative log ratio between the probability of observing the mutant sequence and the probability of observing the wild-type sequence (i.e. sequence without mutation) [24, 1]. The EI has been shown to be an intuitive score to reflect a protein language model's ability to capture information encoded in amino acid sequences [1, 21]. A higher value in EI indicates stronger deviation of the mutant sequence from the wild-type sequence. Therefore, the component with a higher mean value represents a higher chance of being pathogenic. A straightforward classification can be based on the probability of assigning a mutation to the pathogenic cluster as the pathogenicity score.

The strategy allow us to test pre-trained protein language models solely using their representations, avoiding any confounding factors with further tuning a downstream classifier.

We extracted 20 common cancer proteins from ClinVar [5], which documents and provides supporting evidence for the relationship between variation and phenotype in humans. ClinVar provided the assessment criteria for the clinical significance of variation. ClinVar's annotations are widely accepted as ground truth. We screened for human cancer proteins, aiming to select as many representative cancer proteins as possible with clinical data. It is important to note that the importation of the protein into the ESM-1b network, the evolutionary index, and the final pathogenicity score were unsupervised and unadulterated with homologous sequence information.

Testing on the ClinVar labels, ESM-1b obtained an AUC of 0.874 and an average ACC of 0.826 (shown in Figure 2), while ESM-1v achieved better results with an AUC of 0.909 (shown in Table 2), outperforming the current leading performance reported [1]. The pathogenic score makes the performance highly explainable. The degree of separation between two Gaussian components directly assesses the strength of the predictive signal captured by ESM-1b. By aligning the score with the

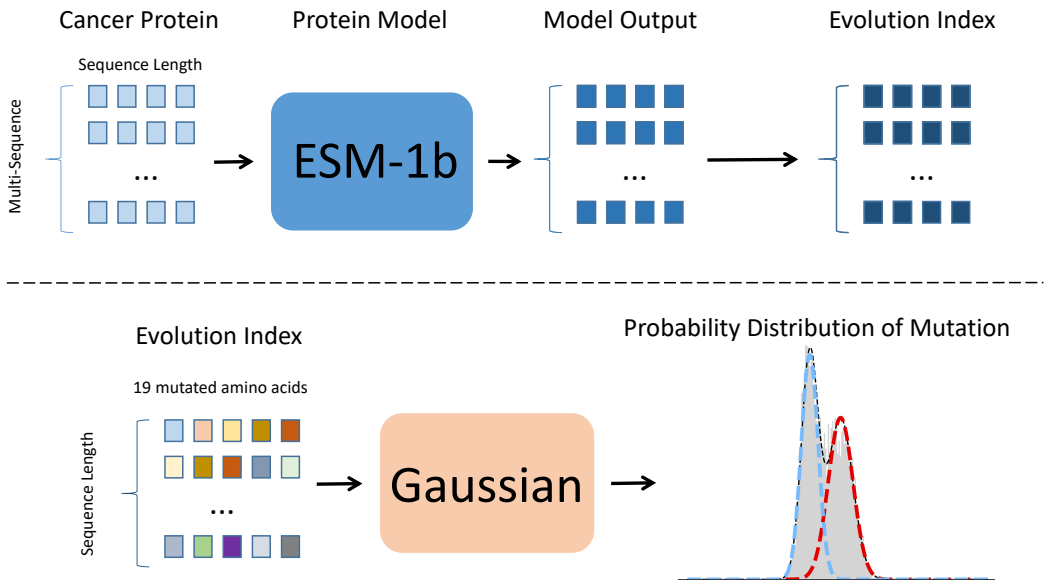
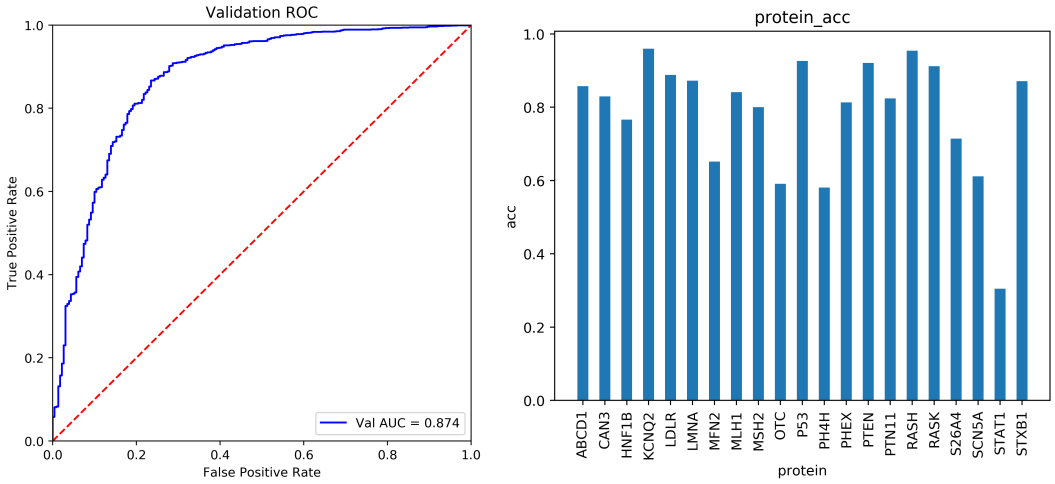


Figure 1: Zero-shot pathogenic mutation prediction framework, using ESM-1b as an example.



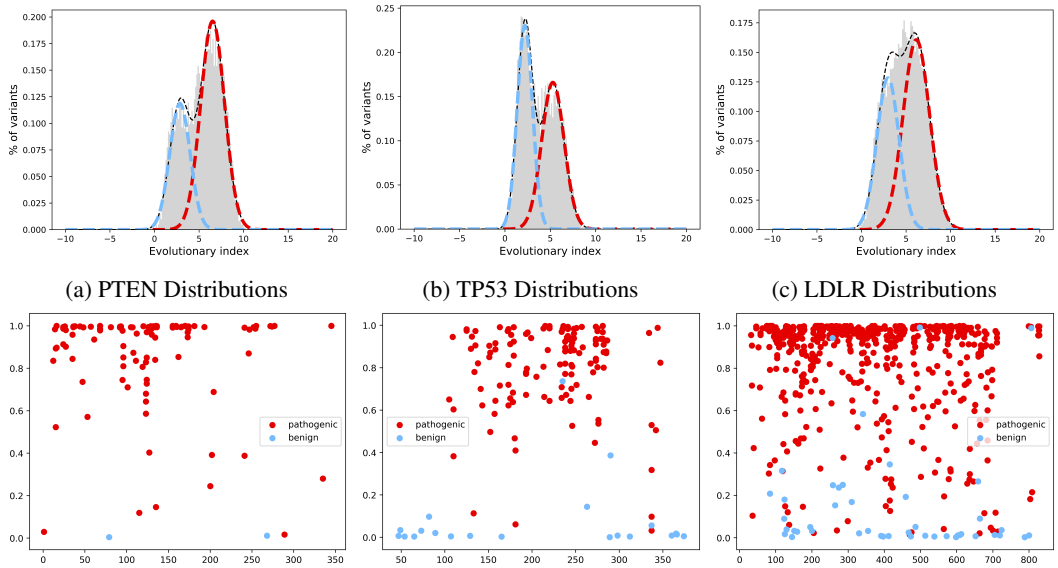
(a) AUC (b) ACC

Figure 2: (a) shows the AUC (area under curve) of the pathogenicity scores of 20 common cancer proteins under the Clinvar clinical label, and (b) shows the accuracy of the pathogenicity labels of 20 cancer proteins.

genomic position of corresponding mutations, one can further examine the biological implication of the predictions.

2.2 Benchmark Protein Language Models with Zero-Shot Pathogenicity Prediction

Here, we examine whether pre-trained protein language models can learn functional changes caused by mutations in amino acid sequences and identify high-risk mutant cancer proteins, using the zero-shot prediction task described in the previous subsection. We base our evaluation on six recently proposed large-scale pretrained protein models. The models include alignment-based methods such as EVE [1] and ESM-MSA [23], and alignment free approaches such as ESM-1, ESM-1b [16], ESM-1v [25],ESM-2[18],and ProGen2 [22].



(a) PTEN Distributions (b) TP53 Distributions (c) LDLR Distributions

(d) PTEN Clustering (e) TP53 Clustering (f) LDLR Clustering

Figure 3: (a), (b), and (c) show the fitting of two-component Gaussian mixture models on evolutionary index scores in PTEN, TP53 and LDLR. There is a clear distinction between benign (blue dashed line) and pathogenic (red dashed line) components. (d), (e), and (f) show the relationship between the pathogenicity score (i.e., the probability of being in the pathogenic component) colour-coded using Clinvar annotations where red denotes pathogenic and blue denotes benign.

To test whether wild-type context information aid the prediction of mutation pathogenicity, we introduce an additional consideration when testing the model. The input protein sequences follow two settings: one is making the input protein sequence completely visible to the model; the other is we mask amino acid sites of the input protein sequence. When we do not mask, the model learns the probability of this amino acid mutating into other amino acids based on the context information of the mutation site and the wild-type. When masking, the model learns what the masked amino acid should be based on the context information of the mutation site. In our experiment results, we found that masking amino acid sites on the input protein sequences has worse performance on the model's prediction ability (without masked ESM-1b: AUC=0.874; masked ESM-1b: AUC=0.706, as shown in Table 2).

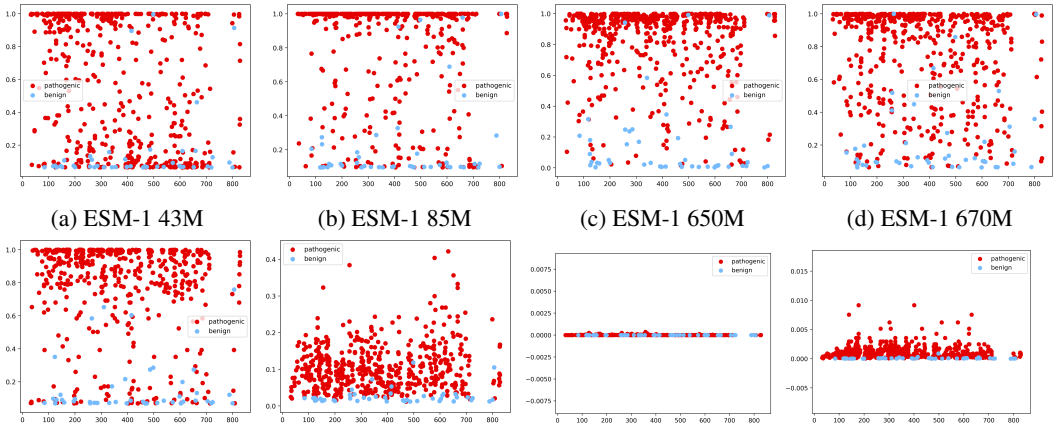
At the same time, we found that models trained without MSA, such as ESM-1b and ESM-1v, were no worse or even better than models trained with MSA, such as EVE (ESM-1v: AUC=0.909; EVE: AUC=0.887). This result suggests that biological information learned from large-scale protein databases was richer than that from specialized homologous sequences, with the advantage of much less computational time. At the same time, we found that the larger the training dataset, the better the model's prediction ability. We also evaluated the performance among models of different scales. We found that the size of a pre-trained protein model is not proportional to its ability to learn the pathogenicity of protein mutations (With the increase in model scale, the AUC of ESM-1 was 0.725, 0.769, 0.874, and 0.856, respectively, shown in Figure 4,and the same as ESM-2). The Zero-shot pathogenicity prediction performance of ESM-1 and ESM-2 does not improve with the increase in model scale; it reaches a peak (at 650M parameters) and then gradually decreases. We also tested the generative model ProGen2, as shown in Table 1. We found that the generative model could also perform a good pathogenicity prediction, and the best 6.4B model reached AUC = 0.862.

Table 1: ProGen2 Model.

Scale	151M	764M	2.7B	6.4B
AUC	0.794	0.825	0.850	0.862

Table 2: Benchmark protein language models with zero-shot pathogenicity prediction.

Model	MSA	Params	AUC	
			Mask Predict	Without Mask
EVE	1	-	-	0.887
ESM-1	0	43M	0.722	0.725
		85M	0.780	0.769
		670M	0.866	0.856
ESM-1b	0	650M	0.706	0.874
ESM-1v	0	650M	0.758	0.909
ESM-MSA	1	100M	-	0,598
ESM-2	0	8M	0.695	0.701
		35M	0.723	0.729
		150M	0.764	0.800
		650M	0.827	0.822
		3B	0.812	0.807
		15B	-	0.799



(a) ESM-1 43M (b) ESM-1 85M (c) ESM-1 650M (d) ESM-1 670M

(e) ProGen2 151M (f) ProGen2 764M (g) ProGen2 2.7B (h) ProGen2 6.4B

Figure 4: Variation in pathogenicity score of mutations in LDLR with different number of parameters in ESM-1 and ProGen2.

2.3 Predict the Clinical Prognosis of Cancer Patients

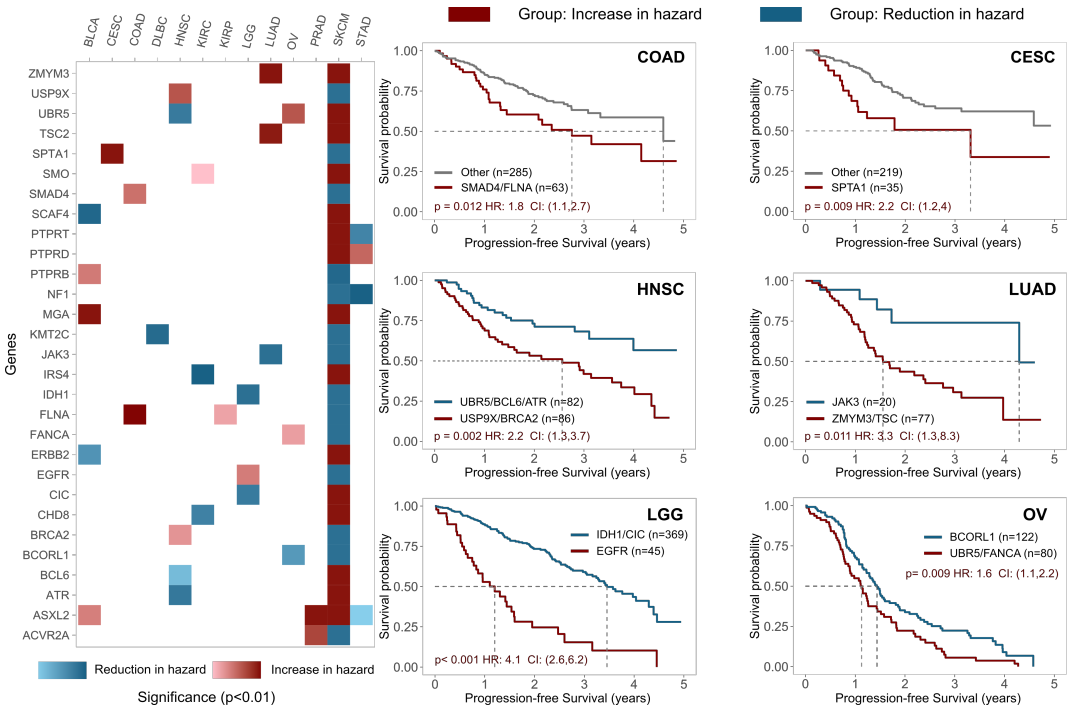
To investigate the clinical utility of pre-trained protein language models, we examined the prognostic value of the evolutionary index computed using ESM-1b on real-world cancer patients' data. TCGA is one of the largest datasets from which matched tumour genomic sequencing and clinical outcome data are publicly available [26]. We selected 412 cancer driver genes with suitable protein canonical sequences out of the 579 Tier-1 genes in the COSMIC (Catalogues of somatic mutation in cancer) [27] cancer gene census. EI was estimated by the ESM-1b model for each protein sequence in our driver gene list for each patient.

We performed multivariate Cox proportional-hazards regression (stratified by gender and age) on proteins' EIs for 10,248 patients across different TCGA cohorts with progression-free interval (PFI) as the adverse outcome. For the 13 cancer types for which our framework was effective, we found

that high EI values of specific proteins significantly contribute to better/worse survival (Figure 5a). For example, the high EI value of SMAD4 and FLNA proteins in colorectal cancer (COAD) showed significant evidence of increased patient hazard risk ($p<0.01$, log-rank test). In contrast, the high EI value of JAK3 protein contributed to a lower hazard risk ($p<0.01$, log-rank test) in lung cancer (LUAD).

Furthermore, we stratified patients into the hazard-increase and hazard-reduction groups based on whether EI>0 for specific proteins suggested by Cox regression for each cancer type. We drew Kaplan-Meier(KM) curves and tested the survival difference between the two groups with estimated hazard ratios(HR) and p values of the log-rank test. Predictive accuracy was assessed using the concordance index (c-index). The c-index compares the ranks of subjects as predicted by the PFI using the stratified Cox model based on the actual disease outcome. We observed significant difference between hazard-increase and hazard-reduction/other group across 6 cancer types (COAD: p=0.012,HR=1.8,CI=1.1-2.7; CESC: p=0.009,HR=2.2,CI=1.2-4; HNSC: p=0.002,HR=2.2,CI=1.3-3.7; LUAD: p=0.011,HR=3.3,CI=1.3-8.3; LGG: p<0.001,HR=4.1,CI=2.6-6.2; OV: p=0.009,HR=1.6,CI=1.1-2.2; Figure 5b).

These results demonstrate that the evolutionary index can achieve statistically significant survival prediction in multiple cancer types.



(a) (b)

Figure 5: a) Genes with the significant prognostic power across tumours, suggested by multivariate Cox proportional-hazards regression; values in each cell denote the p-value of the log-rank test for corresponding driver genes. b) Kaplan–Meier curves for patient subgroups.

3 Conclusions

In this paper, we verified that the large-scale pre-trained protein language models can efficiently and accurately predict the effect of mutations in cancer driver genes. Comparable results were obtained with models learned from homologous sequences and those learned from single sequences.

We propose a systematic benchmark based on a zero-short pathogenic mutation prediction task. The experimental results show that BERT-like models such as ESM-1b are better suited to the task than those that rely on generative models. We also found that the size of a pre-trained protein model is

not proportional to its performance in predicting pathogenic mutations. This observation aligns with DeepMind's finding that model performance might drop as the number of model parameters increases, because a large model might be under-trained with limited data [28]. In our case, the complexity and diversity of protein sequences might have been a limiting factor for sufficiently training large models. It is widely hypothesised that existing protein databases only capture a fraction of proteins that exist in living organisms. Finally, we demonstrated the prognostic value of protein language model in TCGA cohorts. The pathogenicity information captured by pre-trained protein model can separate high and low risk patients in six cancer types, while traditional methods have yet been demonstrated success.

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References

[1] Jonathan Frazer, Pascal Notin, Mafalda Dias, Aidan Gomez, Joseph K Min, Kelly Brock, Yarin Gal, and Debora S Marks. Disease variant prediction with deep generative models of evolutionary data. *Nature*, 599(7883):91–95, 2021.

[2] Melissa J Landrum, Jennifer M Lee, Mark Benson, Garth R Brown, Chen Chao, Shanmuga Chitipiralla, Baoshan Gu, Jennifer Hart, Douglas Hoffman, Wonhee Jang, et al. Clinvar: improving access to variant interpretations and supporting evidence. *Nucleic acids research*, 46(D1):D1062–D1067, 2018.

[3] Cristopher V Van Hout, Ioanna Tachmazidou, Joshua D Backman, Joshua D Hoffman, Daren Liu, Ashutosh K Pandey, Claudia Gonzaga-Jauregui, Shareef Khalid, Bin Ye, Nilanjana Banerjee, et al. Exome sequencing and characterization of 49,960 individuals in the uk biobank. *Nature*, 586(7831):749–756, 2020.

[4] Konrad J Karczewski, Laurent C Francioli, Grace Tiao, Beryl B Cummings, Jessica Alföldi, Qingbo Wang, Ryan L Collins, Kristen M Laricchia, Andrea Ganna, Daniel P Birnbaum, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*, 581(7809):434–443, 2020.

[5] Melissa J Landrum and Brandi L Kattman. Clinvar at five years: Delivering on the promise. *Human mutation*, 39(11):1623–1630, 2018.

[6] Daniele Raimondi, Ibrahim Tanyalcin, Julien Ferté, Andrea Gazzo, Gabriele Orlando, Tom Lenaerts, Marianne Rooman, and Wim Vranken. Deogen2: prediction and interactive visualization of single amino acid variant deleteriousness in human proteins. *Nucleic acids research*, 45(W1):W201–W206, 2017.

[7] Bing-Jian Feng. Perch: a unified framework for disease gene prioritization. *Human mutation*, 38(3):243–251, 2017.

[8] Nilah M Ioannidis, Joseph H Rothstein, Vikas Pejaver, Sumit Middha, Shannon K McDonnell, Saurabh Baheti, Anthony Musolf, Qing Li, Emily Holzinger, Danielle Karyadi, et al. Revel: an ensemble method for predicting the pathogenicity of rare missense variants. *The American Journal of Human Genetics*, 99(4):877–885, 2016.

[9] Karthik A Jagadeesh, Aaron M Wenger, Mark J Berger, Harendra Guturu, Peter D Stenson, David N Cooper, Jonathan A Bernstein, and Gill Bejerano. M-cap eliminates a majority of variants of uncertain significance in clinical exomes at high sensitivity. *Nature genetics*, 48(12):1581–1586, 2016.

[10] Ivan A Adzhubei, Steffen Schmidt, Leonid Peshkin, Vasily E Ramensky, Anna Gerasimova, Peer Bork, Alexey S Kondrashov, and Shamil R Sunyaev. A method and server for predicting damaging missense mutations. *Nature methods*, 7(4):248–249, 2010.

[11] Iuliana Ionita-Laza, Kenneth McCallum, Bin Xu, and Joseph D Buxbaum. A spectral approach integrating functional genomic annotations for coding and noncoding variants. *Nature genetics*, 48(2):214–220, 2016.

[12] Philipp Rentzsch, Daniela Witten, Gregory M Cooper, Jay Shendure, and Martin Kircher. Cadd: predicting the deleteriousness of variants throughout the human genome. *Nucleic acids research*, 47(D1):D886–D894, 2019.

[13] Sue Richards, Nazneen Aziz, Sherri Bale, David Bick, Soma Das, Julie Gastier-Foster, Wayne W Grody, Madhuri Hegde, Elaine Lyon, Elaine Spector, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the american college of medical genetics and genomics and the association for molecular pathology. *Genetics in medicine*, 17(5):405–423, 2015.

[14] Alec Radford, Jeffrey Wu, Rewon Child, David Luan, Dario Amodei, Ilya Sutskever, et al. Language models are unsupervised multitask learners. *OpenAI blog*, 1(8):9, 2019.

[15] Jacob Devlin, Ming-Wei Chang, Kenton Lee, and Kristina Toutanova. Bert: Pre-training of deep bidirectional transformers for language understanding. *arXiv preprint arXiv:1810.04805*, 2018.

[16] Alexander Rives, Joshua Meier, Tom Sercu, Siddharth Goyal, Zeming Lin, Jason Liu, Demi Guo, Myle Ott, C Lawrence Zitnick, Jerry Ma, et al. Biological structure and function emerge from scaling unsupervised learning to 250 million protein sequences. *Proceedings of the National Academy of Sciences*, 118(15):e2016239118, 2021.

[17] Baris E Suzek, Hongzhan Huang, Peter McGarvey, Raja Mazumder, and Cathy H Wu. Uniref: comprehensive and non-redundant uniprot reference clusters. *Bioinformatics*, 23(10):1282–1288, 2007.

[18] Zeming Lin, Halil Akin, Roshan Rao, Brian Hie, Zhongkai Zhu, Wenting Lu, Nikita Smetanin, Robert Verkuil, Ori Kabeli, Yaniv Shmueli, Allan dos Santos Costa, Maryam Fazel-Zarandi, Tom Sercu, Salvatore Candido, and Alexander Rives. Evolutionary-scale prediction of atomic level protein structure with a language model. *bioRxiv*, 2022.

[19] Zeming Lin, Halil Akin, Roshan Rao, Brian Hie, Zhongkai Zhu, Wenting Lu, Allan dos Santos Costa, Maryam Fazel-Zarandi, Tom Sercu, Sal Candido, et al. Language models of protein sequences at the scale of evolution enable accurate structure prediction. *bioRxiv*, 2022.

[20] Mingyang Hu, Fajie Yuan, Kevin K Yang, Fusong Ju, Jin Su, Hui Wang, Fei Yang, and Qiuyang Ding. Exploring evolution-aware &-free protein language models as protein function predictors. *Advances in Neural Information Processing Systems*, 2022.

[21] Brian Hie, Ellen D Zhong, Bonnie Berger, and Bryan Bryson. Learning the language of viral evolution and escape. *Science*, 371(6526):284–288, 2021.

[22] Erik Nijkamp, Jeffrey Ruffolo, Eli N Weinstein, Nikhil Naik, and Ali Madani. Progen2: exploring the boundaries of protein language models. *arXiv preprint arXiv:2206.13517*, 2022.

[23] Roshan M Rao, Jason Liu, Robert Verkuil, Joshua Meier, John Canny, Pieter Abbeel, Tom Sercu, and Alexander Rives. Msa transformer. In *International Conference on Machine Learning*, pages 8844–8856. PMLR, 2021.

[24] Adam J Riesselman, John B Ingraham, and Debora S Marks. Deep generative models of genetic variation capture the effects of mutations. *Nature methods*, 15(10):816–822, 2018.

[25] Joshua Meier, Roshan Rao, Robert Verkuil, Jason Liu, Tom Sercu, and Alex Rives. Language models enable zero-shot prediction of the effects of mutations on protein function. *Advances in Neural Information Processing Systems*, 34:29287–29303, 2021.

[26] Carolyn Hutter and Jean Claude Zenklusen. The cancer genome atlas: creating lasting value beyond its data. *Cell*, 173(2):283–285, 2018.

[27] Simon A Forbes, David Beare, Harry Boutselakis, Sally Bamford, Nidhi Bindal, John Tate, Charlotte G Cole, Sari Ward, Elisabeth Dawson, Laura Ponting, et al. Cosmic: somatic cancer genetics at high-resolution. *Nucleic acids research*, 45(D1):D777–D783, 2017.

[28] Jordan Hoffmann, Sebastian Borgeaud, Arthur Mensch, Elena Buchatskaya, Trevor Cai, Eliza Rutherford, Diego de Las Casas, Lisa Anne Hendricks, Johannes Welbl, Aidan Clark, et al. Training compute-optimal large language models. *arXiv preprint arXiv:2203.15556*, 2022.