GENERATIVE MODELS FOR PROTEIN DESIGN

ABSTRACT

Engineered proteins offer the potential to solve many problems in biomedicine, energy, and materials science, but creating designs that succeed is difficult in practice. A significant aspect of this challenge is the complex coupling between protein sequence and 3D structure, and the task of finding a viable design is often referred to as the *inverse protein folding problem*. We develop generative models for protein sequences conditioned on a graph-structured specification of the design target. Our approach efficiently captures the complex dependency structures in proteins by focusing on dependencies that are long-range in sequence but local in 3D space. Our framework significantly improves upon prior parametric models of protein sequences given structure, and takes a step toward rapid and targeted biomolecular design with the aid of deep generative models.

1 INTRODUCTION

A central goal for computational protein design is to automate the invention of protein molecules with defined structural and chemical properties. This field has seen tremendous progess in the past two decades (Huang et al., 2016), including the design of novel 3D folds (Kuhlman et al., 2003), enzymes (Siegel et al., 2010), and complexes (Bale et al., 2016). However, the current practice often requires multiple rounds of trial-and-error, with first designs frequently failing (Koga et al., 2012; Rocklin et al., 2017). Several of the challenges stem from the bottom-up nature of contemporary approaches that rely on both the accuracy of *energy functions* to describe protein physics as well as on the efficiency of *sampling algorithms* to explore the protein sequence and structure space.

Here, we explore an alternative, top-down framework for protein design that directly learns a conditional generative model for protein *sequences* given a specification of the target structure, which is represented as a *graph* over the sequence elements. Specifically, we augment the autoregressive self-attention of recent sequence models (Vaswani et al., 2017) with graph-based descriptions of the 3D structure. By composing multiple layers of structured self-attention, our model can effectively capture higher-order, interaction-based dependencies between sequence and structure, in contrast to previous parameteric approaches (O'Connell et al., 2018; Wang et al., 2018) that are limited to only the first-order effects.

The graph-structured conditioning of a sequence model affords several benefits, including favorable computational efficiency, inductive bias, and representational flexibility. We accomplish the first two by leveraging a well-evidenced finding in protein science, namely that long-range dependencies in sequence are generally short-range in 3D space (Morcos et al., 2011; Marks et al., 2011; Balakrishnan et al., 2011). By making the graph and self-attention similarly sparse and localized in 3D space, we achieve computational scaling that is linear in sequence length. Additionally, graph structured inputs offer representational flexibility, as they accomodate both coarse, 'flexible backbone' (connectivity and topology) as well as fine-grained (precise atom locations) descriptions of structure.

We demonstrate the merits of our approach via a detailed empirical study. Specifically, we evaluate our model at *structural generalization* to sequences of protein folds that were outside of the training set. Our model achieves considerably improved generalization performance over the recent deep models of protein sequence given structure as well as structure-naïve language models.

1.1 RELATED WORK

Generative models for proteins A number of works have explored the use of generative models for protein engineering and design (Yang et al., 2018). Recently O'Connell et al. (2018) and Wang et al. (2018) proposed neural models for sequences given 3D structure, where the amino acids at different positions in the sequence are predicted independently of one another. Greener et al. (2018) introduced a generative model for protein sequences conditioned on a 1D, context-free grammar based specification of the fold topology. Boomsma & Frellsen (2017) and Weiler et al. (2018) used deep neural networks to model the conditional distribution of letters in a specific position given the structure and sequence of all surrounding residues. In contrast to these works, our model captures the joint distribution of the full protein sequence while grounding these dependencies in terms of long-range interactions arising from the structure.

In parallel to the development of structure-based models, there has been considerable work on deep generative models for protein sequences in individual protein families with directed (Riesselman et al., 2018; Sinai et al., 2017) and undirected (Tubiana et al., 2018) latent variable models. These methods have proven useful for protein engineering, but presume the availability of a large number of sequences from a particular family.

In a complementary line of research, deep models of protein structure (Anand & Huang, 2018; Ingraham et al., 2019; AlQuraishi, 2018) have been proposed recently that could be used to craft 3D structures for input to sequence design.

Protein design For classical approaches to computational protein design, which are based on joint modeling of structure and sequence, we refer the reader to a review of both methods and accomplishments in Huang et al. (2016). More recently, Zhou et al. (2018) proposed a non-parametric approach to protein design in which a target design is decomposed into substructural motifs that are then queried against a protein database. In this work we will focus on comparisons with direct parametric models of the sequence-structure relationship.

Self-Attention Our model extends the Transformer (Biswas et al., 2018) to additionally capture sparse, pairwise relational information between sequence elements. The dense variation of this problem was explored in Shaw et al. (2018) and Huang et al. (2018). As noted in those works, incorporating general pairwise information incurs $\mathcal{O}(L^2)$ memory (and computational) cost for sequences of length L, which can be highly limiting for training on GPUs. We circumvent this cost by instead restricting the self-attention to the sparsity of the input graph. Given this graph-structured self-attention, our model may also be reasonably cast in the framework of message-passing or graph neural networks (Gilmer et al., 2017; Battaglia et al., 2018).

2 METHODS

2.1 REPRESENTING STRUCTURE

We represent molecular structure in terms of an attributed graph $\mathcal{G} = (\mathcal{V}, \mathcal{E})$ with node features $\mathcal{V} = \{\boldsymbol{v}_1, \ldots, \boldsymbol{v}_N\}$ and edge features $\mathcal{E} = \{\boldsymbol{e}_{ij}\}_{i \neq j}$. This formulation can accommodate different variations on the macromolecular design problem, including both the 'rigid backbone' design where the precise coordinates of backbone atoms are fixed, as well as the 'flexible backbone' design where looser constraints such as blueprints of hydrogen-bonding connectivity (Koga et al., 2012) or 1D architectures (Greener et al., 2018) could define the structure of interest.

3D considerations For a rigid-body design problem, the structure for conditioning is a fixed set of backbone coordinates $\mathcal{X} = \{\boldsymbol{x}_i \in \mathbb{R}^3 : 1 \leq i \leq N\}$, where N is the number of positions[1]. We desire a graph representation of the coordinates $\mathcal{G}(\mathcal{X})$ that has two properties:

- *Invariance*. The features are invariant to rotations and translations.

---

[1]Here we consider a single representative coordinate per position when deriving edge features but may revisit multiple atom types per position for features such as backbone angles or hydrogen bonds.

Figure 1: **An autoregressive self-attention model for protein sequences given 3D structures.** (A) The encoder develops position-wise representations of structure using multi-head self-attention (Vaswani et al., 2017) over nodes and edges of an input graph. The attention heads are structured by the sparsity of the input graph, enabling efficient computation for large molecules with thousands of atoms (See Figure 2 for examples). (B) For rigid-body protein design, the graph encodings of atomic structure (left, top) are based on an encoding of relative positioning of C_α coordinates x_i (left, middle), which are endowed with local coordinate systems O_i based on backbone geometry. The graph edge features (left, bottom) encode the 6DoF transformations between local coordinate systems (x_i, O_i) and (x_j, O_j). For sparsity, we derive a k-Nearest Neighbors graph from the Euclidean distances (right, top) and restrict all subsequent computation such as orientation calculations (right, bottom) to this graph.

- *Locally informative.* The edge features incident to v_i due to its neighbors $N(i)$, i.e. $\{e_{ij}\}_{j \in N(i)}$, contain sufficient information to reconstruct all adjacent coordinates $\{x_j\}_{j \in N(i)}$ up to rigid-body motion.

While invariance is motivated by standard symmetry considerations, the second property is motivated by the limitations of current graph neural networks (Gilmer et al., 2017). In these networks, updates to node features v_i depend only on the edge and node features adjacent to v_i. However, typically, these features are insufficient to reconstruct the relative neighborhood positions $\{x_j\}_{j \in N(i)}$, so individual updates cannot fully depend on the 'local environment'. For example, pairwise distances D_{ij} and D_{il} are insufficient to determine if x_j and x_l are on the same or opposite sides of x_i.

Structural encodings We develop invariant and locally informative features by first augmenting the points x_i with 'orientations' O_i that define a local coordinate system at each point. We define these in terms of the backbone geometry as

$$O_i = [b_i \ \ n_i \ \ b_i \times n_i], \tag{1}$$

where b_i is the negative bisector of angle between the rays $(x_{i-1} - x_i)$ and $(x_{i+1} - x_i)$, and n_i is a unit vector normal to that plane. Formally, we have

$$u_i = \frac{x_i - x_{i-1}}{||x_i - x_{i-1}||}, \ \ b_i = \frac{u_i - u_{i+1}}{||u_i - u_{i+1}||}, \ \ n_i = \frac{u_i \times u_{i+1}}{||u_i \times u_{i+1}||}. \tag{2}$$

Finally, we derive the spatial edge features $e_{ij}^{(s)}$ from the rigid body transformation that relates reference frame (x_i, O_i) to reference frame (x_j, O_j). While this transformation has 6 degrees of freedom, we decompose it into features for distance, direction, and orientation as

$$e_{ij}^{(s)} = \text{Concat}\left(r\left(||x_j - x_i||\right), \ \ O_i^T \frac{x_j - x_i}{||x_j - x_i||}, \ \ q\left(O_i^T O_j\right) \right). \tag{3}$$

Here $\mathbf{r}(\cdot)$ is a function that lifts the distances into a radial basis[2], the term in the middle corresponds to the relative direction of $\boldsymbol{x}_j$ in the reference frame of $(\boldsymbol{x}_i, \boldsymbol{O}_i)$, and $\mathbf{q}(\cdot)$ converts the 3×3 relative rotation matrix to a quaternion representation. Quaternions represent rotations as four-element vectors that can be efficiently and reasonably compared by inner products Huynh (2009).[3]

Positional encodings Taking a cue from the original Transformer model, we obtain positional embeddings $\boldsymbol{e}_{ij}^{(p)}$ that encode the role of local structure around node i. Specifically, we need to model the positioning of each neighbor j relative to the node under consideration i. Therefore, we obtain the position embedding as a sinusoidal function of the gap $i - j$. Note that this is in stark contrast to the absolute positional encodings of the original Transformer, and instead matches the relative encodings in Shaw et al. (2018).

Node and edge features Finally, we obtain an aggregate edge encoding vector $\boldsymbol{e}_{ij}$ by concatenating the structural encodings $\boldsymbol{e}_{ij}^{(s)}$ and the positional encodings $\boldsymbol{e}_{ij}^{(p)}$. We only include edges in the k-nearest neighbors graph of $\mathcal{X}$, with $k = 30$ for all experiments.

For node features, we compute the three dihedral angles of the protein backbone $(\phi_i, \psi_i, \omega_i)$ and embed these on the 3-torus as $\{\sin, \cos\} \times (\phi_i, \psi_i, \omega_i)$.

2.2 STRUCTURED TRANSFORMER

In this work, we introduce a *Structured Transformer* model that draws inspiration from the self-attention based *Transformer* model (Vaswani et al., 2017). First, since L is often very large (several hundred amino acids being common), global all-vs-all attention leads to prohibitive computational and memory bottlenecks. We avert these bottlenecks by restricting attention for each node i to the set $\mathrm{N}(i, k)$ of its k-nearest neighbors in 3D space. Since our architecture is multilayered, iterated local attention can derive progressively more global estimates of context for each node i. Second, unlike the standard Transformer, we also include edge features to embed the spatial and positional dependencies in deriving the attention. Thus, our model generalizes Transformer to spatially structured settings.

Encoder Our encoder module is designed as follows. A transformation $\boldsymbol{W}_h : \mathbb{R}^{d_v} \mapsto \mathbb{R}^d$ produces initial embeddings $\boldsymbol{h}_i = \boldsymbol{W}_h(\boldsymbol{v}_i)$ from the node features $\boldsymbol{v}_i$ pertaining to position $i \in [N] \triangleq \{1, 2, \ldots, N\}$. Each layer of the encoder implements a multi-head self-attention component, where head $\ell \in [L]$ attends to a separate subspace of the embeddings. We write $\boldsymbol{h}_i = \text{Concat}\left(\boldsymbol{h}_i^{(1)}, \ldots, \boldsymbol{h}_i^{(L)}\right)$, where each $\boldsymbol{h}_i^{(\ell)} \in \mathbb{R}^{d/L}$, to emphasize the subspaces. We refine the embeddings via additional learned transformations. Specifically, $\boldsymbol{W}_q^{(\ell)}$ maps $\boldsymbol{h}_i^{(\ell)}$ to *query* embeddings $\boldsymbol{q}_i^{(\ell)}$, $\boldsymbol{W}_z^{(\ell)}$ maps pairs $(\boldsymbol{h}_j^{(\ell)}, \boldsymbol{e}_{ij})$ to *key* embeddings $\boldsymbol{z}_{ij}^{(\ell)}$ for $j \in \mathrm{N}(i, k)$, and $\boldsymbol{W}_v^{(\ell)}$ maps the same pairs $(\boldsymbol{h}_j^{(\ell)}, \boldsymbol{e}_{ij})$ to *value* embeddings $\boldsymbol{v}_{ij}^{(\ell)}$ for each $i \in [N], \ell \in [L]$. Decoupling the mappings for keys and values allows each to depend on different subspaces of the representation.

We compute the similarity of key $\boldsymbol{z}_{ij}^{(\ell)}$ to query $\boldsymbol{q}_i^{(\ell)}$ as a function of their scaled inner product:

$$a_{ij}^{(\ell)} = \frac{\exp(m_{ij}^{(\ell)})}{\sum\limits_{j' \in \mathrm{N}(i,k)} \exp(m_{ij'}^{(\ell)})}, \qquad \text{where} \quad m_{ij}^{(\ell)} = \frac{\boldsymbol{q}_i^{(\ell)\top} \boldsymbol{z}_{ij}^{(\ell)}}{\sqrt{d}}.$$

This similarity is interpreted as the importance of $\boldsymbol{v}_{ij}^{(\ell)}$ in refining $\boldsymbol{h}_i^{(\ell)}$, and we obtain the updates

$$\boldsymbol{h}_i^{(\ell)} \leftarrow \boldsymbol{h}_i^{(\ell)} + \sum_{j \in N(i,k)} a_{ij}^{(\ell)} \boldsymbol{v}_{ij}^{(\ell)}.$$

Thus, effectively, we refine embeddings in each subspace separately via output of a residual connection at each layer. We alternate between these self-attention layers and position-wise feedforward

---

[2]We used 16 Gaussian RBFs isotropically spaced from 0 to 20 Angstroms.

[3]We represent quaternions in terms of their vector of real coefficients.

Table 1: Null perplexities

Null model	Perplexity	Conditioned on
Uniform	20.00	-
Natural frequencies	17.83	Random position in a natural protein
Pfam HMM profiles	11.64	Specific position in a specific protein family

layers as in the original Transformer (Vaswani et al., 2017). We stack multiple layers atop each other, and thereby obtain continually refined embeddings as we traverse the layers bottom up. The encoder yields the embeddings produced by the topmost layer as its output.

Decoder   Our decoder module is laid out similarly but for some important differences. First, we augment the edge information e_{ij} with s_j, i.e., the jth amino acid in the output sequence. This combination is then propagated bottom up in the decoder stack analogous to the role of e_{ij} during encoding. Second, unlike encoding, we perform decoding *autoregressively*, i.e., conditioned on the partial sequence already decoded. Therefore, while training, we mask the information pertaining to the neighbor nodes subsequent to the position being decoded. Finally, as in the Transformer decoder (Vaswani et al., 2017), the embeddings output by the encoder are treated as keys that dictate, in part, the embeddings output by the decoder. The general layout of our structural decoder, modulo the influx of augmented edge information and restriction to k-neighborhood, is similar to the decoder in the original Transformer setting (Vaswani et al., 2017) so we omit the details.

2.3   TRAINING

Dataset   To evaluate the ability of the models to generalize across different protein folds, we collected a dataset based on the CATH hierarchical classification of protein structure (Orengo et al., 1997). For all domains in the CATH 4.2 40% non-redundant set of proteins, we obtained full chains up to length 500 (which may contain more than one domain) and then cluster-split these at the CATH topology level (i.e. fold level) into training, validation, and test sets at an 80/10/10 split. Chains containing multiple CATH tpologies were purged with precedence for test over validation over train. Our splitting procedure ensured that no two domains from different sets would share the same topologies (folds). The final splits contained 18025 chains in the training set, 1637 chains in the validation set, and 1911 chains in the test set.

Optimization   We trained models using the learning rate schedule and initialization of (Vaswani et al., 2017), a dropout (Srivastava et al., 2014) rate of 10%, and early stopping based on validation perplexity.

Table 2: Per-residue perplexities for test sets (lower is better).

Test set	Short	Single chain	All
Structure-conditioned models			
Structured Transformer (ours)	9.07	9.54	7.04
SPIN2 ($\lambda = 0.001$)	11.81	12.50	-
SPIN2 ($\lambda = 0.01$)	11.52	12.08	-
SPIN2 ($\lambda = 0.1$)	13.62	13.95	-
Language models			
Structured Transformer, no encoder	16.03	16.36	16.98
RNN ($h = 128$)	16.08	16.34	16.93
RNN ($h = 256$)	16.09	16.32	16.93
RNN ($h = 512$)	16.01	16.34	16.94
Test set size	94	107	1911

3 RESULTS

Many protein sequences may reasonably design the same 3D structure (Li et al., 1996), and so we focus on likelihood-based evaluations of model performance. Specifically, we evaluate the perplexity per letter of test protein folds (topologies) that were held out from the training and validation sets.

Protein perplexities What kind of perplexities might be useful? To provide context, we first present perplexities for some simple models of protein sequences in Table 1. The amino acid alphabet and its natural frequencies upper-bound perplexity at 20 and ∼17.8, respectively. Random protein sequences under these null models are unlikely to be functional without further selection (Keefe & Szostak, 2001). First order profiles of protein sequences such as those from the Pfam database (El-Gebali et al., 2018), however, are widely used for protein engineering. We found the average perplexity per letter of profiles in Pfam 32 (ignoring alignment uncertainty) to be ∼11.6. This suggests that even models with high perplexities of this order have the potential to be useful models for the space of functional protein sequences.

The importance of structure We found that there was a significant gap between unconditional language models of protein sequences and models conditioned on structure. Remarkably, for a range of structure-independent language models, the typical test perplexities turned out to be ∼16-17 (Table 2), which were barely better than null letter frequencies (Table 1).

All structure-based models were observed to have a considerably lower perplexity. In particular, our Structured Transformer model attained a perplexity of ∼7 on the full test set.

Improvement over profile-based methods We also compared to a recent method SPIN2 that predicts, using deep neural networks, protein sequence profiles given protein structures (O'Connell et al., 2018). Since SPIN2 is computationally intensive (minutes per protein for small proteins) and was trained on complete proteins rather than chains, we evaluated it on two subsets of the full test set: a 'Small' subset of the test set containing chains up to length 100 and a 'Single chain' subset containing only those models where the single chain accounted for the entire protein record in the Protein Data Bank. Both subsets discarded any chains with structural gaps. Additionally, since SPIN2 sometimes output zero-probability predictions truncated by round-off error, we added a small pseudocount λ to its predictions at varying strengths to avoid infinite perplexities. We found that our Structured Transformer model considerably improved upon the perplexities of SPIN2 (specifically, an improvement on the order of ∼2-2.5).

4 CONCLUSION

We presented a new deep generative model to 'design' protein sequences given a graph specification of their structure. Our model augments the traditional sequence-level self-attention of Transformers (Vaswani et al., 2017) with relational 3D structural encodings and is able to leverage the spatial locality of dependencies in molecular structures for efficient computation. When evaluated on unseen folds, the model achieves significantly improved perplexities over the state-of-the-art parametric generative models. Our framework suggests the possibility of being able to efficiently design and engineer protein sequences with structurally-guided deep generative models, and underscores the central role of modeling sparse long-range dependencies in biological sequences.

REFERENCES

Mohammed AlQuraishi. End-to-end differentiable learning of protein structure. *bioRxiv*, pp. 265231, 2018.

Namrata Anand and Possu Huang. Generative modeling for protein structures. In *Advances in Neural Information Processing Systems*, pp. 7505–7516, 2018.

Sivaraman Balakrishnan, Hetunandan Kamisetty, Jaime G Carbonell, Su-In Lee, and Christopher James Langmead. Learning generative models for protein fold families. *Proteins: Structure, Function, and Bioinformatics*, 79(4):1061–1078, 2011.

Jacob B Bale, Shane Gonen, Yuxi Liu, William Sheffler, Daniel Ellis, Chantz Thomas, Duilio Cascio, Todd O Yeates, Tamir Gonen, Neil P King, et al. Accurate design of megadalton-scale two-component icosahedral protein complexes. *Science*, 353(6297):389–394, 2016.

Peter W Battaglia, Jessica B Hamrick, Victor Bapst, Alvaro Sanchez-Gonzalez, Vinicius Zambaldi, Mateusz Malinowski, Andrea Tacchetti, David Raposo, Adam Santoro, Ryan Faulkner, et al. Relational inductive biases, deep learning, and graph networks. *arXiv preprint arXiv:1806.01261*, 2018.

Surojit Biswas, Gleb Kuznetsov, Pierce J Ogden, Nicholas J Conway, Ryan P Adams, and George M Church. Toward machine-guided design of proteins. *bioRxiv*, pp. 337154, 2018.

Wouter Boomsma and Jes Frellsen. Spherical convolutions and their application in molecular modelling. In *Advances in Neural Information Processing Systems*, pp. 3433–3443, 2017.

Sara El-Gebali, Jaina Mistry, Alex Bateman, Sean R Eddy, Aurélien Luciani, Simon C Potter, Matloob Qureshi, Lorna J Richardson, Gustavo A Salazar, Alfredo Smart, et al. The pfam protein families database in 2019. *Nucleic acids research*, 47(D1):D427–D432, 2018.

Justin Gilmer, Samuel S Schoenholz, Patrick F Riley, Oriol Vinyals, and George E Dahl. Neural message passing for quantum chemistry. In *Proceedings of the 34th International Conference on Machine Learning-Volume 70*, pp. 1263–1272. JMLR. org, 2017.

Joe G Greener, Lewis Moffat, and David T Jones. Design of metalloproteins and novel protein folds using variational autoencoders. *Scientific reports*, 8(1):16189, 2018.

Cheng-Zhi Anna Huang, Ashish Vaswani, Jakob Uszkoreit, Noam Shazeer, Curtis Hawthorne, Andrew M Dai, Matthew D Hoffman, and Douglas Eck. An improved relative self-attention mechanism for transformer with application to music generation. *arXiv preprint arXiv:1809.04281*, 2018.

Po-Ssu Huang, Scott E Boyken, and David Baker. The coming of age of de novo protein design. *Nature*, 537(7620):320, 2016.

Du Q Huynh. Metrics for 3d rotations: Comparison and analysis. *Journal of Mathematical Imaging and Vision*, 35(2):155–164, 2009.

John Ingraham, Adam Riesselman, Chris Sander, and Debora Marks. Learning protein structure with a differentiable simulator. In *International Conference on Learning Representations*, 2019. URL https://openreview.net/forum?id=Byg3y3C9Km.

Anthony D Keefe and Jack W Szostak. Functional proteins from a random-sequence library. *Nature*, 410(6829):715, 2001.

Nobuyasu Koga, Rie Tatsumi-Koga, Gaohua Liu, Rong Xiao, Thomas B Acton, Gaetano T Montelione, and David Baker. Principles for designing ideal protein structures. *Nature*, 491(7423):222, 2012.

Brian Kuhlman, Gautam Dantas, Gregory C Ireton, Gabriele Varani, Barry L Stoddard, and David Baker. Design of a novel globular protein fold with atomic-level accuracy. *science*, 302(5649): 1364–1368, 2003.

Hao Li, Robert Helling, Chao Tang, and Ned Wingreen. Emergence of preferred structures in a simple model of protein folding. *Science*, 273(5275):666–669, 1996.

Debora S Marks, Lucy J Colwell, Robert Sheridan, Thomas A Hopf, Andrea Pagnani, Riccardo Zecchina, and Chris Sander. Protein 3d structure computed from evolutionary sequence variation. *PloS one*, 6(12):e28766, 2011.

Faruck Morcos, Andrea Pagnani, Bryan Lunt, Arianna Bertolino, Debora S Marks, Chris Sander, Riccardo Zecchina, José N Onuchic, Terence Hwa, and Martin Weigt. Direct-coupling analysis of residue coevolution captures native contacts across many protein families. *Proceedings of the National Academy of Sciences*, 108(49):E1293–E1301, 2011.

James O'Connell, Zhixiu Li, Jack Hanson, Rhys Heffernan, James Lyons, Kuldip Paliwal, Abdollah Dehzangi, Yuedong Yang, and Yaoqi Zhou. Spin2: Predicting sequence profiles from protein structures using deep neural networks. *Proteins: Structure, Function, and Bioinformatics*, 86(6): 629–633, 2018.

Christine A Orengo, AD Michie, S Jones, David T Jones, MB Swindells, and Janet M Thornton. Cath–a hierarchic classification of protein domain structures. *Structure*, 5(8):1093–1109, 1997.

Adam J Riesselman, John B Ingraham, and Debora S Marks. Deep generative models of genetic variation capture the effects of mutations. *Nat. Methods*, 15:816–822, 2018.

Gabriel J Rocklin, Tamuka M Chidyausiku, Inna Goreshnik, Alex Ford, Scott Houliston, Alexander Lemak, Lauren Carter, Rashmi Ravichandran, Vikram K Mulligan, Aaron Chevalier, et al. Global analysis of protein folding using massively parallel design, synthesis, and testing. *Science*, 357 (6347):168–175, 2017.

Peter Shaw, Jakob Uszkoreit, and Ashish Vaswani. Self-attention with relative position representations. In *Proceedings of the 2018 Conference of the North American Chapter of the Association for Computational Linguistics: Human Language Technologies, Volume 2 (Short Papers)*, volume 2, pp. 464–468, 2018.

Justin B Siegel, Alexandre Zanghellini, Helena M Lovick, Gert Kiss, Abigail R Lambert, Jennifer L St Clair, Jasmine L Gallaher, Donald Hilvert, Michael H Gelb, Barry L Stoddard, et al. Computational design of an enzyme catalyst for a stereoselective bimolecular diels-alder reaction. *Science*, 329(5989):309–313, 2010.

Sam Sinai, Eric Kelsic, George M Church, and Martin A Nowak. Variational auto-encoding of protein sequences. *arXiv preprint arXiv:1712.03346*, 2017.

Nitish Srivastava, Geoffrey Hinton, Alex Krizhevsky, Ilya Sutskever, and Ruslan Salakhutdinov. Dropout: a simple way to prevent neural networks from overfitting. *The Journal of Machine Learning Research*, 15(1):1929–1958, 2014.

Jérôme Tubiana, Simona Cocco, and Rémi Monasson. Learning protein constitutive motifs from sequence data. *arXiv preprint arXiv:1803.08718*, 2018.

Ashish Vaswani, Noam Shazeer, Niki Parmar, Jakob Uszkoreit, Llion Jones, Aidan N Gomez, Łukasz Kaiser, and Illia Polosukhin. Attention is all you need. In *Advances in Neural Information Processing Systems*, pp. 5998–6008, 2017.

Jingxue Wang, Huali Cao, John ZH Zhang, and Yifei Qi. Computational protein design with deep learning neural networks. *Scientific reports*, 8(1):6349, 2018.

Maurice Weiler, Mario Geiger, Max Welling, Wouter Boomsma, and Taco Cohen. 3d steerable cnns: Learning rotationally equivariant features in volumetric data. In *Advances in Neural Information Processing Systems*, pp. 10402–10413, 2018.

Kevin K Yang, Zachary Wu, and Frances H Arnold. Machine learning in protein engineering. *arXiv preprint arXiv:1811.10775*, 2018.

Jianfu Zhou, Alexandra E Panaitiu, and Gevorg Grigoryan. A general-purpose protein design framework based on mining sequence-structure relationships in known protein structures. *bioRxiv*, pp. 431635, 2018.

5 APPENDIX

Figure 2: **Example structures from the dataset.**