DIRECT MOLECULAR CONFORMATION GENERATION

ABSTRACT

Molecular conformation generation, which is to generate 3 dimensional coordinates of all the atoms in a molecule, is an important task for bioinformatics and pharmacology. Most existing machine learning based methods first predict interatomic distances and then generate conformations based on them. This two-stage approach has a potential limitation that the predicted distances may conflict with each other, e.g., violating the triangle inequality. In this work, we propose a method that directly outputs the coordinates of atoms, so that there is no violation of constraints. The conformation generator of our method stacks multiple blocks, and each block outputs a conformation which is then refined by the following block. We adopt the variational auto-encoder (VAE) framework and use a latent variable to generate diverse conformations. To handle the roto-translation equivariance, we adopt a loss that is invariant to rotation and translation of molecule coordinates, by computing the minimal achievable distance after any rotation and translation. Our method outperforms strong baselines on four public datasets, which shows the effectiveness of our method and the great potential of the direct approach. The code is released at `https://github.com/DirectMolecularConfGen/DMCG`.

1 INTRODUCTION

Molecular conformation generation aims to generate the 3D coordinates of all the atoms of molecules, which then can be used in molecular property prediction (Axelrod & Gomez-Bombarelli, 2021), docking (Roy et al., 2015), structure-based virtual screening (Kontoyianni, 2017), etc. Molecular conformation can be physically observed using X-ray crystallography, but it is prohibitively costly for industry-scale tasks (Mansimov et al., 2019). *Ab initio* methods, such as density functional theory (DFT) (Parr, 1980; Baseden & Tye, 2014), can accurately predict the molecular shapes, but take up to several hours per small molecule (Hu et al., 2021).To handle large-scale molecules, people turn to leverage the classical force fields, like UFF (Rappe et al., 1992) or MMFF (Halgren, 1996), to estimate conformations, which is efficient but unacceptably inaccurate (Kanal et al., 2018).

Recently, people start to explore machine learning methods to generate conformation. Simm & Hernández-Lobato (2020), Shi et al. (2020) and Shi et al. (2021) leveraged variational auto-encoder (VAE), flow-based models and score-based methods for conformation generation, respectively. With learned models, molecule conformations can be sampled independently (Simm & Hernández-Lobato, 2020; Shi et al., 2020) or using Langevin dynamics (Xu et al., 2021; Shi et al., 2021). The common part of these methods is that they are all built upon the interatomic distances among atoms (i.e., the distance between atom pairs). Specifically, Simm & Hernández-Lobato (2020); Shi et al. (2020); Xu et al. (2021) use various generative methods to model the distribution of interatomic distances, and then reconstruct conformations based on distances. Shi et al. (2021) leveraged a score-matching network to model the density gradient of interatomic distances. A major reason of using distance-based methods is that the interatomic distances are invariant to rotation and translation of the conformation. However, a potential drawback is that the predicted distances might conflict with each other. For example, they did not explicitly consider the triangle inequality for distances among any three atoms. The underlying degree-of-freedom of these distances is only $3N - 6$ (N is the number of atoms in a molecule) while practically, these methods often generate distances with the degree-of-freedom of roughly $N(N-1)/2$, which usually lead to violations of the triangle inequality. For example, we found that in GraphDG (Simm & Hernández-Lobato, 2020), a prevailing representative of distance-based method, 8.65% of molecular graphs in the GEOM-Drugs test set will produce distance matrix that violates the triangle inequality.

In this work, we explore the possibility to directly generate the coordinates of all atoms. This is a straightforward and more natural choice and avoids violations of the triangle inequality, and has demonstrated remarkable performance on protein structure prediction by the AlphaFold 2 (Jumper et al., 2021). We design a model that generates atom coordinates directly. The generator in our model stacks multiple blocks, and each block outputs a conformation which is then refined by the following block. A block consists of several modules that can encode the previous conformation as well as the features of bonds, atoms and global information. At the end of each block, we add a normalization layer that centers the coordinates at the origin. Since a molecule may have multiple conformations, we use the variational auto-encoder (VAE) framework which allows diverse generation. To realize roto-translation equivariance, i.e., rotating and translating a set of atom coordinates do not change the conformation, we adopt a loss that is invariant to rotation and translation of atom coordinates.

We conduct experiments on four benchmark settings, which are GEOM-QM9 and GEOM-Drugs with small-scale setting (Shi et al., 2021) and large-scale setting (Axelrod & Gomez-Bombarelli, 2021). Compared with previous methods, ours achieves state-of-the-art results on GEOM-Drugs (both small-scale and large-scale settings), and outperforms almost all previous baselines on GEOM-QM9, demonstrating the effectiveness of our method.

Our contributions are summarized as follows:

(1) We explore a new direction of conformation generation, that directly generates the coordinates of a molecular conformation without generating interatomic distances. We empirically show that directly generating conformation achieves state-of-the-art results on several tasks.

(2) We leverage a fine-grained loss function for training, that is invariant to the roto-translation of the conformation.

(3) We propose a new model that iteratively refines the conformations. Our model is inspired by multiple advanced architectures like GATv2 (Brody et al., 2021), graph network (GN) block (Battaglia et al., 2018), that can effectively model molecules.

2 PRELIMINARIES

In this section, we introduce the notations used in this work, give a formal definition of the molecular conformation generation problem, and briefly introduce how to align two conformations under rotation and translation.

Notations: Let $G = (V, E)$ denote a molecular graph, where V and E are collections of atoms and bonds, respectively. Specifically, $V = \{v_1, v_2, \cdots, v_{|V|}\}$ with the i-th atom v_i. Let e_{ij} denote the bond between atom v_i and v_j. Let $N(i)$ denote the neighbors of atom i, i.e., $N(i) = \{v_j \mid e_{ij} \in E\}$. We use R to represent the conformation of G, where $R \in \mathbb{R}^{|V| \times 3}$. The i-th row of R is the coordinate of atom v_i.

Let $\rho(\cdot)$ denote a roto-translation operation, i.e. an affine transformation $\rho(R) = R\boldsymbol{t} + \boldsymbol{b}$ where the orientation-preserving orthogonal transformation $\boldsymbol{t} \in \mathrm{SO}(3) \subset \mathbb{R}^{3 \times 3}$ represents a rotation and the vector $\boldsymbol{b} \in \mathbb{R}^{1 \times 3}$ represents a translation. Let $\mathrm{MLP}(\cdots)$ denote the multi-layer perception network, where the inputs are concatenated together and then processed by linear mapping, Batch Normalization (Ioffe & Szegedy, 2015) and ReLU activation sequentially.

Problem definition: Given a graph $G = (V, E)$, our task is to learn a mapping, that can output the coordinates R of all atoms in V, i.e., $R \in \mathbb{R}^{|V| \times 3}$.

Matching loss: Let R_1 and R_2 denote two conformations, both of which are $N \times 3$ matrices (N is the number of rows in R_1 and R_2). The matching loss between R_1 and R_2 is defined as follows:

$$\ell_M(R_1, R_2) = \min_{\rho} \|\rho(R_1) - R_2\|_F^2, \tag{1}$$

where $\| \cdot \|_F$ denotes the Frobenius norm, i.e., $\|A\|_F^2 = \sum_{i,j} |A_{ij}|^2$. The matching loss is invariant to the rotation and translation of either of the two input conformations. It is obviously nonnegative, and is zero if and only if R_1 and R_2 represent the same molecule structure.

Karney (2007) proposed to use quaternions to solve Eqn.(1). A quaternion q is an extension of complex numbers, $q = q_0\mathbf{u} + q_1\mathbf{i} + q_2\mathbf{j} + q_3\mathbf{k}$, where q_0, q_1, q_2, q_3 are scalars and $\mathbf{u}, \mathbf{i}, \mathbf{j}, \mathbf{k}$ are

orientation vectors. With quaternions, any rotation operation is specified by a 3×3 matrix, where each element in the matrix is the basic algebra of q_0 to q_3 and some constants. The solution to Eqn.(1) is the minimal eigenvalue of a 4×4 matrix obtained by the basic algebra on R_1 and R_2, the corresponding optimal rotation and translation ρ^* can be obtained from the eigenvectors. Note that the best ρ^* is related to both R_1 and R_2. To stabilize training, we set the gradients of ρ^* w.r.t. the model parameter as zero. More details can be found in Karney (2007).

3 OUR METHOD

We introduce our proposed method in this section.

3.1 CONFORMATION GENERATION FRAMEWORK

Our goal can be formalized as learning to generate samples from the conditional distribution $p(R|G)$. We follow the variational auto-encoder (VAE) scheme (Kingma & Welling, 2014; Rezende et al., 2014; Sohn et al., 2015) as it enables flexible model architecture (vs. flow-based models), efficient (i.e., i.i.d) generation (vs. score-matching methods), and stable training and diverse sample generation (vs. adversarial methods). Given a molecule graph G, the conditional variant of VAE generates R using a likelihood model $p(R|z, G)$ where z is a latent code drawn from a prior $p(z) = \mathcal{N}(0, \boldsymbol{I})$. The corresponding $p(R|G) = \int p(z)p(R|z, G)\,\mathrm{d}z$ is however intractable to evaluate and maximize to fit data. VAE handles this by introducing a (conditional) inference model $q(z|R, G)$ which leads to the following identity:

$$\log p(R|G) = \mathbb{E}_{q(z|R,G)}\left[\log p(R|z,G)\right] - D_{\mathrm{KL}}(q(z|R,G)\|p(z)) + D_{\mathrm{KL}}(q(z|R,G)\|p(z|R,G)), \tag{2}$$

where D_{KL} denotes the Kullback-Leibler (KL) divergence, and $p(z|R, G) = \frac{p(z)p(R|z,G)}{\int p(z)p(R|z,G)\,\mathrm{d}z}$ is the true posterior. Due to the nonnegativity of the third term, the first two terms form a lower bound of $\log p(R|G)$. When $q(z|R, G)$ is properly chosen, the bound is tractable to optimize. Moreover, as the left-hand-side is independent of $q(z|R, G)$, tightening the bound (i.e., minimizing the third term) can be achieved by maximizing the bound w.r.t $q(z|R, G)$. Therefore, as a loss function to be minimized, the objective is:

$$\mathbb{E}_{q(z|R,G)}[-\log p(R|z,G)] + D_{\mathrm{KL}}(q(z|R,G)\|p(z)). \tag{3}$$

Now we develop our specification for the conformation generation task. We construct the likelihood model with regard to a decoded reference conformation $\hat{R}(z, G)$:

$$p(R|z,G) \propto \exp\left\{-\frac{1}{\gamma}\ell_M\big(\hat{R}(z,G), R\big)\right\}, \tag{4}$$

where $\gamma > 0$ is a variance parameter that we choose to fix as 1 (see Sec. 3.4 (1) for explanation). Note that the usage of ℓ_M guarantees all roto-translation equivalent conformations have the same probability. For the inference model, we choose

$$q(z|R,G) = \mathcal{N}(z|\mu_{R,G}, \Sigma_{R,G}), \tag{5}$$

where the conditional mean and variance are outputs from some encoder. It enables tractable loss optimization via reparameterization: $z \sim q(z|R, G)$ is equivalent to $z = \mu_{R,G} + \Sigma_{R,G}\epsilon$ where $\epsilon \sim \mathcal{N}(0, \boldsymbol{I})$. With the above specification, the loss function becomes:

$$\min \mathbb{E}_{\epsilon \sim \mathcal{N}(0,\boldsymbol{I})}\ell_M(\hat{R}(\mu_{R,G} + \Sigma_{R,G}\epsilon, G), R) + \beta D_{\mathrm{KL}}(\mathcal{N}(\mu_{R,G}, \Sigma_{R,G})\|\mathcal{N}(0, \boldsymbol{I})), \tag{6}$$

where the minimization is applied over all model parameters. Note that in Eqn.(6), an additional hyperparameter β is introduced in the spirit of β-VAE (Higgins et al., 2016) to handle the strength of the prior regularization. The second term in Eqn.(6) has a closed-form expression for optimization.

3.2 WORKFLOW

According to Eqn.(3), we need a module φ_{3D} to model $q(z|G, R)$, a module φ_{dec} to model $p(R|z, G)$ and a module φ_{2D} to model the conditional input G. In this subsection, we introduce the general

Figure 1: The workflow of our method. Green and orange lines represent $p(R|z, G)$ and $q(z|R, G)$ respectively.

workflow of how they work together (see Figure 1). Details of the $\varphi_{...}$ modules will be covered in the next subsection.

The training workflow is shown as follows:

(1) The encoder φ_{2D} takes the molecular graph G as its input, and outputs d-dimensional graph representations $H_V^{(0)} \in \mathbb{R}^{|V| \times d}$ for all atoms, $H_E^{(0)} \in \mathbb{R}^{|E| \times d}$ for all bonds, a global graph feature $U^{(0)} \in \mathbb{R}^d$, and initial conformation $\hat{R}^{(0)} \in \mathbb{R}^{|V| \times 3}$. Formally, $(H_V^{(0)}, H_E^{(0)}, U^{(0)}, \hat{R}^{(0)}) = \varphi_{2D}(G)$.

(2) We use another encoder φ_{3D} to extract features of the conformation R for constructing the conditional inference model $q(z|R, G)$. According to the above specification, this encoder only needs to output the mean and variance of the Gaussian, or formally, $(\mu_{R,G}, \Sigma_{R,G}) = \varphi_{3D}(R, G)$.

(3) We randomly sample a variable z from the Gaussian distribution $\mathcal{N}(\mu_{R,G}, \Sigma_{R,G})$, and then feed $H_V^{(0)}, H_E^{(0)}, U^{(0)}, \hat{R}^{(0)}, z$ into the decoder φ_{dec} to obtain the conformation $\hat{R}$. That is, $\hat{R}(z, G) = \varphi_{dec}(\varphi_{2d}(G), z) = \varphi_{dec}(H_V^{(0)}, H_E^{(0)}, \hat{R}^{(0)}, U^{(0)}, z)$. Note that sampling $z \sim \mathcal{N}(\mu_{R,G}, \Sigma_{R,G})$ is equivalent to sampling $\epsilon \sim \mathcal{N}(0, \boldsymbol{I})$ and then setting $z = \mu_{R,G} + \Sigma_{R,G}\epsilon$.

(4) After obtaining $\hat{R}(z, G)$ and and $\mathcal{N}(\mu_{R,G}, \Sigma_{R,G})$, we use Eqn.(6) as the objective function for training. Remind that $\hat{R}$ is related to $\varphi_{2D}, \varphi_{3D}, \varphi_{dec}$, and $\mu_{R,G}, \Sigma_{R,G}$ are related to φ_{3D}.

For the inference workflow, there are three steps in total: (1) Given a molecular graph G, we use φ_{2D} to encode G and obtain $\hat{R}^{(0)}, H_V^{(0)}, H_E^{(0)}, U^{(0)}$; (2) sample a random variable z from Gaussian $\mathcal{N}(0, \boldsymbol{I})$; (3) feed $\hat{R}^{(0)}, H_V^{(0)}, H_E^{(0)}, U^{(0)}, z$ into φ_{dec} and obtain the eventual conformation $\hat{R}(z, G)$. Note that φ_{3D} is not used in inference phase.

3.3 DETAILED MODEL ARCHITECTURE

The encoders $\varphi_{2D}, \varphi_{3D}$ and the decoder φ_{dec} share the same architecture. They are all stacks of L identical blocks. We introduce the l-th block in the decoder φ_{dec}, and leave the introduction of φ_{2D} and φ_{3D} in Appendix A. In contrast to previous work, we introduce some novel designs that handle the task at a finer level. (1) The conformation is iteratively refined by each decoding block. Specifically, we use a normalization technique that moves the center of each decoded conformation to the origin. (2) We use more advanced model components, like attention models (Brody et al., 2021) and the GN block (Battaglia et al., 2018), that can more effectively represent the molecules.

The detailed architecture is shown in Figure 2. The inputs include the following information output by the $(l-1)$-th block: the conformation $\hat{R}^{(l-1)}$, atom features $H_V^{(l-1)} \in \mathbb{R}^{|V| \times 3}$, edge features $H_E^{(l-1)}$ and the global feature $U^{(l-1)}$. Note $H_V^{(0)}, H_E^{(0)}, U^{(0)}$ and $\hat{R}^{(0)}$ are the outputs of φ_{2D}. $\hat{R}^{(l-1)}$ is first mapped by MLP and get $F^{(l)} = \text{MLP}(\hat{R}^{(l-1)})$. The atom features are then fused with the coordinate features by $H_V^{(l-1)} \leftarrow H_V^{(l-1)} + F^{(l)} + z$, where $z \sim \mathcal{N}(\mu_{R,G}, \Sigma_{R,G})$.

We use a variant of the GN block (Battaglia et al., 2018) as the backbone of the model due to its superior performance in molecular modeling. In each block, we update the bond features, atom features and global features sequentially. Each block will output its own prediction on the coordinates of atoms. For ease of reference, we use $h_i^{(l)}$ (single subscript) to denote the feature of atom i output by the l-th block, and use $h_{ij}^{(l)}$ (two subscripts) to denote the bond features between atom i and j output by the l-th block. The workflow of the l-th block is shown as follows:

Figure 2: Network architecture of the l-th block. Yellow dashed boxes are inputs, orange boxes denote the operations, and green boxes are outputs.

(1) *Update bond features*: For each bond feature $h_{ij}^{(l-1)}$ in $H_E^{(l-1)}$, it is updated by

$$h_{ij}^{(l)} = h_{ij}^{(l-1)} + \text{MLP}(h_i^{(l-1)}, h_j^{(l-1)}, h_{ij}^{(l-1)}, U^{(l-1)}). \tag{7}$$

(2) *Update atom features*: The atom features are updated using an attentive way. For any $i \in [|V|]$,

$$\bar{h}_i^{(l)} = \sum_{j \in \mathcal{N}(i)} \alpha_j W_v \text{concat}(h_{ij}^{(l)}, h_j^{(l-1)});$$

$$\alpha_j \propto \exp(\boldsymbol{a}^\top \text{LeakyReLU}(W_q h_i^{(l-1)} + W_k \text{concat}(h_j^{(l-1)}, h_{ij}^l))); \tag{8}$$

$$h_i^{(l)} = h_i^{(l-1)} + \text{MLP}\left(h_i^{(l-1)}, \bar{h}_i^{(l)}, U^{(l-1)}\right).$$

In Eqn.(8), $\boldsymbol{a}$, W_q, W_v and W_k are the parameters to be learned and $\text{concat}(\cdot, \cdot)$ is the concatenation of two vectors. For atom v_i, we first use GATv2 (Brody et al., 2021) to aggregate the features from its connected bonds and obtain $\bar{h}_i$, and then update v_i based on $\bar{h}_i^{(l)}$, $h_i^{(l-1)}$ and $U^{(l-1)}$.

(3) *Update global features*: The global feature is updated as follows:

$$U^{(l)} = U^{(l-1)} + \text{MLP}\left(\frac{1}{|V|}\sum_{i=1}^{|V|} h_i^{(l)}, \frac{1}{|E|}\sum_{i,j} h_{ij}^{(l)}, U^{(l-1)}\right). \tag{9}$$

(4) *Output the prediction*: After obtaining the new features of the l-th block, for any atom $i \in [|V|]$, it predicts the conformations $\hat{R}_i^{(l)}$ as follows:

$$\bar{R}_i^{(l)} = \text{MLP}(h_i^{(l)}), \quad m^{(l)} = \frac{1}{|V|}\sum_{j=1}^{|V|} \bar{R}_j^{(l)}, \quad \hat{R}_i^{(l)} = \bar{R}_i^{(l)} - m^{(l)} + \hat{R}_i^{(l-1)}. \tag{10}$$

An important step in Eqn.(10) is that, after making initial prediction $\bar{R}_i^{(l)}$, we calculate its center and normalize their coordinates by moving the center to the origin. We add this kind of normalization to ensure that input coordinates of each block are in reasonable numeric ranges.

We use $\hat{R}^{(L)}$ output by the last block in φ_{dec} as the final prediction of the conformation.

3.4 DISCUSSIONS

In this section, we discuss the relation between our method and two previous work.

Comparison with CVGAE: Mansimov et al. (2019) developed an early attempt to generate conformation by raw coordinates, but its performance is not as desired as distance-based methods developed afterwards (Shi et al., 2020; Simm & Hernández-Lobato, 2020). Our method, pursuing the same spirit, makes several finer designs and implementations.

(1) Under the VAE formulation, Mansimov et al. (2019) also leaned the variance parameter γ of the likelihood model. i.e., the γ in Eqn.(4). However, a decent VAE analysis (Dai & Wipf, 2019, Thm. 3) revealed that the VAE objective prefers a vanishing γ. This is desired in the ideal case when the encoder and decoder are optimized perfectly (Dai & Wipf, 2019, Thm. 4), but in practice it would distract the optimizer towards a "lazy way" to focus on γ and stagnate the optimization of the encoder and decoder. Therefore, we choose to fix γ. When γ is fixed, the loss function is equivalent to using $\gamma = 1$ and a β parameter as the formulation in Eqn.(6). We hence choose a small value of β corresponding to fixing γ to a small value in the original formulation (i.e. without β) as advocated (Dai & Wipf, 2019).

(2) Our model consists of more advanced modules, including GATv2 (Brody et al., 2021) and GN block (Battaglia et al., 2018) to better model the input. In comparison, Mansimov et al. (2019) mainly leveraged GRU (Bahdanau et al., 2015) and its variant on graphs, which is outperformed by the modules used in our model.

(3) We iteratively refine the output of each block, while (Mansimov et al., 2019) only outputs the conformation in the last layer.

Comparison with ConfGF: Shi et al. (2021) used score matching to model the gradients w.r.t interatomic distances, and then recover the coordinates based on the gradients. That is, (Shi et al., 2021) still tries to model the interatomic distances, while we completely abandon modeling the distances. We propose a new direction for molecular conformation generation. In addition, (Shi et al., 2021) is not so efficient as our method, since it relies on Langevin dynamics, which requires much more sampling cost.

4 EXPERIMENT

4.1 SETTINGS

Datasets: Following prior works (Xu et al., 2021; Shi et al., 2021), we use the GEOM-QM9 and GEOM-Drugs datasets (Axelrod & Gomez-Bombarelli, 2021) for conformation generation. We verify our method on both small-scale setting and large-scale setting. First, we use the same datasets provided by Shi et al. (2021) for fair comparison with prior works. The training, validation and test sets of the two datasets consist of 200K, 2.5K and 22408 (for GEOM-QM9)/14324 (for GEOM-Drugs) molecule-conformation pairs respectively. After that, we sample larger datasets from the original GEOM to validate the scalability of our method. We use all data in GEOM-QM9 and $2.2M$ molecule-conformation pairs for GEOM-Drugs. The numbers of training, validation and test sets for the larger GEOM-QM9 setting are 1.37M, 165K and 174K, and those for larger GEOM-Drugs are 2M, 100K and 100K.

Model configuration: All of φ_{2D}, φ_{3D} and φ_{dec} have six blocks. The dimension d of the features is 256. Inspired by the feed-forward layer in Transformer (Vaswani et al., 2017), `MLP` also consists of two sub-layers, where the first one maps the input features from dimension 256 to 1024, followed by Batch Normalization and ReLU activation. Then the 1024-dimension feature is mapped to 256 again using linear mapping.

Training: We use AdamW optimizer (Loshchilov & Hutter, 2019) with initial learning rate $\eta_0 = 2 \times 10^{-4}$ and weight decay 0.01. In the first 4000 iterations, the learning rate is linearly increased from 10^{-6} to 2×10^{-4}. After that, we use cosine learning rate scheduler (Loshchilov & Hutter, 2016), where the learning rate at the t-th iteration is $\eta_0(1 + \cos(\pi\frac{t}{T}))/2$, where T is the half of the period (i.e., the iteration numbers of 10 epochs in our setting). Similarly, we also use the cosine scheduler to dynamically set the β at range $[0.0001, 0.008]$. The batch size is fixed as 128. All models are trained for 100 epochs. For the two small-scale settings, the experiments are conducted on a single V100 GPU. For the two large-scale settings, we use two V100 GPUs for experiments.

Evaluation: Assume in the test set, the molecule x has N_x conformations. Following Shi et al. (2020; 2021), for each molecule x in the test set, we generate $2N_x$ conformations. Let $\mathbb{S}_g(\mathcal{G})$ and $\mathbb{S}_r(\mathcal{G})$ denote all generated and groundtruth conformations respectively. We use coverage score (COV) and matching score (MAT) to evaluate the generation quality. Mathematically,

$$\text{COV}(\mathbb{S}_g(\mathcal{G}), \mathbb{S}_r(\mathcal{G})) = \frac{1}{|\mathbb{S}_r|} |\{\boldsymbol{R} \in \mathbb{S}_r \mid \ell_M(\boldsymbol{R}, \boldsymbol{R}') < \delta, \exists \boldsymbol{R}' \in \mathbb{S}_g\}|; \tag{11}$$

$$\text{MAT}(\mathbb{S}_g(\mathcal{G}), \mathbb{S}_r(\mathcal{G})) = \frac{1}{|\mathbb{S}_r|} \sum_{\boldsymbol{R}' \in \mathbb{S}_r} \min_{\boldsymbol{R} \in \mathbb{S}_g} \ell_M(\boldsymbol{R}, \boldsymbol{R}'), \tag{12}$$

where ℓ_M is defined in Eqn.(1). A good method should have a high COV score and a low MAT score. Following (Shi et al., 2021; Xu et al., 2021), the δ's are set as 0.5 and 1.25 for QM9 and Drugs, respectively.

Baselines: (1) RDKit, which is a widely used toolkit and generates the conformation based on the force fields; (2) CVGAE (Mansimov et al., 2019), which is an early attempt to generate raw coordinates; (3) GraphDG (Simm & Hernández-Lobato, 2020), a representative distance-based method

with VAE; (4) CGCF (Xu et al., 2021), which is another distance-based method leveraging continuous normalizing flow; (5) ConfGF (Shi et al., 2021), which uses score matching to generate the gradients w.r.t. distances and then recover the conformation. Note that ConfGF leverages Langevin dynamics, which requires much more additional decoding time.

4.2 RESULTS

The results on the small-scale datasets are reported in Table 1, and those on large-scale datasets are in Table 2.

Dataset	QM9				Drugs			
Methods	COV(%)↑		MAT (Å)↓		COV(%)↑		MAT (Å)↓	
	Mean	Median	Mean	Median	Mean	Median	Mean	Median
RDkit	83.26	90.78	0.3447	0.2935	60.91	65.70	1.2026	1.1252
CVGAE	0.09	0.00	1.6713	1.6088	0.00	0.00	3.0702	2.9937
GraphDG	73.33	84.21	0.4245	0.3973	8.27	0.00	1.9722	1.9845
CGCF	78.05	82.48	0.4219	0.3900	53.96	57.06	1.2487	1.2247
ConfGF	**88.49**	94.13	**0.2673**	0.2685	62.15	70.93	1.1629	1.1596
Ours	81.44	**96.61**	0.2879	**0.2428**	**77.78**	**86.09**	**1.0657**	**1.0563**

Table 1: Experimental results on small-scale datasets.

Dataset	QM9				Drugs			
Methods	COV(%)↑		MAT (Å)↓		COV(%)↑		MAT (Å)↓	
	Mean	Median	Mean	Median	Mean	Median	Mean	Median
RDkit	81.61	85.71	0.2643	0.2472	69.42	77.45	1.0880	1.0333
GraphDG	13.48	5.71	0.9511	0.9180	1.95	0.00	2.6133	2.6132
ConfGF	**89.21**	95.12	0.2809	0.2837	70.92	85.71	1.0940	1.0917
Ours	86.58	**100.00**	**0.2457**	**0.1756**	**90.68**	**100.00**	**0.8934**	**0.8703**

Table 2: Experimental results on large-scale datasets.

We have the following observations:

(1) On the four settings in Table 1 and Table 2, our method achieves state-of-the-art results on most of them, i.e., the two large-scale settings and small dataset setting on GEOM-drug. The only exception is that our method is not so good as ConfGF in terms of the COV mean and MAT mean on small-scale QM9, and the mean COV score on large-scale QM9. These results already showed the effectiveness and scalability of our method.

(2) Our method achieves more improvement on molecules with more heavy atoms. Take the results in Table 1 as an example. On average, QM9 and Drugs have 8.8 and 24.9 heavy atoms respectively. In terms of MAT median, on QM9, our method improves ConfGF by 0.026 points, while on Drugs, the improvement is 0.103. To verify our conjecture, on Drugs, we categorize the molecules based on their numbers of heavy atoms. The number of heavy atoms in the i-th group lie in $[10i+1, 10(i+1)]$. We compare our method against ConfGF and GraphDG. The results are in Table 3.

Metric	COV(%)↑				MAT(Å)↓			
	$i = 1$	$i = 2$	$i = 3$	average	$i = 1$	$i = 2$	$i = 3$	average
ConfGf	99.95	66.28	15.34	62.54	0.7764	1.1510	1.5345	1.1637
GraphDG	15.11	1.78	0.0	3.12	2.0578	2.5863	2.9849	2.5847
Ours	98.13	80.18	51.72	77.78	0.8502	1.0552	1.2737	1.0657

Table 3: COV and MAT mean scores w.r.t. numbers of heavy atoms on small-scale GEOM-Drugs. The i indicates that the heavy atom number lies in range $[10i + 1, 10(i + 1)]$, $i \in \{1, 2, 3\}$.

We can observe that among the three algorithms, ConfGF achieves the best results on the molecules whose numbers of heavy atoms lie in $[11, 20]$, and our method is slightly worse than it. However, when the numbers of heavy atoms are larger than 21, our method significantly outperforms ConfGF

Figure 3: Visualization of different conformations.

and GraphDG. This is consistent with our conjecture, and also demonstrates the potential of directly generating coordinates for complex molecules with more heavy atoms.

(3) Our method is much more sample efficient than ConfGF. Based on the implementation of ConfGF, generating a conformation requires 5000 sequential forward steps. With our method, we only need to sample one variable from $\mathcal{N}(0, \boldsymbol{I})$ and then generate a corresponding conformation by forwarding once.

For a fair comparison, following the official implementation of ConfGF, we split the test sets of small-scale GEOM-QM9 and GEOM-Drugs into 200 batches. ConfGF requires 8511.60 and 11830.42 seconds to decode QM9 and Drugs test sets, while our method only requires 32.68 and 54.89 seconds respectively. That is, our method speeds up the decoding more than 200 times, which shows the effectiveness of our method.

In Figure 3, we visualize the conformation of different methods. We randomly select four molecules from the small-scale GEOM-drug dataset, generate several conformations, and visualize the best-aligned ones with the groundtruth. We can see that our method can generate high-quality conformations than previous methods, which are the most similar to the groundtruth.

4.3 PROPERTY PREDICTION

In addition to conformation generation task, we also conduct experiments on property prediction task, which is to predict molecular property based on an ensemble of generated conformation (Axelrod & Gomez-Bombarelli, 2021). We first randomly choose 30 molecules from QM9 and Drugs test sets, and then sample 50 conformations for each molecule using RDkit, ConfGF and our method as initial coordinates. Then we will use MMFF function in RDkit to optimize the generated coordinates and quantum chemical calculation package Psi4 (Smith et al., 2020) to calculate the energy for each conformation. We also calculate energy for each groundtruth conformation of these molecules in the same way, as the groundtruth energy. Finally, we use the mean absolute error (MAE) of the average energy (denoted as "Average E") and the lowest energy (denoted as "Lowest E") among these conformations to measure the accuracy of each method on ensemble property prediction task.

Methods	QM9		Drugs	
	Average E	Lowest E	Average E	Lowest E
RDkit	0.0507	0.0276	0.1720	0.1255
ConfGF	0.0458	0.0229	2.7344	0.1379
GraphDG	0.0859	0.0567	5.6440	0.3520
Ours	0.0418	0.0213	0.1573	0.1129

Table 4: MAE of property prediction based on the generated conformations.

Methods	COV(%)↑		MAT (Å)↓	
	Mean	Median	Mean	Median
Ours	**77.78**	**86.09**	**1.0657**	**1.0563**
Ours w/o attention	72.18	82.76	1.1257	1.1138
Ours w/o conformation normalization	61.82	64.76	1.2045	1.1974
Ours w/ FF	85.00	92.05	0.8959	0.8992

Table 5: Ablation study on small-scale GEOM-Drugs.

The results are shown in Table 4. Our method outperforms all previous baselines. On QM9, our method achieves slightly better results than ConfGF, while on Drugs, the results are significantly better than ConfGF and GraphDG. This shows that our method can provide good conformations for property prediction tasks.

4.4 ABLATION STUDY

To further verify our algorithm, we conduct the following ablation study: (1) We remove the attentive node aggregation; instead, it is replaced by a simple `MLP` network. That is, Eqn.(8) is replaced by

$$h_i^{(l)} = h_i^{(l-1)} + \texttt{MLP}(h_i^{(l-1)}, U^{(l-1)}, \frac{1}{|\mathcal{N}(i)|} \sum_{j \in \mathcal{N}(i)} h_j^{(l-1)}); \qquad (13)$$

(2) We remove the normalization step in Eqn.(10), i.e., the $m^{(l)}$ is not used; (3) We further use the force field in RDKit to refine the output of our method. The results are summarized in Table 5. We can see that: (1) Without attentively aggregating the atom features, the performances drops: The mean COV drops 5.6 points and MAT score increases 0.06 points. (2) Without the conformation normalization, the performance is greatly effected, which shows the importance of this step; (3) Our method can still be refined by the RDKit, which shows that our method can be combined with the classic methods for more improvements.

Finally, we compute the COV and MAT scores of $\hat{R}^{(l)}$ against the groundtruth, which is the output conformation of the l-th block in the decoder. $\hat{R}^{(0)}$ is the output of φ_{2D}. The results are shown in Figure 4. We can see that iteratively refining the conformations can improve the performances, which shows the effectiveness of our design.

Figure 4: MAT and COV scores of $\hat{R}^{(l)}$ against the grondtruth.

5 CONCLUSIONS AND FUTURE WORK

In this work, we propose a new method, that directly generates the coordinates of conformations. For this purpose, we leverage a loss function that is invariant to roto-translation, and design a new model with many advanced modules (i.e., GATv2, GN block) that can iteratively refine the conformations. Experimental results on both small-scale and large-scale GEOM-QM9 and GEOM-Drugs demonstrate the effectiveness of our method.

For future work, there are many interesting directions. First, we will combine with more generative methods like flow-based model, Langevin dynamics and so on. Second, during case analyzing, we found that there are still some cases that current methods cannot successfully handle (e.g., rotatable rings) and we will improve them in the future. Third, current methods are mainly non-autoregressive, where all coordinates are generated simultaneously. We will study the autoregressive setting so as to further improve the accuracy.

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A   DETAILS OF φ_{2D} AND φ_{3D}

The model architectures of φ_{2D} and φ_{3D} are similar to φ_{dec}, with the following differences.

Comparing φ_{2D} with φ_{dec}, the differences are the initial conformation $\hat{R}^{(0)}$ and initial features (i.e., the $H_V^{(0)}$, $H_E^{(0)}$ and $U^{(0)}$). φ_{2D} takes a random conformation sampled from uniform distribution in $[-1, 1]$ as input. The initial atom and edge features are the embeddings of the atoms and edges respectively. φ_{2D} will also output a prediction of the conformation. Note that the random variable z sampled from Gaussian $\mathcal{N}(\mu_{R,G}, \Sigma_{R,G})$ is not used in φ_{2D}.

Comparing φ_{3D} with φ_{dec}, the differences are the initial conformation $\hat{R}^{(0)}$, initial features (i.e., the $H_V^{(0)}$, $H_E^{(0)}$ and $U^{(0)}$) too. φ_{3D} takes the groundtruth conformation as input. The initial atom and edge features are the embeddings of the atoms and edges respectively. Another difference is that the fourth step of φ_{dec}, i.e., Eqn.(10), is not used. More details are available in our code.