FROM SALT-AND-PEPPER TO PINWHEELS: NEURAL MECHANISMS FOR EFFICIENT AND ROBUST VISUAL CODING

Anonymous authors
Paper under double-blind review

ABSTRACT

Extensive experimental studies have shown that in lower mammals, neuronal orientation preference in the primary visual cortex is organized in a disordered "salt-and-pepper" pattern. In contrast, higher-order mammals display a continuous variation in orientation preference, forming structured pinwheel-like patterns. Despite these observations, the spiking mechanisms underlying the emergence of these distinct topological structures and their functional roles in visual processing remain poorly understood. To address this, we developed a self-evolving spiking neural network model with Hebbian plasticity, trained using physiological parameters characteristic of rodents, cats, and primates, including retinotopy, neuronal morphology, and connectivity patterns. Our results identify critical factors, such as the degree of input visual field overlap, neuronal density, and the balance between localized connectivity and long-range competition, that determine the emergence of either salt-and-pepper or pinwheel-like topologies. Furthermore, we demonstrate that pinwheel structures exhibit lower wiring cost and enhanced sparse coding capabilities compared to salt-and-pepper organizations. They also maintain greater coding robustness against noise in naturalistic visual stimuli. These findings suggest that such topological structures confer significant computational advantages in visual processing and highlight their potential application in the design of brain-inspired deep learning networks and algorithms.

1 INTRODUCTION

In the primary visual cortex (V1) of visually-advanced mammals, seminal works (Hubel & Wiesel, 1962; 1974; Blasdel & Salama, 1986; Bonhoeffer & Grinvald, 1991) revealed that neurons are organized into distinct spatial clusters, forming “pinwheel” structures centered around singularities within the orientation preference map. This organization contrasts with that of some mammals, such as rodents, which exhibit “salt-and-pepper” organizations, characterized by randomly interspersed neurons with varying orientation preferences, or “mini-columns” that show only weak spatial clustering, revealed by studies(Girman et al., 1999; Ringach et al., 2016; Bargmann & Newsome, 2014; Kaschube et al., 2010). Despite these structural differences, the spiking mechanisms that give rise to these species-specific patterns, and their roles in visual processing, remain poorly understood.

Existing computational models, such as self-organizing maps (SOM) (Kohonen, 1982), ON-OFF theoretical models (Miller, 1994; Jang et al., 2020a; Song et al., 2021; Najafian et al., 2022), and various artificial neural networks (Margalit et al., 2023; Chizhov & Graham, 2021) have provided insights into the generation of orientation preference maps through mathematical abstractions or firing rate models. However, these approaches often lack the biological realism of spiking neural networks, which better capture the intricate temporal dynamics of real neuronal systems. Furthermore, many existing SNN models (Lufkin et al., 2022; Chizhov & Graham, 2021; Srinivasa & Jiang, 2013) struggle to reproduce the emergent properties of orientation preference maps over time, as they often overlook key anatomical and physiological factors, such as shared visual inputs (Zhang et al., 2018; Jang et al., 2020a; Najafian et al., 2022), cortical size (Meng et al., 2012), localized neuronal densities (Weigand et al., 2017), and genetically encoded connectivity patterns (Blauch et al., 2022).

In response to these challenges, our research contributes the following:

- We propose a retina-V1 realistic self-evolving spiking neural network (SESNN) that integrates comprehensive data on retinotopy, neuronal morphology, and connectivity from rats, cats and primates. This model is capable of dynamically simulating cortical structure formation across species using Hebbian-like learning rules and natural scene inputs. By leveraging visual field overlap and neuronal density, the SESNN accurately distinguishes between salt-and-pepper and pinwheel cortical patterns.
- Pinwheel structures arise from local synaptic plasticity rules in spiking neural networks, where lateral connections are strengthened between neurons with similar stimulus responses and weakened between those with dissimilar responses. This selective connectivity forms iso-orientation domains with consistent orientation tuning. Near pinwheel centers, connections become more homogeneous, while neurons maintain diverse orientation preferences and dynamic firing patterns. This balance between connectivity and neural diversity enables efficient, sparse coding of both simple and complex visual stimuli, optimizing visual information processing.
- We further demonstrate that pinwheel organizations exhibit superior information-cost efficiency, increased coding sparsity, and greater robustness to noise compared to salt-and-pepper patterns. These properties enhance the detection and coding of varied orientation features, indicating that the spatial structure of pinwheels promotes highly efficient and reliable visual processing.

2 RETINA-V1 TOPOLOGICAL SELF-EVOLVING SPIKING NEURAL NETWORK

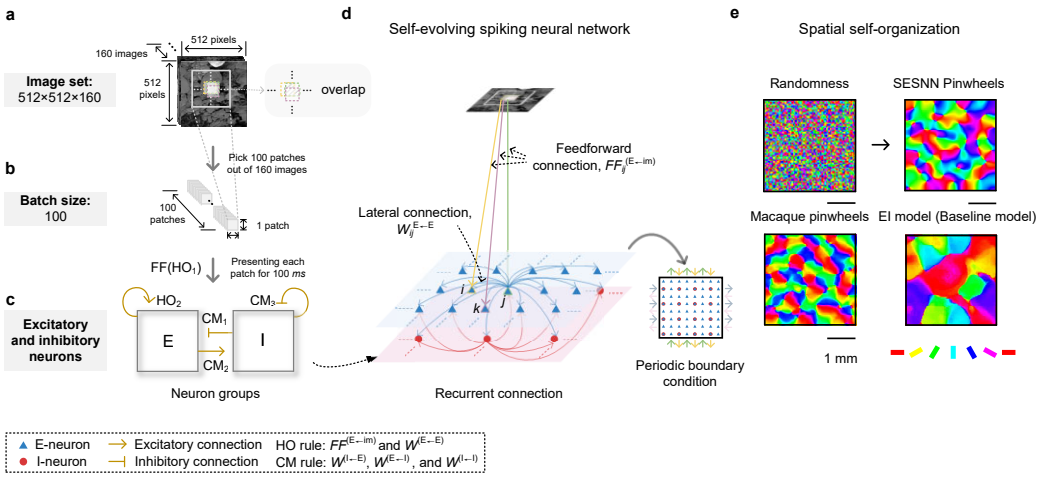

Figure 1: Architecture of proposed SESNN model. **a**. The image set with 160 whitened natural images. **b**. Stimulus images are randomly selected from the image set and presented to E neurons for 100 ms. **c**. The connections between excitatory (E-) and inhibitory (I-) neurons follow HO and CM rules. **d**. Recurrent connection among neurons. The spatial arrangement and periodic boundary conditions of E- and I- neurons. E- and I- neurons share the same spatial coordinates, and the boundaries are linked together according to the arrows shown in the diagram. Arrows of the same color represent identical connections. **e**. Spatial self-organization within the neuronal population post-training and the comparison among the SESNN model's orientation preference map, macaque's orientation preference map and SNN-based model (baseline) (Srinivasa & Jiang, 2013). The color bars represent orientation preferences, and the scale bar corresponds to 1 mm on the V1 cortical surface. This comparative analysis is detailed in Table 1.

2.1 STIMULUS TRAINING SETS

We use 20 original whitened images which reduce pairwise correlations among pixels, mimicking the processing of the lateral geniculate nucleus ((Dan et al., 1996; Zylberberg et al., 2011; Paul D. King et al., 2013)). To ensure these images encompassed orientation details, each image is rotated

Table 1: SESNN pinwheels vs. macaque pinwheels. (mean ± SD) for SESNN results.

Metric	E-I baseline	SESNN (macaque)	Macaque
Pinwheel density (pinwheels/Λ^2)	~ 2.941	3.175 ± 0.397	~ 3.327
Nearest-neighbor pinwheel distance (mm)	N/A	0.277 ± 0.043	~ 0.242
Hypercolumn size (mm)	N/A	0.839 ± 0.054	~ 0.760

clockwise every 90-degree clockwise rotation and a subsequent flip, producing 8 variations per original image. This method generates a total of 160 images comprising our training dataset (refer to Fig. 1a).

In each trial, all E-neurons process visual input from 100 random patches, each presented for 100 ms(see Fig. 1b). Each E-neuron perceives a small patch of 16×16. The visual fields of the E-neurons are modeled as overlapping on the retina (see Fig. 1a), with the total visual input dimension being $[(16-\varepsilon)\sqrt{n}+\varepsilon]$ pixels on each side (n is neuron number, visual overlap metric ε). To address the visual overlap among adjacent neurons, which adheres to periodic boundary conditions, the input design features a wraparound effect to ensure continuity and uniformity throughout the neural network.

2.2 Neural Model and the Plasticity Rules

The SESNN model includes 4900 E- and 1225 I- neurons, forming a recurrent network with an E-I ratio of 4:1 ((Alreja et al., 2022; Tao et al., 2004; Pfeffer et al., 2013)). E-neurons are stimulated by natural images and receive background noise from other brain areas, while I-neurons indirectly process natural image stimuli through their interactions with E-neurons. The spiking dynamics of the network are simulated the classical leaky integrate-and-fire neurons ((Stevens et al., 2013)), which also feature refractory periods of 3 ms and adaptive firing thresholds ((Földiák, 1990)). The detailed description of the neuronal model can be found in the appendix A.1.

The learning process utilizes two primary forms of Hebbian-like learning rules: Hebbian Oja's variant (HO) and Correlation Measuring (CM) ((Oja, 1982; Zylberberg et al., 2011; Paul D. King et al., 2013)). Consistent with experimental evidence Holmgren et al. (2003); Hofer et al. (2011), V1 pyramidal neurons exhibit weaker synaptic strengths compared to other types, crucial for avoiding over-excitation and ensuring neural balance. To accurately model these self-evolving dynamics within our SESNN, we implement E-E connections governed by the HO rule Oja (1982), with a normalization factor (the second term in Eq. 1) to regulate synaptic weight growth within a 0 to 1 range. In contrast, lateral connections between excitatory and inhibitory neurons are governed by the CM rule, which inherently allows for stronger synaptic weights without normalization (Zylberberg et al. (2011); Paul D. King et al. (2013)), enabling these connections to better capture correlations between input patterns and neuronal responses. This distinction in learning rules is crucial for maintaining functional balance and enhancing representational efficiency within the network. Specifically, the HO rule learns the principal component of input patterns, allowing E-E connections to align with the largest variance in the input space. While this supports self-organized structure formation, such as oriented receptive fields, it can lead to redundancy across excitatory and inhibitory neurons. By comparison, the CM rule captures correlations between input activity and a neuron's response, ensuring that predictable components of excitatory activity are expressed in inhibitory neurons. These components can then be suppressed through E-I-E loops, a mechanism that supports predictive coding by minimizing redundancy and promoting sparse, efficient neural representations. ~~These rules allow the SESNN to adaptively modify synaptic weights based on firing pattern correlations, emulating fundamental learning processes in biological neural networks.~~ Both HO and CM rules support long-term potentiation and depression, functioning similarly to other rate-based learning rules that do not require precise spike timing. We selected the HO and CM rules for their minimal tuning requirements and their ability to stabilize recurrent excitation.

Following every 100 ms of stimulus presentation, synaptic weight changes ΔW within the network are computed:

$$\text{HO}: \ \Delta W_{ij}^{(\text{K}\leftarrow\text{K}^*)} \propto y_i x_j - y_i^2 W_{ij}^{(\text{K}\leftarrow\text{K}^*)}, \tag{1}$$

$$\text{CM}: \ \Delta W_{ij}^{(\mathrm{K}\leftarrow\mathrm{K}^*)} \propto y_i x_j - \langle y_i \rangle \langle x_j \rangle \left(1 + W_{ij}^{(\mathrm{K}\leftarrow\mathrm{K}^*)}\right), \tag{2}$$

where x, y refer to the spike rate of presynaptic (j) and postsynaptic neurons (i) and K, K^* determine the type of neurons (E or I), the operator $\langle\cdot\rangle$ indicates the lifetime average value. Following each 100 ms stimulus presentation, we compute the network's neuronal instantaneous spike rates as exponential moving averages, which accumulate spikes over time (see Eq. 3). Exponential moving averages are utilized to track recent neuronal activity levels. Concurrently, lifetime average values are also calculated using exponential moving averages, which are crucial for maintaining homeostatic stability. This method helps stabilize the neural network by adjusting neuronal properties or synaptic strengths to sustain consistent activity levels over time.

$$x_j(t) = (1-\zeta)x_j(t-1) + \zeta \cdot z_j(t), \tag{3}$$

where $\zeta = 1 - e^{-\frac{1}{10}}$, indicating that the 10 ms is a temporal window of the moving average weighted with exponential decay. The initialization of x_j is 0. The exponential moving average is calculated dynamically and updated along with synaptic weights. $z_j^{(\mathrm{K}^*)}(t) = 1$ represents the spike output from neuron j at time t, and 0 otherwise.

$$\langle x_j \rangle := (1-\xi) \cdot \langle x_j \rangle + \xi \cdot \overline{x}_j, \tag{4}$$

where $\xi = 1 - e^{-1}$. It is dynamically updated to ensure the sum of the weights remains constant over time. Eq. 3 captures the short-term firing rate of neurons by tracking recent activity through a short time window, while Eq. 4 computes the long-term average to monitor activity trends and maintain homeostatic stability, together ensuring the network's adaptability and balanced dynamics over multiple timescales. $\overline{x}_j$ represents the average firing rate of neuron j in one trial. The hyperparameters of the SESNN model are as follows: the learning rates are $\eta_{\mathrm{FF}} = 0.2$ (image to E-neuron), $\eta_{\mathrm{EE}} = 0.01$ (E- to E-neuron), $\eta_{\mathrm{EI}} = 0.7$ (I- to E-neuron), $\eta_{\mathrm{II}} = 1.5$ (I- to I-neuron), and $\eta_{\mathrm{IE}} = 0.7$ (E- to I-neuron), while the neural connectivity parameters are $\alpha_{\text{max, E}} = 1.0$ (E- max weight) and $\alpha_{\text{max, I}} = 0.5$ (I- max weight). These learning rate settings are crucial for stabilizing the training of the neural network. Specifically, setting a slower learning rate for E-E connections than for others helps prevent over-excitation among E-neurons. This approach is consistent with empirical findings Hofer et al. (2011); Holmgren et al. (2003); Sato et al. (2016).

2.3 Experimental Data-Justified Neural Connectivity in a 2D Cortical Area

E- and I- neurons are arranged symmetrically on a two-dimensional lattice within SESNN. The initial connection weights between neurons are modeled by a Gaussian function of Euclidean distance, which can be expressed as:

$$W_0^{\mathrm{K}\leftarrow\mathrm{K}'}(i,j) = \alpha_{\mathrm{K}'} \times \exp\left(\frac{-d(i,j)^2}{2{\sigma_{\mathrm{K}'}}^2}\right), \tag{5}$$

where $d(i,j)$ represents the Euclidean distance from neuron i to neuron j in a grid, α determines the maximum connection weight, which is set to $\alpha_{\mathrm{EE}} = 1$, $\alpha_{\mathrm{EI}} = 1$, $\alpha_{\mathrm{IE}} = 0.5$, $\alpha_{\mathrm{II}} = 0.5$, and σ governs the rate at which the weight decays with distance. The synaptic types predominantly determine the parameters for this connection weight distribution function. To accurately replicate the neuronal architecture of V1 in macaques. The synaptic connection range, denoted by σ, are set to $\sigma_{\mathrm{EE}} = 3.5$, $\sigma_{\mathrm{EI}} = 2.9$, $\sigma_{\mathrm{IE}} = 2.6$, $\sigma_{\mathrm{II}} = 2.1$. These values are based on anatomical data indicating that the axon lengths of E- and I-neurons are approximately 200 μm and 100 μm, respectively, while the dendrite lengths are around 150 μm for E-neurons and 75 μm for I-neurons in the V1 ((Tao et al., 2004; Stepanyants et al., 2009; Amatrudo et al., 2012)). This careful selection of parameters and neuron distribution ensures our model's biological fidelity, particularly in representing synaptic connectivity and neuronal density. We discard any connection strengths below a threshold of 0.01 to maintain computational efficiency and biological plausibility.

Given that the majority of RFs are concentrated in the area centralis Van Essen et al. (1984); Born et al. (2015), which projects to a disproportionately large area of V1, we focus our analysis solely on RFs within this central visual field (see table 2). To control for variables, we use a constant RF size near the central vision, where eccentricity is close to zero.

The anatomical data for neural connectivity is based on the findings of (Tao et al., 2004; Stepanyants et al., 2009; Amatrudo et al., 2012). The plasticity rule settings are based on the findings of (Holmgren

et al., 2003; Hofer et al., 2011). For retinotopic data, we refer to (Srinivasan et al., 2015; Tehovnik & Slocum, 2007; Scholl et al., 2013; Veit et al., 2014; Engelmann & Peichl, 1996; Weigand et al., 2017; Law et al., 1988; Huberman et al., 2006; Niell & Stryker, 2008; van Beest et al., 2021; Foik et al., 2020). The anatomical data on orientation preference maps is sourced from (Najafian et al., 2022). The details of the data usage and visual input overlap calculation can be found in the appendix A.2,A.3,A.4.

Table 2: Comparative anatomical data of the retina and V1 across species.

Species	V1 neurons density (neurons/mm^2)	V1 RF size (deg)	CMF (mm/deg)	Pinwheel numbers (per mm^2)
Macaque	243,000^a	0.2^b	2.0^b	8
Cat	99,200^a	1.0^c	0.67^d	3
Mouse	86,600^a	4.0^e	0.02^f	N/A

^{a} Srinivasan et al. (2015) ^{b} Tehovnik & Slocum (2007) ^{c} Scholl et al. (2013) ^{d} Veit et al. (2014) ^{e} Niell & Stryker (2008) ^{f} van Beest et al. (2021)

3 SESNN REVEALS THAT VISUAL OVERLAP BETWEEN NEURONS IS CRITICAL FOR V1 ORIENTATION MAP FORMATION

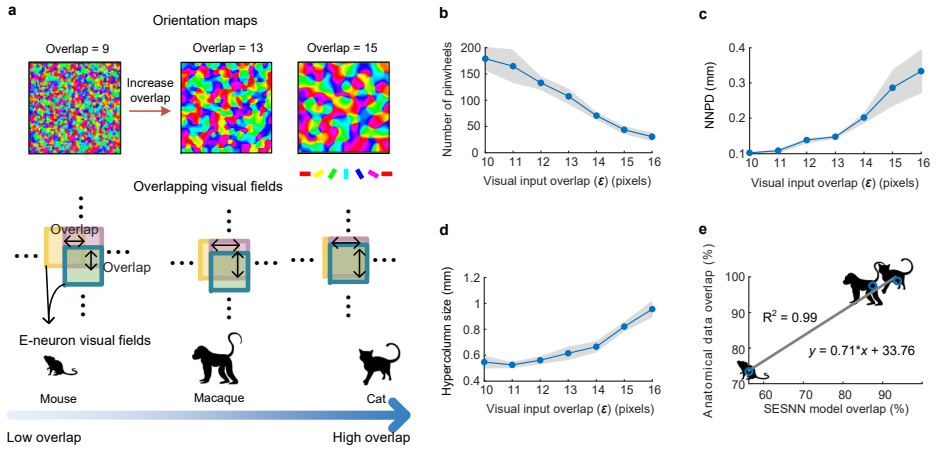

Figure 2: The overlap of visual fields among adjacent neurons contributes to the formation of pinwheel structures. **a**. The diverse overlapping extent of visual fields in neurons forms various iso-orientation domains of species in the orientation preference maps when perceiving natural images. The increasing overlap is consistent with the transition from lower to higher mammals (see the animal sketches). **b**. The correlation between visual overlap and pinwheel counts. **c**. The relationship between visual overlap and NNPD. **d**. The hypercolumn size varies with different visual overlaps. **e**. Comparing the SESNN model overlap with actual anatomical data overlap percentages in different species (mice, cats, and macaques). Color scheme: orientation preference. All data: mean ± SD.

For quantitative analysis of orientation maps, we utilize metrics including pinwheel density (pinwheels/mm^2), pinwheel counts ((Kaschube et al., 2010; Stevens et al., 2013)) (Fig. 2b), nearest-neighbor pinwheel distance (NNPD) (mm) ((Müller et al., 2000; Kaschube et al., 2010)) (Fig. 2c), and hypercolumn size (mm) ((Stevens et al., 2013)) (Fig. 2d). Pinwheel density is defined as the number of pinwheels per V1 surface area mm^2. NNPD refers to the shortest Euclidean distance between pinwheel centers. Pinwheel centers are identifiable via 2D Fast Fourier Transform ((Kaschube et al., 2010)), located where the real and imaginary components of the transform intersect at zero ((Stevens et al., 2013)). A hypercolumn is defined as a region with periodicity measured by 2D FFT. It's noteworthy that pinwheel density is excluded from the Fig. 2 analysis, because, regardless of

hypercolumn size, the observed pinwheel density consistently approaches π pinwheels/Λ^2(Fig. 3d), conforming to topological constraints ((Stevens et al., 2013; Kaschube et al., 2010)). This paper does not account for left- and right-eye dominance columns, so the hypercolumn size is defined as the full 180° cycle of repeating column spacing (Λ) (mm).

Our SESNN model effectively generates various orientation selectivity patterns, from salt-and-peppers to pinwheel structures. We modify the overlap parameter ε to range from 10 to 15 pixels (see Fig. 2). This visualization highlights how variations in the degree of overlap in visual stimuli can influence the density and arrangement of pinwheel structures in the visual cortex. Notably, setting the overlap to $\varepsilon_2 = 9$ pixels results in the model producing salt-and-pepper organizations (see the first panel of Fig. 2a), rather than the anticipated pinwheel structures. Thus, it is evident that above the threshold of overlap (in our scenarios, $threshold_\varepsilon = 10$ pixels), pinwheel structures emerge. Specifically, we find that a greater overlap (e.g., $\varepsilon_2 = 15$ pixels) fosters the formation of stronger local clusters, resulting in larger hypercolumn sizes (Fig. 2d), a reduced number of pinwheels (Fig. 2b), and extended NNPDs (Fig. 2c), compared with the scenario of a smaller overlap (e.g., $\varepsilon_1 = 12$ pixels). These findings suggest that visual input overlap critically determines the species-specific organization patterns of V1. The overlap degrees (ε) in visual fields vary across species (including macaques, cats, rodents, etc.) (Fig. 2e) to accommodate the size of iso-orientation domains in the visual cortex representation as observed by the study of (Najafian et al., 2022). When the overlap of visual stimuli is large, multiple neurons receive similar visual inputs. This causes these neurons to form stronger local connections through Hebbian plasticity. Neurons with similar receptive fields tend to develop highly consistent local connections, and their orientation preferences gradually align, creating a region of locally continuous orientation preferences. Additionally, Neuronal density is another factor that impacts the size of hypercolumns. Although this has been reported previously Philips et al. (2017), we have included additional details in Appendix A.3.2 for completeness.

3.1 THE EMERGENCE OF THE PINWHEEL STRUCTURE AND MECHANISM

We investigate the development of orientation preference maps in V1 of animals at eye-opening using the SESNN model, where the initial visual experience of natural stimuli are retinotopically mapped to V1 neurons (Fig. 3a) ((Stevens et al., 2013)). Our comparative analysis with empirical ferret data ((Stevens et al., 2013; Chapman et al., 1996; Chapman & Stryker, 1993)) shows a high correlation ($r = 0.98$, Pearson correlation) for orientation preference maps observed from postnatal day 33 (P33) to P41 when they reach high stability (Fig. 3b). By normalizing the time scale, we are able to align the SESNN model's developmental trajectory with ferret empirical data. To quantitatively assess how the stability change over time, we use the orientation stability index (SI) ((Chapman et al., 1996)), calculated as follows:

$$\text{SI} = 1 - \frac{4}{n\pi} \sum_{i=1}^{n} \left| (F_i - O_i) \bmod \left(\frac{\pi}{2}\right) \right|. \tag{6}$$

The SI is computed by iterating over all n data points, where i indexes individual pixels, F_i represents the final orientation preference value of the neurons, and O_i denotes the orientation preference at a prior developmental stage. The higher SI at later development stage of the orientation preference maps implying that maps remain unchanged during their continued exposure to varying natural stimuli (Fig. 3b). Furthermore, we also investigated whether the model's performance would change if trained with a new set of images after stabilization. Our results show that while some detailed aspects of the pinwheel structure altered slightly, the overall locations of the pinwheel centers and the domains remained largely unchanged, indicating strong stability. The changes in neuron tuning toward specific orientations were also observed (detailed in Appendix A.5). Additionally, we examine the relationship between dimensionless pinwheel densities (ρ) and hypercolumn areas (Λ^2) across the SESNN model, ferrets, galagos, and tree shrews, comparing findings with empirical data (Fig. 3d).

Considering the trained connections W_{ij}, we follow the hypothesis that the information capacity can be depicted by the entropy of weight distribution in the complete connection matrix (distribution of all the trained connection strength) and the energy cost of synaptic transmission is inversely proportional to the connection strength. We have the Information-Cost Efficiency ICE defined as:

$$\text{ICE} = \frac{H}{C}; H = -\sum_{k} P_k \log\left(P_k\right); C = \sum_{i,j} \frac{1}{W_{ij}}, \tag{7}$$

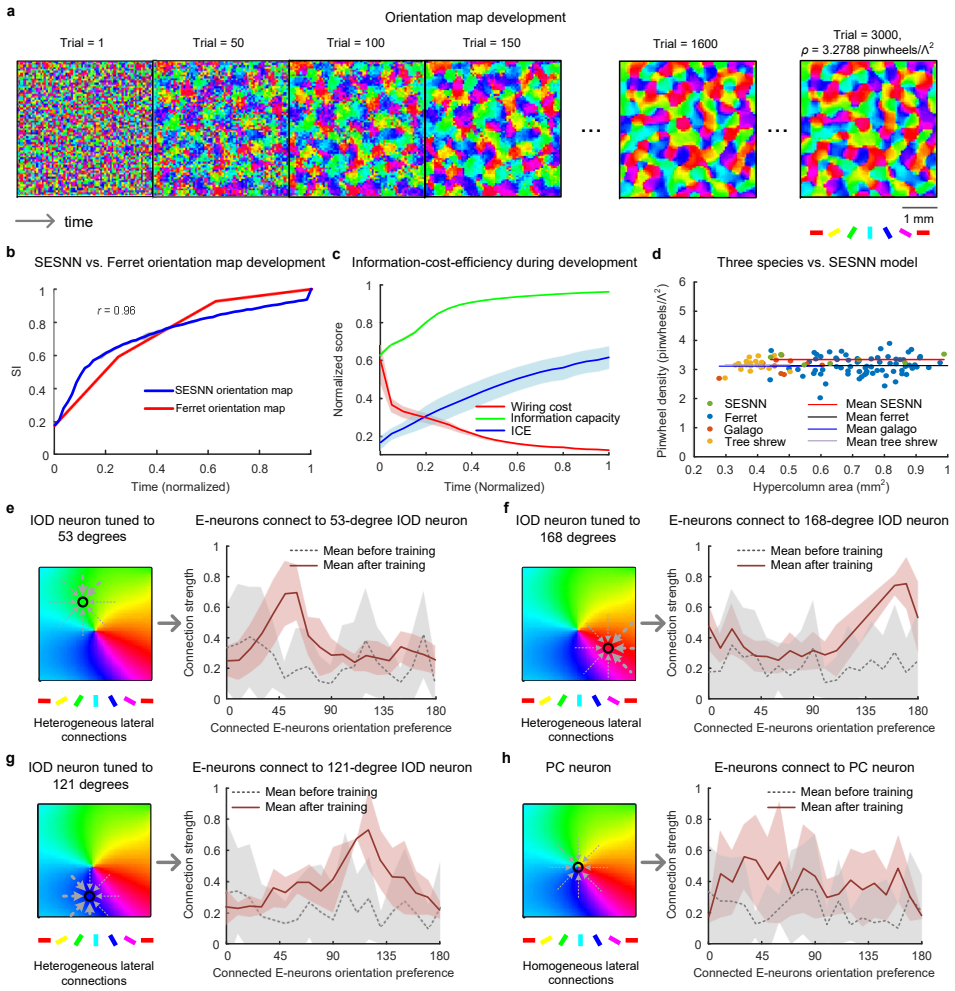

Figure 3: Orientation preference map development in SESNN model. **a**. The temporal evolution of the SESNN model's orientation preference maps, presented in HSV format. The final trial orientation map displays measured density per hypercolumn area (Λ^2). **b**. The correlation between orientation preference maps from the SESNN model and observed orientation preference maps in the ferret over a normalized time scale.**c**. The changes in three different metrics—wiring cost, information capacity, and information-cost-efficiency during orientation map development. **d**. Comparative analysis of pinwheel densities relative to hypercolumn areas for ferrets, galagos, and tree shrews ((Kaschube et al., 2010; Stevens et al., 2013)), alongside the SESNN model. **e-h**. Subplots with insets of pinwheel structures identify focal neurons and depict changes in connection strength among E-neurons with various orientation tunings before and after training (dashed and solid lines: mean). **e-g**. Display heterogeneous connection weights that bias toward iso-orientation domains. **h**. The subplot shows that the homogeneous connection weights contributed equally from nearby iso-orientation domains. The thickness of the arrows represents connection strength. All shaded areas: SD.

where H represents the information encoding capacity, and C represents the energy cost of synaptic transmission, P_k is the discrete probability of the distribution, that is, the normalized count of W_{ij} in k-bins of the same size (0.01). Here, the calculation of wiring costs is based on Eq. 7 from Bertens & Lee (2022). In this approach, the network wiring cost is inversely proportional to the summed synaptic connection weights between pairs of neurons, indicating that stronger synaptic weights not only facilitate efficient communication but also contribute to lower wiring costs. In Fig. 3c, we observed decreasing wiring costs during the development, which is consistent with previous research Koulakov & Chklovskii (2001). Meanwhile, the growing information capacity and the ICE

suggest that the emergence of the pinwheel structure effectively maximizes coding capacity under the constraint of energy cost.

To further understand the circuit mechanisms underlying the emergence of pinwheel structure, we examine the connections between E-neurons for ones at the iso-orientation domains (e.g., Fig. 3e-g) and ones at the pinwheel centers (e.g., Fig. 3h). All connections of E-neurons are initialized with Gaussian-distributed strengths (see Eq. 5) and distributed uniformly across neurons of different orientation preference (determined after stabilized), as indicated by the gray dashed curves. After the orientation preference maps stabilized, the connectivity patterns has significantly evolved leading to a selective strong connection to neurons with similar orientation preference, clearly demonstrated by the red solid curves and the varied thickness of arrows. The temporal in-variance of the connection distribution at the pinwheel center over time (except for increase in overall strength) therefore clearly demonstrated that the iso-orientation domains must take form first, as also visually verified in early trials of our model (Fig. 3a). This local continuity of iso-orientation domains is consistent with the demand of Hebbian-like learning rule (Eq. 1) together the overlap between RFs. The exact size of the domain, as explained earlier, is determined by the extent of the E-connections; while their swirling patterns are instead demanded by the rich local orientations in each stimuli (as described in Section 2.1) such that the animal would be able to perceive different orientations in the same local visual field but still satisfying most of the continuity locally. Note that in the salt-and-pepper organization, the mechanisms are disrupted due to reduced overlap. The local continuity of iso-orientation domains is consistent with the Hebbian-like learning rule (Eq. 1) and the overlap between RFs. The domain size is determined by the extent of E-connections, while swirling patterns arise from the diverse local orientations in stimuli (as described in Section 2.1). This allows animals to perceive varying orientations within the same local visual field while maintaining local continuity. In salt-and-pepper organizations, reduced overlap disrupts these mechanisms. Hence, visual stimulus overlap critically shapes orientation preference maps in V1: greater overlap promotes pinwheel structures, while reduced overlap leads to salt-and-pepper patterns.

3.2 Coding efficiency and robustness emergence within pinwheels

To assess the response reliability of neurons under stochastic conditions—vital for trustworthy and robust neural information processing—we analyze neuron reliability within orientation preference maps under noisy environments ($\mathcal{N}(0, \sigma_r^2)$). We utilize a spike-based measurement approach to quantify this reliability. Specifically, for each neuron, we examine spike time sequences generated under identical stimulus conditions with added noise, calculating the mean cross-correlation between these sequences. We then determine the neuron's reliability by identifying the maximum in the average cross-correlation. For each neuron, we analyze n spike trains of duration T. After normalizing the mean of each train to zero, we systematically calculate the cross-correlation coefficient between each pair of sequences from these n trains:

$$R_{xy}(m) = E\left[x_{T+m} y_T\right], \tag{8}$$

where m denotes lag, the range of which can extend over the length of the time sequence T. x and y denote two different spike trains. Through this procedure, the average correlation coefficient of the same neuron can be determined:

$$\overline{R}_{xy}(m) = \frac{\sum_{i=1}^{n} R_{xy}(m)}{n}. \tag{9}$$

We posit that the maximum lag for similar time sequences generated by the same neuron cannot exceed half the length of the time sequence. Consequently, the reliability is obtained as follows:

$$\text{Reliability} = \max_{-\frac{T}{2} \le m \le \frac{T}{2}} \overline{R}_{xy}(m). \tag{10}$$

To assess the response reliability of the SESNN model, we use a noise-corrupted natural image as the input. This involves superimposing Gaussian white noise onto the image. The model is then stimulated with this noise-modified image, with the same variance noise applied in 10 trials.

Our findings indicate that neurons within orientation maps featuring distinct pinwheel structures demonstrate higher reliability compared to those in salt-and-peppers or mini-columns (Fig. 4b-d). This suggests that pinwheel structures in higher mammals (Fig. 4c-d.) enhance the ability to process signals reliably against noise and contribute to improved visual perception. In Fig. 4e, we analyze

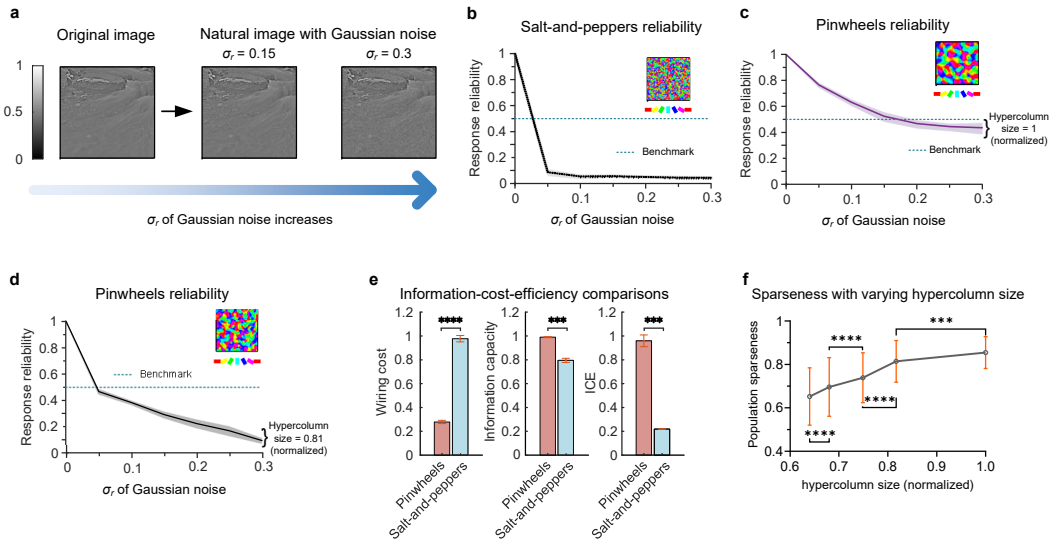

Figure 4: Population response analysis investigates neurons across different orientation preference maps. **a**. The SESNN model is trained using natural image inputs that include background disturbances characterized by Gaussian noise ($\sigma_r \in [0, 0.3]$). **b**. The response reliability of all neurons in the salt-and-peppers gradually decreases as σ_r increases. **c-d**. The response reliability about noise level decreases distinctly for neurons within pinwheel center and surrounding areas across different hypercolumn sizes. Insets illustrate these patterns, with color bars indicating orientation preferences. A dashed blue line at $y = 0.5$ acts as a benchmark for comparing the responses of neuron types under specific noise conditions. **e**. Comparisons of three metrics—wiring cost, information capacity, and ICE—across between pinwheels and salt-and-peppers. **f**. The averaged population response sparseness of neurons increases for pinwheels with larger hypercolumn sizes. (Data presented as mean ± SD, statistical significance denoted as ***p<0.001, ****p<0.0001).

three metrics—wiring cost, information capacity, and Information Cost Efficiency (ICE)—across pinwheels and salt-and-pepper organizations. Pinwheels show lower wiring costs but high information capacity compared to salt-and-peppers. The ICE metric Crumiller et al. (2013), which measures the efficiency of wiring cost relative to information capacity, indicates that pinwheels are more efficient due to their significantly greater information capacity and lower costs. We also calculate the population sparseness Vinje & Gallant (2000) for neurons across different orientation map types when exposed to 100 natural image patches within a 100 ms time window each (Fig. 4d). Population sparseness refers to the distribution of responses among a population of neurons to a single patch, as depicted below:

$$\text{Population sparseness} = \frac{1 - \left(\sum \frac{\langle a_i \rangle}{m}\right)^2 / \sum \left(\frac{\langle a_i \rangle^2}{m}\right)}{1 - \frac{1}{m}}, \tag{11}$$

where a_i refers to the mean spike rate of a neuron i out of a total of m neurons in a time window.

The model's findings reveal a pattern in which population sparseness intensifies with the enlargement of hypercolumn sizes, spanning from small to large across various species. This pattern is illustrated in Fig. 4f, which displays the progressively rising average population sparseness about hypercolumn size across 100 patches. Our model's findings are consistent with empirical data (population sparseness: SESNN pinwheels ~ 0.74 vs. ~ 0.67 macaque pinwheels Zandvakili & Kohn (2015), SESNN salt-and-pepper organizations ~ 0.25 vs. ~ 0.36 mice Froudarakis et al. (2014).

4 Related Works

Traditional models. Traditional models such as SOM (Kohonen, 1982), ON-OFF theoretical models (Miller, 1994; Jang et al., 2020a; Song et al., 2021; Najafian et al., 2022), and artificial neural networks (Margalit et al., 2023; Chizhov & Graham, 2021) have explored orientation map mechanisms but

often lack the complexity found in spiking neural networks. Recent spiking neural networks (Lufkin et al., 2022; Keane & Gong, 2015; Chizhov & Graham, 2021) have struggled with static inputs and connections, failing to accurately capture neural dynamics. Most importantly, no previous spiking neuronal networks with plasticity have actually successfully reproduced pinwheel structures. Line discontinuity instead discrete point discontinuity (as in real pinwheel and our model) plague the functional map (Srinivasa & Jiang, 2013).

Thus, as a major improvement, our model's quantitative match with experiment results greatly enhanced the explaining power of its mechanisms.

Visual overlap among neighboring neurons. Numerous studies have explored the variability in V1 neuronal organization and its impact on afferent segregation, with factors like visual sampling density playing a significant role (Baden et al., 2016; Román Rosón et al., 2019; Srinivasan et al., 2015; Najafian et al., 2022; Jin et al., 2011; Lee et al., 2016; Kremkow et al., 2016; Jang et al., 2020a; Lien & Scanziani, 2013). While these studies have highlighted the importance of anatomical factors such as visual overlap and neuronal density in distinguishing pinwheel structures from salt-and-pepper organizations, they primarily rely on phenomenological static models, lacking detailed mechanisms for how these factors influence functional organization. Our study provides a clear mechanism by demonstrating that visual input overlap is crucial in forming pinwheel structures. Through our spiking neural network model, we show that increased overlap between neighboring neurons enhances local connectivity and functional clustering in iso-orientation domains, leading to the emergence of pinwheels. In contrast, reduced overlap weakens these local connections, resulting in the disorganized salt-and-pepper patterns. This finding establishes a direct link between the structure of input overlap and the functional organization of V1, making the connection between structure and function explicit.

Wiring cost minimization and sparse coding strategy. Research suggests that wiring minimization optimizes orientation maps (Koulakov & Chklovskii, 2001; Petitot, 2017; Dai et al., 2018). Our study expands on this by not only examining the wiring costs but also the information capacity, thereby finding that the ICE of pinwheel structures is much improved compared to salt-and-pepper organizations. Studies also suggest that sparseness in neural coding enhances precision and reduces redundancy (Zhou & Yu, 2018; Yu et al., 2014; Ba & Dj, 1997; Olshausen & Field, 1997; Rolls & Tovee, 1995; Vinje & Gallant, 2000). We found that pinwheel structures strike a balance between reliability and coding sparsity. Lateral connections in pinwheels strengthen synapses among neurons with similar orientations and weaken those with dissimilar orientations. Thus, by enhancing functional clustering in the iso-orientation domains, these neurons boost reliability with some redundancy but still effectively implement sparse coding by suppressing nearby iso-orientation domains. While near pinwheel centers, where connections are less selective, neurons display varied response patterns instead of clear orientation preferences, therefore implements sparse coding of high-order pattern in addition to plain feature space of single orientations.

5 Conclusion

The SESNN model integrates anatomical and physiological factors like visual overlap, neuronal density, and synaptic plasticity, providing a more realistic, dynamic representation of spiking mechanisms in V1, especially in pinwheel formation. Our study identifies visual overlap, neuron density, and local plasticity as key factors driving the transition from salt-and-pepper to pinwheel topologies, linking structural features to functional performance—a significant advancement in understanding visual coding efficiency. By addressing wiring cost minimization and information capacity, our findings reveal optimal cortical network strategies for sparse and robust visual encoding. These insights can inform the design of more efficient brain-inspired computational models.

References

Arish Alreja, Ilya Nemenman, and Christopher J. Rozell. Constrained brain volume in an efficient coding model explains the fraction of excitatory and inhibitory neurons in sensory cortices. *PLOS Computational Biology*, 18(1):e1009642, January 2022. ISSN 1553-7358. doi: 10.1371/journal.pcbi.1009642. URL https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1009642. Publisher: Public Library of Science.

Joseph M. Amatrudo, Christina M. Weaver, Johanna L. Crimins, Patrick R. Hof, Douglas L. Rosene, and Jennifer I. Luebke. Influence of Highly Distinctive Structural Properties on the Excitability of Pyramidal Neurons in Monkey Visual and Prefrontal Cortices. *Journal of Neuroscience*, 32(40): 13644–13660, October 2012. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.2581-12.2012. URL https://www.jneurosci.org/content/32/40/13644. Publisher: Society for Neuroscience Section: Articles.

Olshausen Ba and Field Dj. Sparse coding with an overcomplete basis set: a strategy employed by V1? *Vision research*, 37(23), December 1997. ISSN 0042-6989. doi: 10.1016/s0042-6989(97)00169-7. URL https://pubmed.ncbi.nlm.nih.gov/9425546/. Publisher: Vision Res.

Tom Baden, Philipp Berens, Katrin Franke, Miroslav Román Rosón, Matthias Bethge, and Thomas Euler. The functional diversity of retinal ganglion cells in the mouse. *Nature*, 529(7586):345–350, January 2016. ISSN 1476-4687. doi: 10.1038/nature16468. URL https://www.nature.com/articles/nature16468. Publisher: Nature Publishing Group.

Cornelia I. Bargmann and William T. Newsome. The Brain Research Through Advancing Innovative Neurotechnologies (BRAIN) Initiative and Neurology. *JAMA Neurology*, 71(6): 675–676, June 2014. ISSN 2168-6149. doi: 10.1001/jamaneurol.2014.411. URL https://jamanetwork.com/journals/jamaneurology/fullarticle/1852982. Publisher: American Medical Association.

Paul Bertens and Seong-Whan Lee. Emergence of hierarchical layers in a single sheet of self-organizing spiking neurons. *Advances in Neural Information Processing Systems*, 35:6680–6691, 2022.

Gary G. Blasdel and Guy Salama. Voltage-sensitive dyes reveal a modular organization in monkey striate cortex. *Nature*, 321(6070):579–585, June 1986. ISSN 1476-4687. doi: 10.1038/321579a0. URL https://www.nature.com/articles/321579a0. Number: 6070 Publisher: Nature Publishing Group.

Nicholas M. Blauch, Marlene Behrmann, and David C. Plaut. A connectivity-constrained computational account of topographic organization in primate high-level visual cortex. *Proceedings of the National Academy of Sciences*, 119(3):e2112566119, January 2022. doi: 10.1073/pnas.2112566119. URL https://www.pnas.org/doi/10.1073/pnas.2112566119. Publisher: Proceedings of the National Academy of Sciences.

Tobias Bonhoeffer and Amiram Grinvald. Iso-orientation domains in cat visual cortex are arranged in pinwheel-like patterns. *Nature*, 353(6343):429–431, October 1991. ISSN 1476-4687. doi: 10.1038/353429a0. URL https://www.nature.com/articles/353429a0. Number: 6343 Publisher: Nature Publishing Group.

Richard T. Born, Alexander R. Trott, and Till S. Hartmann. Cortical magnification plus cortical plasticity equals vision? *Vision Research*, 111:161–169, June 2015. ISSN 0042-6989. doi: 10.1016/j.visres.2014.10.002. URL https://www.sciencedirect.com/science/article/pii/S0042698914002351.

William H. Bosking, Ying Zhang, Brett Schofield, and David Fitzpatrick. Orientation Selectivity and the Arrangement of Horizontal Connections in Tree Shrew Striate Cortex. *Journal of Neuroscience*, 17(6):2112–2127, March 1997. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.17-06-02112.1997. URL https://www.jneurosci.org/content/17/6/2112. Publisher: Society for Neuroscience Section: Articles.

B. Chapman and M. P. Stryker. Development of orientation selectivity in ferret visual cortex and effects of deprivation. *Journal of Neuroscience*, 13(12):5251–5262, December 1993. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.13-12-05251.1993. URL https://www.jneurosci.org/content/13/12/5251. Publisher: Society for Neuroscience Section: Articles.

Barbara Chapman, Michael P. Stryker, and Tobias Bonhoeffer. Development of Orientation Preference Maps in Ferret Primary Visual Cortex. *Journal of Neuroscience*, 16(20):6443–6453, October 1996. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.16-20-06443.1996. URL https:

//www.jneurosci.org/content/16/20/6443. Publisher: Society for Neuroscience Section: Articles.

Anton V. Chizhov and Lyle J. Graham. A strategy for mapping biophysical to abstract neuronal network models applied to primary visual cortex. *PLOS Computational Biology*, 17(8):e1009007, August 2021. ISSN 1553-7358. doi: 10.1371/journal.pcbi.1009007. URL https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1009007. Publisher: Public Library of Science.

Marshall Crumiller, Bruce Knight, and Ehud Kaplan. The Measurement of Information Transmitted by a Neural Population: Promises and Challenges. *Entropy*, 15(9):3507–3527, September 2013. ISSN 1099-4300. doi: 10.3390/e15093507. URL https://www.mdpi.com/1099-4300/15/9/3507. Number: 9 Publisher: Multidisciplinary Digital Publishing Institute.

Wei P. Dai, Douglas Zhou, David W. McLaughlin, and David Cai. Mechanisms underlying contrast-dependent orientation selectivity in mouse V1. *Proceedings of the National Academy of Sciences*, 115(45):11619–11624, November 2018. doi: 10.1073/pnas.1719044115. URL https://www.pnas.org/doi/abs/10.1073/pnas.1719044115. Publisher: Proceedings of the National Academy of Sciences.

Yang Dan, Joseph J. Atick, and R. Clay Reid. Efficient Coding of Natural Scenes in the Lateral Geniculate Nucleus: Experimental Test of a Computational Theory. *Journal of Neuroscience*, 16(10):3351–3362, May 1996. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.16-10-03351.1996. URL https://www.jneurosci.org/content/16/10/3351. Publisher: Society for Neuroscience Section: Articles.

Ralf Engelmann and Leo Peichl. Unique Distribution of Somatostatin-immunoreactive Cells in the Retina of the Tree Shrew (Tupaia belangeri). *European Journal of Neuroscience*, 8 (1):220–228, 1996. ISSN 1460-9568. doi: 10.1111/j.1460-9568.1996.tb01183.x. URL https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1460-9568.1996.tb01183.x. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1460-9568.1996.tb01183.x.

Andrzej T. Foik, Leo R. Scholl, Georgina A. Lean, and David C. Lyon. Visual Response Characteristics in Lateral and Medial Subdivisions of the Rat Pulvinar. *Neuroscience*, 441:117–130, August 2020. ISSN 0306-4522. doi: 10.1016/j.neuroscience.2020.06.030. URL https://www.sciencedirect.com/science/article/pii/S0306452220304073.

Emmanouil Froudarakis, Philipp Berens, Alexander S. Ecker, R. James Cotton, Fabian H. Sinz, Dimitri Yatsenko, Peter Saggau, Matthias Bethge, and Andreas S. Tolias. Population code in mouse V1 facilitates read-out of natural scenes through increased sparseness. *Nature neuroscience*, 17(6):851–857, June 2014. ISSN 1097-6256. doi: 10.1038/nn.3707. URL https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4106281/.

P. Földiák. Forming sparse representations by local anti-Hebbian learning. *Biological Cybernetics*, 64(2):165–170, December 1990. ISSN 1432-0770. doi: 10.1007/BF02331346. URL https://link.springer.com/article/10.1007/BF02331346. Company: Springer Distributor: Springer Institution: Springer Label: Springer Number: 2 Publisher: Springer-Verlag.

Sergej V. Girman, Yves Sauvé, and Raymond D. Lund. Receptive Field Properties of Single Neurons in Rat Primary Visual Cortex. *Journal of Neurophysiology*, 82(1):301–311, July 1999. ISSN 0022-3077. doi: 10.1152/jn.1999.82.1.301. URL https://journals.physiology.org/doi/full/10.1152/jn.1999.82.1.301. Publisher: American Physiological Society.

W C Hall, J H Kaas, H Killackey, and I T Diamond. Cortical visual areas in the grey squirrel (Sciurus carolinesis): a correlation between cortical evoked potential maps and architectonic subdivisions. *Journal of Neurophysiology*, 34(3):437–452, May 1971. ISSN 0022-3077. doi: 10.1152/jn.1971.34.3.437. URL https://journals.physiology.org/doi/abs/10.1152/jn.1971.34.3.437. Publisher: American Physiological Society.

Sonja B. Hofer, Ho Ko, Bruno Pichler, Joshua Vogelstein, Hana Ros, Hongkui Zeng, Ed Lein, Nicholas A. Lesica, and Thomas D. Mrsic-Flogel. Differential connectivity and response dynamics of excitatory and inhibitory neurons in visual cortex. *Nature Neuroscience*, 14(8):1045–1052, August 2011. ISSN 1546-1726. doi: 10.1038/nn.2876. URL https://www.nature.com/articles/nn.2876. Number: 8 Publisher: Nature Publishing Group.

Carl Holmgren, Tibor Harkany, Björn Svennenfors, and Yuri Zilberter. Pyramidal cell communication within local networks in layer 2/3 of rat neocortex. *The Journal of Physiology*, 551(Pt 1):139–153, August 2003. ISSN 0022-3751. doi: 10.1113/jphysiol.2003.044784. URL https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2343144/.

D. H. Hubel and T. N. Wiesel. Receptive fields, binocular interaction and functional architecture in the cat's visual cortex. *The Journal of Physiology*, 160(1):106–154, 1962. ISSN 1469-7793. doi: 10.1113/jphysiol.1962.sp006837. URL https://onlinelibrary.wiley.com/doi/abs/10.1113/jphysiol.1962.sp006837. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1113/jphysiol.1962.sp006837.

David H. Hubel and Torsten N. Wiesel. Sequence regularity and geometry of orientation columns in the monkey striate cortex. *Journal of Comparative Neurology*, 158(3):267–293, 1974. ISSN 1096-9861. doi: 10.1002/cne.901580304. URL https://onlinelibrary.wiley.com/doi/abs/10.1002/cne.901580304. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1002/cne.901580304.

Andrew D. Huberman, Colenso M. Speer, and Barbara Chapman. Spontaneous Retinal Activity Mediates Development of Ocular Dominance Columns and Binocular Receptive Fields in V1. *Neuron*, 52 (2):247–254, October 2006. ISSN 0896-6273. doi: 10.1016/j.neuron.2006.07.028. URL https://www.cell.com/neuron/abstract/S0896-6273(06)00625-8. Publisher: Elsevier.

A. Hughes. A schematic eye for the rat. *Vision Research*, 19(5):569–588, January 1979. ISSN 0042-6989. doi: 10.1016/0042-6989(79)90143-3. URL https://www.sciencedirect.com/science/article/pii/0042698979901433.

Jaeson Jang, Min Song, and Se-Bum Paik. Retino-Cortical Mapping Ratio Predicts Columnar and Salt-and-Pepper Organization in Mammalian Visual Cortex. *Cell Reports*, 30(10):3270–3279.e3, March 2020a. ISSN 2211-1247. doi: 10.1016/j.celrep.2020.02.038. URL https://www.cell.com/cell-reports/abstract/S2211-1247(20)30199-6. Publisher: Elsevier.

Jaeson Jang, Min Song, and Se-Bum Paik. Retino-Cortical Mapping Ratio Predicts Columnar and Salt-and-Pepper Organization in Mammalian Visual Cortex. *Cell Reports*, 30(10):3270–3279.e3, March 2020b. ISSN 2211-1247. doi: 10.1016/j.celrep.2020.02.038. URL https://www.cell.com/cell-reports/abstract/S2211-1247(20)30199-6. Publisher: Elsevier.

Jianzhong Jin, Yushi Wang, Harvey A. Swadlow, and Jose M. Alonso. Population receptive fields of ON and OFF thalamic inputs to an orientation column in visual cortex. *Nature Neuroscience*, 14(2):232–238, February 2011. ISSN 1546-1726. doi: 10.1038/nn.2729. URL https://www.nature.com/articles/nn.2729. Publisher: Nature Publishing Group.

Matthias Kaschube, Michael Schnabel, Siegrid Löwel, David M. Coppola, Leonard E. White, and Fred Wolf. Universality in the Evolution of Orientation Columns in the Visual Cortex. *Science*, 330(6007):1113–1116, November 2010. doi: 10.1126/science.1194869. URL https://www.science.org/doi/10.1126/science.1194869. Publisher: American Association for the Advancement of Science.

Adam Keane and Pulin Gong. Propagating Waves Can Explain Irregular Neural Dynamics. *Journal of Neuroscience*, 35(4):1591–1605, January 2015. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.1669-14.2015. URL https://www.jneurosci.org/content/35/4/1591. Publisher: Society for Neuroscience Section: Articles.

Jason Keller, Hans Strasburger, Daniel T Cerutti, and Bernhard A Sabel. Assessing spatial vision — automated measurement of the contrast-sensitivity function in the hooded rat. *Journal of Neuroscience Methods*, 97(2):103–110, April 2000. ISSN 0165-0270. doi: 10.

1016/S0165-0270(00)00173-4. URL https://www.sciencedirect.com/science/article/pii/S0165027000001734.

Teuvo Kohonen. Self-organized formation of topologically correct feature maps. *Biological Cybernetics*, 43(1):59–69, January 1982. ISSN 1432-0770. doi: 10.1007/BF00337288. URL https://doi.org/10.1007/BF00337288.

Alexei A. Koulakov and Dmitri B. Chklovskii. Orientation Preference Patterns in Mammalian Visual Cortex: A Wire Length Minimization Approach. *Neuron*, 29(2):519–527, February 2001. ISSN 0896-6273. doi: 10.1016/S0896-6273(01)00223-9. URL https://www.sciencedirect.com/science/article/pii/S0896627301002239.

Jens Kremkow, Jianzhong Jin, Yushi Wang, and Jose M. Alonso. Principles underlying sensory map topography in primary visual cortex. *Nature*, 533(7601):52–57, May 2016. ISSN 1476-4687. doi: 10.1038/nature17936. URL https://www.nature.com/articles/nature17936. Publisher: Nature Publishing Group.

Margaret I. Law, Kathleen R. Zahs, and Michael P. Stryker. Organization of primary visual cortex (area 17) in the ferret. *Journal of Comparative Neurology*, 278(2):157–180, 1988. ISSN 1096-9861. doi: 10.1002/cne.902780202. URL https://onlinelibrary.wiley.com/doi/abs/10.1002/cne.902780202. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1002/cne.902780202.

Kuo-Sheng Lee, Xiaoying Huang, and David Fitzpatrick. Topology of ON and OFF inputs in visual cortex enables an invariant columnar architecture. *Nature*, 533(7601):90–94, May 2016. ISSN 1476-4687. doi: 10.1038/nature17941. URL https://www.nature.com/articles/nature17941. Publisher: Nature Publishing Group.

Anthony D. Lien and Massimo Scanziani. Tuned thalamic excitation is amplified by visual cortical circuits. *Nature Neuroscience*, 16(9):1315–1323, September 2013. ISSN 1546-1726. doi: 10.1038/nn.3488. URL https://www.nature.com/articles/nn.3488. Publisher: Nature Publishing Group.

Leon Lufkin, Ashish Puri, Ganlin Song, Xinyi Zhong, and John Lafferty. *Emergent organization of receptive fields in networks of excitatory and inhibitory neurons*. May 2022. doi: 10.48550/arXiv.2205.13614.

Eshed Margalit, Hyodong Lee, Dawn Finzi, James J. DiCarlo, Kalanit Grill-Spector, and Daniel L. K. Yamins. A Unifying Principle for the Functional Organization of Visual Cortex, May 2023. URL https://www.biorxiv.org/content/10.1101/2023.05.18.541361v1. Pages: 2023.05.18.541361 Section: New Results.

Yicong Meng, Shigeru Tanaka, and Chi-Sang Poon. Comment on "Universality in the Evolution of Orientation Columns in the Visual Cortex". *Science*, 336(6080):413–413, April 2012. doi: 10.1126/science.1205737. URL https://www.science.org/doi/10.1126/science.1205737. Publisher: American Association for the Advancement of Science.

K. D. Miller. A model for the development of simple cell receptive fields and the ordered arrangement of orientation columns through activity-dependent competition between ON- and OFF-center inputs. *Journal of Neuroscience*, 14(1):409–441, January 1994. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.14-01-00409.1994. URL https://www.jneurosci.org/content/14/1/409. Publisher: Society for Neuroscience Section: Articles.

T. Müller, M. Stetter, M. Hübener, F. Sengpiel, T. Bonhoeffer, I. Gödecke, B. Chapman, S. Löwel, and K. Obermayer. An analysis of orientation and ocular dominance patterns in the visual cortex of cats and ferrets. *Neural Computation*, 12(11):2573–2595, November 2000. ISSN 0899-7667. doi: 10.1162/089976600300014854.

Sohrab Najafian, Erin Koch, Kai Lun Teh, Jianzhong Jin, Hamed Rahimi-Nasrabadi, Qasim Zaidi, Jens Kremkow, and Jose-Manuel Alonso. A theory of cortical map formation in the visual brain. *Nature Communications*, 13(1):2303, April 2022. ISSN 2041-1723. doi: 10.1038/s41467-022-29433-y. URL https://www.nature.com/articles/s41467-022-29433-y. Number: 1 Publisher: Nature Publishing Group.

Cristopher M. Niell and Michael P. Stryker. Highly Selective Receptive Fields in Mouse Visual Cortex. *Journal of Neuroscience*, 28(30):7520–7536, July 2008. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.0623-08.2008. URL https://www.jneurosci.org/content/28/30/7520. Publisher: Society for Neuroscience Section: Articles.

Erkki Oja. Simplified neuron model as a principal component analyzer. *Journal of Mathematical Biology*, 15(3):267–273, November 1982. ISSN 1432-1416. doi: 10.1007/BF00275687. URL https://doi.org/10.1007/BF00275687.

B. A. Olshausen and D. J. Field. Sparse coding with an overcomplete basis set: a strategy employed by V1? *Vision Research*, 37(23):3311–3325, December 1997. ISSN 0042-6989. doi: 10.1016/s0042-6989(97)00169-7.

Paul D. King, Joel Zylberberg, and Michael R. DeWeese. Inhibitory Interneurons Decorrelate Excitatory Cells to Drive Sparse Code Formation in a Spiking Model of V1. *The Journal of Neuroscience*, 33(13):5475, March 2013. doi: 10.1523/JNEUROSCI.4188-12.2013. URL http://www.jneurosci.org/content/33/13/5475.abstract.

Jean Petitot. *Elements of Neurogeometry: Functional Architectures of Vision*. Lecture Notes in Morphogenesis. Springer International Publishing, Cham, 2017. ISBN 978-3-319-65589-5 978-3-319-65591-8. doi: 10.1007/978-3-319-65591-8. URL http://link.springer.com/10.1007/978-3-319-65591-8.

Carsten K. Pfeffer, Mingshan Xue, Miao He, Z. Josh Huang, and Massimo Scanziani. Inhibition of inhibition in visual cortex: the logic of connections between molecularly distinct interneurons. *Nature Neuroscience*, 16(8):1068–1076, August 2013. ISSN 1546-1726. doi: 10.1038/nn.3446. URL https://www.nature.com/articles/nn.3446. Number: 8 Publisher: Nature Publishing Group.

Ryan T. Philips, Mriganka Sur, and V. Srinivasa Chakravarthy. The influence of astrocytes on the width of orientation hypercolumns in visual cortex: A computational perspective. *PLOS Computational Biology*, 13(10):e1005785, October 2017. ISSN 1553-7358. doi: 10.1371/journal.pcbi.1005785. URL https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1005785. Publisher: Public Library of Science.

Dario L. Ringach, Patrick J. Mineault, Elaine Tring, Nicholas D. Olivas, Pablo Garcia-Junco-Clemente, and Joshua T. Trachtenberg. Spatial clustering of tuning in mouse primary visual cortex. *Nature Communications*, 7(1):12270, August 2016. ISSN 2041-1723. doi: 10.1038/ncomms12270. URL https://www.nature.com/articles/ncomms12270. Number: 1 Publisher: Nature Publishing Group.

E. T. Rolls and M. J. Tovee. Sparseness of the neuronal representation of stimuli in the primate temporal visual cortex. *Journal of Neurophysiology*, 73(2):713–726, February 1995. ISSN 0022-3077. doi: 10.1152/jn.1995.73.2.713. URL https://journals.physiology.org/doi/abs/10.1152/jn.1995.73.2.713. Publisher: American Physiological Society.

Miroslav Román Rosón, Yannik Bauer, Ann H. Kotkat, Philipp Berens, Thomas Euler, and Laura Busse. Mouse dLGN Receives Functional Input from a Diverse Population of Retinal Ganglion Cells with Limited Convergence. *Neuron*, 102(2):462–476.e8, April 2019. ISSN 0896-6273. doi: 10.1016/j.neuron.2019.01.040. URL https://www.sciencedirect.com/science/article/pii/S0896627319300674.

Tatsuo K. Sato, Bilal Haider, Michael Häusser, and Matteo Carandini. An excitatory basis for divisive normalization in visual cortex. *Nature Neuroscience*, 19(4):568–570, April 2016. ISSN 1546-1726. doi: 10.1038/nn.4249. URL https://www.nature.com/articles/nn.4249. Number: 4 Publisher: Nature Publishing Group.

Benjamin Scholl, Johannes Burge, and Nicholas J. Priebe. Binocular integration and disparity selectivity in mouse primary visual cortex. *Journal of Neurophysiology*, 109(12):3013–3024, June 2013. ISSN 0022-3077. doi: 10.1152/jn.01021.2012. URL https://journals.physiology.org/doi/full/10.1152/jn.01021.2012. Publisher: American Physiological Society.

Min Song, Jaeson Jang, Gwangsu Kim, and Se-Bum Paik. Projection of Orthogonal Tiling from the Retina to the Visual Cortex. *Cell Reports*, 34(1), January 2021. ISSN 2211-1247. doi: 10.1016/j.celrep.2020.108581. URL https://www.cell.com/cell-reports/abstract/S2211-1247(20)31570-9. Publisher: Elsevier.

Narayan Srinivasa and Qin Jiang. Stable learning of functional maps in self-organizing spiking neural networks with continuous synaptic plasticity. *Frontiers in Computational Neuroscience*, 7, 2013. ISSN 1662-5188. URL https://www.frontiersin.org/articles/10.3389/fncom.2013.00010.

Shyam Srinivasan, C. Nikoosh Carlo, and Charles F. Stevens. Predicting visual acuity from the structure of visual cortex. *Proceedings of the National Academy of Sciences*, 112(25):7815–7820, June 2015. doi: 10.1073/pnas.1509282112. URL https://www.pnas.org/doi/10.1073/pnas.1509282112. Publisher: Proceedings of the National Academy of Sciences.

Armen Stepanyants, Luis M. Martinez, Alex S. Ferecskó, and Zoltán F. Kisvárday. The fractions of short- and long-range connections in the visual cortex. *Proceedings of the National Academy of Sciences*, 106(9):3555–3560, March 2009. doi: 10.1073/pnas.0810390106. URL https://www.pnas.org/doi/abs/10.1073/pnas.0810390106. Publisher: Proceedings of the National Academy of Sciences.

Jean-Luc R. Stevens, Judith S. Law, Ján Antolík, and James A. Bednar. Mechanisms for Stable, Robust, and Adaptive Development of Orientation Maps in the Primary Visual Cortex. *Journal of Neuroscience*, 33(40):15747–15766, October 2013. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.1037-13.2013. URL https://www.jneurosci.org/content/33/40/15747. Publisher: Society for Neuroscience Section: Articles.

Louis Tao, Michael Shelley, David McLaughlin, and Robert Shapley. An egalitarian network model for the emergence of simple and complex cells in visual cortex. *Proceedings of the National Academy of Sciences*, 101(1):366–371, January 2004. doi: 10.1073/pnas.2036460100. URL https://www.pnas.org/doi/full/10.1073/pnas.2036460100. Publisher: Proceedings of the National Academy of Sciences.

Edward J. Tehovnik and Kyoungmin Lee. The dorsomedial frontal cortex of the rhesus monkey: topographic representation of saccades evoked by electrical stimulation. *Experimental Brain Research*, 96(3):430–442, November 1993. ISSN 1432-1106. doi: 10.1007/BF00234111. URL https://doi.org/10.1007/BF00234111.

Edward J. Tehovnik and Warren M. Slocum. Phosphene induction by microstimulation of macaque V1. *Brain Research Reviews*, 53(2):337–343, February 2007. ISSN 0165-0173. doi: 10.1016/j.brainresrev.2006.11.001. URL https://www.sciencedirect.com/science/article/pii/S0165017306001160.

Alex Thomson and Christophe Lamy. Functional maps of neocortical local circuitry. *Frontiers in Neuroscience*, 1, 2007. ISSN 1662-453X. URL https://www.frontiersin.org/articles/10.3389/neuro.01.1.1.002.2007.

Enny H. van Beest, Sreedeep Mukherjee, Lisa Kirchberger, Ulf H. Schnabel, Chris van der Togt, Rob R. M. Teeuwen, Areg Barsegyan, Arne F. Meyer, Jasper Poort, Pieter R. Roelfsema, and Matthew W. Self. Mouse visual cortex contains a region of enhanced spatial resolution. *Nature Communications*, 12(1):4029, June 2021. ISSN 2041-1723. doi: 10.1038/s41467-021-24311-5. URL https://www.nature.com/articles/s41467-021-24311-5. Number: 1 Publisher: Nature Publishing Group.

David C. Van Essen, William T. Newsome, and John H. R. Maunsell. The visual field representation in striate cortex of the macaque monkey: Asymmetries, anisotropies, and individual variability. *Vision Research*, 24(5):429–448, January 1984. ISSN 0042-6989. doi: 10.1016/0042-6989(84)90041-5. URL https://www.sciencedirect.com/science/article/pii/0042698984900415.

Julia Veit, Anwesha Bhattacharyya, Robert Kretz, and Gregor Rainer. On the Relation Between Receptive Field Structure and Stimulus Selectivity in the Tree Shrew Primary Visual Cortex.

Cerebral Cortex, 24(10):2761–2771, October 2014. ISSN 1047-3211. doi: 10.1093/cercor/bht133. URL https://doi.org/10.1093/cercor/bht133.

William E. Vinje and Jack L. Gallant. Sparse Coding and Decorrelation in Primary Visual Cortex During Natural Vision. *Science*, 287(5456):1273–1276, February 2000. doi: 10.1126/science.287.5456.1273. URL https://www.science.org/doi/10.1126/science.287.5456.1273. Publisher: American Association for the Advancement of Science.

Marvin Weigand, Fabio Sartori, and Hermann Cuntz. Universal transition from unstructured to structured neural maps. *Proceedings of the National Academy of Sciences*, 114(20):E4057–E4064, May 2017. doi: 10.1073/pnas.1616163114. URL https://www.pnas.org/doi/full/10.1073/pnas.1616163114. Publisher: Proceedings of the National Academy of Sciences.

Yuguo Yu, Michele Migliore, Michael L. Hines, and Gordon M. Shepherd. Sparse Coding and Lateral Inhibition Arising from Balanced and Unbalanced Dendrodendritic Excitation and Inhibition. *Journal of Neuroscience*, 34(41):13701–13713, October 2014. ISSN 0270-6474, 1529-2401. doi: 10.1523/JNEUROSCI.1834-14.2014. URL https://www.jneurosci.org/content/34/41/13701. Publisher: Society for Neuroscience Section: Articles.

Amin Zandvakili and Adam Kohn. Coordinated Neuronal Activity Enhances Corticocortical Communication. *Neuron*, 87(4):827–839, August 2015. ISSN 0896-6273. doi: 10.1016/j.neuron.2015.07.026. URL https://www.sciencedirect.com/science/article/pii/S0896627315006480.

Qing-fang Zhang, Hao Li, Ming Chen, Aike Guo, Yunqing Wen, and Mu-ming Poo. Functional organization of intrinsic and feedback presynaptic inputs in the primary visual cortex. *Proceedings of the National Academy of Sciences*, 115(22):E5174–E5182, May 2018. doi: 10.1073/pnas.1719711115. URL https://www.pnas.org/doi/full/10.1073/pnas.1719711115. Publisher: Proceedings of the National Academy of Sciences.

Shanglin Zhou and Yuguo Yu. Synaptic E-I Balance Underlies Efficient Neural Coding. *Frontiers in Neuroscience*, 12, February 2018. ISSN 1662-453X. doi: 10.3389/fnins.2018.00046. URL https://www.frontiersin.org/journals/neuroscience/articles/10.3389/fnins.2018.00046/full. Publisher: Frontiers.

Joel Zylberberg, Jason Timothy Murphy, and Michael Robert DeWeese. A Sparse Coding Model with Synaptically Local Plasticity and Spiking Neurons Can Account for the Diverse Shapes of V1 Simple Cell Receptive Fields. *PLOS Computational Biology*, 7(10):1–12, October 2011. doi: 10.1371/journal.pcbi.1002250. URL https://doi.org/10.1371/journal.pcbi.1002250. Publisher: Public Library of Science.

A Appendix

A.1 Detailed neural model

The spiking neuronal dynamics are iteratively formulated as follows:

$$\begin{aligned} u_i^{(\mathrm{K})}(t+1) = u_i^{(\mathrm{K})}(t) e^{-\frac{\eta}{\tau^{(\mathrm{K})}}} + h_{\mathrm{K}}(i) \sum_j FF_{ij}^{(\mathrm{E}\leftarrow\text{image})} X_j \\ + \sum_{\mathrm{K}^*} \sum_j \beta_{ij}^{(\mathrm{K}\leftarrow\mathrm{K}^*)} \cdot W_{ij}^{(\mathrm{K}\leftarrow\mathrm{K}^*)} \cdot z_j^{(\mathrm{K}^*)}(t) + \text{noise}, \end{aligned} \tag{12}$$

$$h_{\mathrm{K}}(i) = \begin{cases} 1, & \text{if } i \text{ is an E-neuron ID}, \\ 0, & \text{if } i \text{ is an I-neuron ID}, \end{cases} \tag{13}$$

$$\Delta\theta_i^{(\mathrm{K}^*)} \propto p_i(z_i^{(\mathrm{K}^*)} = 1) - p_i^{(\mathrm{K}^*)}, \tag{14}$$

where K = E-neurons or I-neurons; $i = 1, 2, \ldots, N$ (numbers of E-neurons and I-neurons).

In Eq. 12, $u_i^{(\mathrm{K})}(t)$ represents the membrane potential of neuron i at time t, applicable to neurons of class K. The decay rate of the membrane potential is governed by the resistor-capacitor circuit time

constant τ, set at $\tau^{(\mathrm{E})} = 10$ ms for E-neurons and $\tau^{(\mathrm{I})} = 5$ ms for I-neurons, reflecting the faster firing rate of I-neurons compared to E-neurons ((Paul D. King et al., 2013; Thomson & Lamy, 2007)). This configuration reduces reconstruction error and speeds up system convergence, enhancing the representation of input stimuli. The feedforward (FF) connection, $\mathrm{FF}_{ij}^{(\mathrm{E}\leftarrow\mathrm{image})}$, connects pixel X_j from a whitened image patch (normalized to zero mean and unit variance) to E-neuron *i*. Connection weight $W_{ij}^{(\mathrm{K}\leftarrow\mathrm{K}^*)}$ from neuron *j* of class K^* to neuron *i* of class K includes a sign $\beta_{ij}^{(\mathrm{K}\leftarrow\mathrm{K}^*)}$ indicating excitatory (+1) or inhibitory (-1) connections. $z_j^{(\mathrm{K}^*)}(t) = H\left(u_j^{(\mathrm{K}^*)}(t) - \theta_j^{(\mathrm{K}^*)}\right)$, where $H(x)$ is the Heaviside step function, defined as $H(x) = 1$ if $x > 0$ and 0 otherwise. Here, $z_j^{(\mathrm{K}^*)}(t)$ represents the spike output from neuron j at time t, which triggers a reset of the membrane potential to 0 after reaching the spike threshold θ (initially set at 2). The firing threshold θ adapts based on the discrepancy between the current and target firing rates $p_i(t)$ and $p_i^{(\mathrm{K})}$ ($p^{(\mathrm{E})} = 0.02,\ p^{(\mathrm{I})} = 0.04$) within a 10 ms time window (see Eq. 14 (Földiák, 1990)). For computational efficiency, the time step η is set at 1 ms.

A.2 Anatomical Data Integration

Neural Connection Data

The experimental subjects include six adult cats with unknown genders, with data sourced from research by Armen Stepanyants et al.(Stepanyants et al., 2009); and eight macaques, aged 5-11 years, including six males and two females, with data sourced from research by Joseph Amatrudo et al.(Amatrudo et al., 2012).

Neuronal Synaptic Plasticity

The subjects are rats aged 14-16 days, with unknown gender and quantity, with data sourced from research by Holmgren et al.(Holmgren et al., 2003); transgenic mice, with unknown quantity and gender, with data sourced from research by Hofer et al. (Hofer et al., 2011).

Retinal-V1 Topological Projection Data

RF data: V1 neuron counts for macaques, cats, tree shrews, ferrets, mice, rats, and gray squirrels respectively come from Tehovnik et al. (Tehovnik & Lee, 1993) (subjects: 3 macaques, unknown gender and age), Scholl et al. (Scholl et al., 2013) (subjects: cats, unknown gender and age), Veit et al.(Veit et al., 2014) (subjects: 9 male and 7 female tree shrews, aged 3-8 years), Huberman et al.(Huberman et al., 2006) (subjects: 8 ferrets, unknown gender and age), Niell et al.(Niell & Stryker, 2008) (subjects: mice, aged 2–6 months, unknown gender), Foik et al.(Foik et al., 2020)(subjects: 21 rats, unknown gender and age), and Hall et al.(Hall et al., 1971) (subjects: 17 gray squirrels, unknown gender and age). V1 neuronal density: neuronal density data for macaques, cats, mice, rats, and gray squirrels come from Srinivasana et al.(Tehovnik & Slocum, 2007) (subjects: unknown gender and age); tree shrew, ferret, and gray squirrel density data respectively come from Weigand et al.(Weigand et al., 2017).

Cortical Magnification Factor

Cortical magnification factor data for macaques, cats, tree shrews, ferrets, mice, rats, and gray squirrels are sourced from Tehovnik et al.(Tehovnik & Lee, 1993) (subjects: 3 macaques, unknown gender and age), Veit et al.(Veit et al., 2014)(subjects: cats, unknown gender and age), Bosking et al.(Bosking et al., 1997)(subjects: tree shrews, unknown gender and age), Rockland et al. (Law et al., 1988)(subjects: 9 ferrets, female, unknown age), Beest et al.(van Beest et al., 2021)(subjects: 28 mice, 11 males and 17 females, ages 2-14 months), Keller et al.(Keller et al., 2000) (subjects: male rats, age 3 months), and Hall et al.(Hall et al., 1971) (subjects: 17 gray squirrels, unknown gender and age).

Table 3: Comparative anatomical data of the retina and V1 across species.

a. Species (mean)	**b.** Retina (mm^2)	**c.** V1 size (mm^2)	**d.** V1 neurons density (neurons/mm^2)	**e.** V1 RF size (deg)	**f.** RFs density ($(c) \times (d)/(b)$) (RFs/mm^2)
Macaque	636[a]	1,090[b]	243,000[b]	0.2[c]	416,462.26
Cat	510[a]	380[b]	99,200[b]	1.0[d]	73,913.73
Tree shrew	122[a,e]	73[b,a]	192,800[f]	2.0[g]	115,363.93
Ferret	83[h,a]	78[b]	95,813[f]	3.0[i]	90,041.13
Mouse	15[a]	2.5[b]	86,600[b]	4.0[j]	14,433.33
Rat	52[a,k]	7.1[b]	90,800[b]	3.0[l]	12,397.69
Gray squirrel	205[a]	32[a]	84,213[f]	2.0[m]	13,145.44

[a] Jang et al. (2020b) [b] Srinivasan et al. (2015) [c] Tehovnik & Slocum (2007)
[d] Scholl et al. (2013) [e] Engelmann & Peichl (1996) [f] Weigand et al. (2017)
[g] Veit et al. (2014) [h] Law et al. (1988) [i] Huberman et al. (2006)
[j] Niell & Stryker (2008) [k] Hughes (1979) [l] Foik et al. (2020)
[m] Hall et al. (1971)

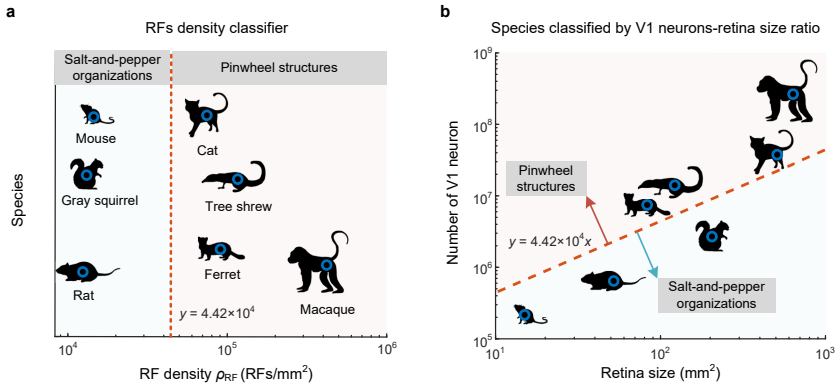

Figure 5: A linear classifier based on RFD ($y = 4.42 \times 10^4 x$) effectively differentiates species with salt-and-pepper organizations (rats, mice, gray squirrels) from those with pinwheel structures (macaques, ferrets, cats, tree shrews). textbfa. This classifier reflects variations in V1 organizations across species. **b**. A plot categorizing species by the ratio of V1 neuron number to retina size acts as a divider, implying a critical ratio for the formation of pinwheel structures.

A.3 Unveiling species-specific factors distinguishing pinwheels and salt-and-peppers

A.3.1 Anatomical data suggests RFs density underlying V1 organizations

We analyzed anatomical data from seven species, including primates (e.g., macaques) and non-primates (e.g., mice, rats, cats, tree shrews, gray squirrels, and ferrets), as detailed in Table 3. We first find that V1 RFD (ρ_{RF}) acts as a linear classifier ($y = 4.42 \times 10^4 x$), effectively distinguishing species with pinwheel structures from those with salt-and-pepper organizations. In this classifier, species like macaques, cats, tree shrews, and ferrets, which have higher RFD, are associated with pinwheel structures (light red area in Fig 5) and exceed the classification threshold. In contrast, species with lower RFD, such as mice, rats, and gray squirrels, are linked to salt-and-pepper organizations (light blue area in Fig 5). Thus, V1 RFD serves as a predictive metric for V1 organizational patterns across species. The ρ_{RF} is calculated as follows:

$$\rho_{\text{RF}} = \frac{n}{s_{\text{r}}} = \frac{n}{\left[\left(s_{\text{RF}} - \varepsilon\right)\left(\sqrt{n} - 1\right) + s_{\text{RF}}\right]^2}, \tag{15}$$

where n denotes the total number of neurons in V1, s_{r} indicates the retinal surface area. The variable ε quantifies the degree of visual input overlap among adjacent neurons, and s_{RF} represents the RF

size. Concerning the Eq. 15 and anatomical data (Table 3), the two main factors influencing RFD ρ_{RF} are the overlap ε of visual inputs between adjacent RFs and V1 neuronal density ρ_{V1}. We discuss the overlap in the main text. Neuronal density is discussed in the following sections.

A.3.2 SESNN REVEALS NEURONAL DENSITY INFLUENCING V1 CLUSTERS

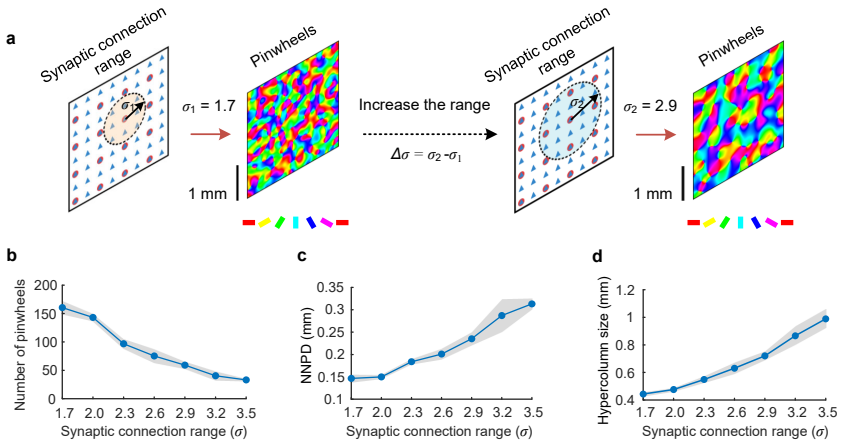

Figure 6: Synaptic connection range within V1 contributes to the formation of pinwheel structures. **a**. Modifying the synaptic connection range reshapes the dimensions of pinwheel structures. **b-d**. The relationship between the synaptic connection range (σ) and the number of pinwheels, NNPD (mm), and hypercolumn size (mm). The scale bar: 1 mm in V1 cortical surface. Color scheme: orientation preference. Lines: mean. Shaded area: SD.

The anatomical data in Table 3d for seven species show variability in V1 neuronal density (ρ_{V1}), which influences inter-neuronal spacing and connection strength. We explore how V1 cortical orientation patterns form by adjusting the lateral connection range, impacting axon reach among E- and I-neurons, as depicted in Fig. 6. We modulate axonal arborization through parameter σ to adjust the connection range, allowing us to simulate neuronal connections in areas with varying densities. This setup enables the SESNN model to predict changes in cortical patterns (Fig. 6). Our observations indicate that increasing axon lengths, thereby extending the connection range, enlarges hypercolumn sizes within pinwheel structures (Fig. 6d), reduces the overall number of pinwheels (Fig. 6b), and increases NNPD (Fig. 6c). These findings underscore the critical role of neural synaptic connection range in organizing orientation maps.

A.4 QUANTITATIVE DATA ON CORTICAL PINWHEEL STRUCTURES

The visual input overlap is quantified as follows:

$$\varepsilon_{\text{percentage}} = \frac{\sqrt[2]{\rho_{\text{V1}} S_{\text{unit}}} - \frac{L_{\text{unit}} M}{RF_{\text{size}}}}{\sqrt[2]{\rho_{\text{V1}} S_{\text{unit}}} - 1} \times 100\% \tag{16}$$

where S_{unit} represents the unit cortical area, RF_{size} denotes the size of the RF in V1, represents the unit area, and ρ_{V1} represents the density of neurons in V1. We consider only an effective cortical layer composed of output neurons. This is because the apparent overlap within a vertical cortical column primarily contributes to intermediary processing stages for the same input. Therefore, such overlaps should not be conflated with overlaps in the input space. Consequently, the 1/3 root of the symbol ρ_{V1} is calculated as the product of two factors: 2/3, representing the overlap between columnar structures, and 1/2, indicating one side of a 2D cortical rectangular area. M refers to the CMF. The trend shown in Fig. 2e indicates increasing overlap for mice, macaques, and cats, which is consistent with the overlap settings of our model. This escalation in overlap corresponds to the expansion of iso-orientation domains and pinwheels, presented by our model's predictions and anatomically detailed by Najafian et al.Najafian et al. (2022).

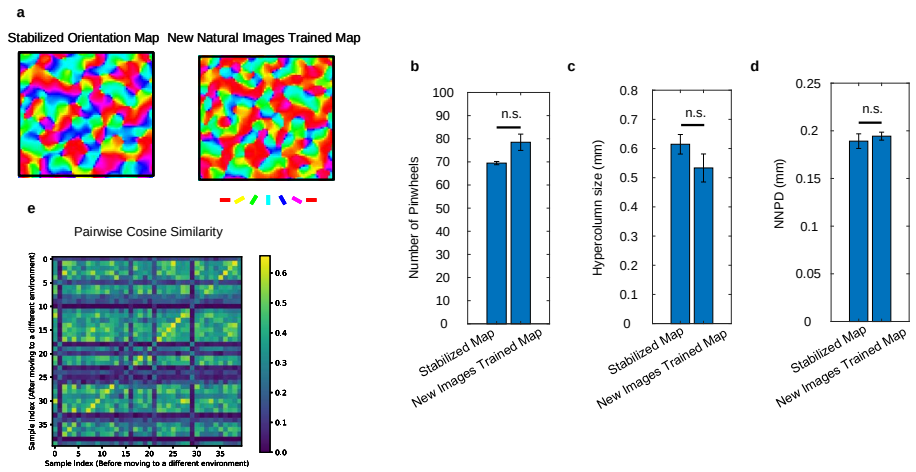

Figure 7: Plasticity of the pinwheel structure in response to new natural images. (a) Stabilized orientation map and orientation map after training with new natural images. (b) Number of pinwheels. (c) Hypercolumn size. (d) Nearest-neighbor pinwheel distance. (e) Pairwise cosine similarity between neuronal representations before and after introducing new environments.

A.5 Stability and Adaptability of Pinwheel Structures

We conducted additional experiments to examine the plasticity of the pinwheel structure in response to new visual environments. A new set of natural images, including forests, large stones, tree branches and leaves, reeds, stones of various sizes, tree trunks, leaves, and grasses, was introduced after the pinwheel structure had stabilized. After further training with these new images, we observed slight modifications in some finer aspects of the pinwheel structure; however, the overall structure, including the locations of the pinwheel centers, remained largely stable (Fig. 7a).

To quantitatively assess these changes, we employed three different metrics: (i) the number of pinwheels, (ii) hypercolumn size, and (iii) nearest-neighbor pinwheel distance. These metrics were used to compare the pinwheel structure before and after introducing the new images (Fig. 7b, c, d). Additionally, we utilized pairwise cosine similarity to measure the representation similarity of the neuronal spike train population (Fig. 7e).

Although we did not observe significant changes in the morphology of the pinwheels, some detailed aspects of the pinwheel structure did change. Specifically, while the locations of the pinwheel centers and the overall domains remained nearly unchanged, some neurons exhibited tuning adjustments towards specific orientations. Furthermore, the representation of neurons to the same stimulus remained overall stable, with minor variations before and after the introduction of new environments, suggesting that the pinwheel structure is highly adaptable (Fig. 7).

These results indicate that the model's organizational structure is deeply rooted in the statistical features of natural images and demonstrates a strong capacity for adaptation to new environments while preserving its core structural properties.

A.6 Orientation Map Formation from Our Proposed Spiking Network Model (New Section)

1. HO Rule Learns V1 RF: The HO rule learns RFs from visual images, as described by the following formula:

$$\Delta FF_{im}^{(\mathrm{E}\leftarrow\mathrm{Image})} \propto y_i x_m - y_i^2 FF_{im}^{(\mathrm{E}\leftarrow\mathrm{Image})}, \tag{17}$$

where x_m and y_i represent the gray value of pixel m and instantaneous firing rate of post-synaptic neuron i. This rule ensures that weights converge when:

$$FF_{im} = \frac{\langle y_i x_m \rangle}{\langle y_i^2 \rangle} = \frac{\text{STA}_i}{\langle y_i^2 \rangle}. \tag{18}$$

Corresponding to the normalized spike-triggered average (STA). HO rule stabilizes learning by constraining synaptic growth, resulting in biologically realistic V1-like RFs.

2. Retinotopic Topographic Constraints: Each neuron encodes only a small portion of the visual field (Fig. 1a), with the small patch $\in \mathbb{R}^{H' \times W'}$ smaller than the input image $I \in \mathbb{R}^{H \times W}$. The population's collective response is needed to represent the full stimulus, aligning with the topographic mapping from retina to cortex.

3. High-Overlap Strengthens Local Excitatory-Excitatory Connections for Orientation Map Continuity: Excitatory connections align adjacent neurons' firing activities, promoting similar orientation preferences. This requires a high overlap (Fig. 2), where correlated feedforward input initially strengthens local excitatory connections, ensuring continuous orientation preference and forming iso-orientation domains with similar orientation preferences (Fig. 3e-h).

4. CM rule's Anti-Correlation via E-I-E loops Ensures Orientation Diversity and Map Coverage: Inhibitory neurons, through the CM rule:

$$\Delta W_{ij}^{(\mathrm{K} \leftarrow \mathrm{K}^*)} \propto y_i x_j - \langle y_i \rangle \langle x_j \rangle \left(1 + W_{ij}^{(\mathrm{K} \leftarrow \mathrm{K}^*)}\right), \tag{19}$$

where x_j and y_i represent the instantaneous firing rates of pre- and post-synaptic neuron j and i. The operator $\langle \cdot \rangle$ denotes the moving average value (refer to Eq. 4).

At equilibrium, the CM rule converges to:

$$W_{ij} = \frac{\langle y_i x_j \rangle - \langle y_i \rangle \langle x_j \rangle}{\langle y_i \rangle \langle x_j \rangle}. \tag{20}$$

We have:

$$W_{ij} \propto \langle y_i x_j \rangle - \langle y_i \rangle \langle x_j \rangle = \text{Cov}(y_i, x_j). \tag{21}$$

Covariance shapes neural connectivity based on activity covariance, where synaptic weights are strengthened between correlated neurons and weakened between uncorrelated ones. This mechanism ensures that inhibitory neurons regulate overall activity between neighbouring E neurons through the E-I-E loops (lateral inhibition), suppressing predictable responses to maintain sparse coding. This also prevents excitatory neurons from excitatory connections overly responding to the same orientation, enabling preference for other orientations and ensuring full coverage. Over larger connection ranges, highly correlated feedforward correlations are disrupted and inhibitory neurons drive diverse responses, resulting in distinct domains with varying preferred orientations, forming pinwheel structures within the functional orientation map.

A.7 Computing infrastructure

CPU	Intel® Xeon(R) Gold 6348 CPU @ 2.60GHz
GPU	A100
Memory	512 GB
Operating system	Ubuntu 20.04.6 LTS
Simulation platform	MATLAB R2023a and Python 3.9

Table 4: Computing infrastructure

A.8 Acronym

Acronym	Term
V1	Primary Visual Cortex
SESNN	Self-Evolving Spiking Neural Network
HO	Hebbian Oja's variant
CM	Correlation Measuring
ICE	Information-Cost Efficiency
CMF	Cortical Magnification Factor
E-I	Excitatory-Inhibitory
FF	Feedforward
RF	Receptive Field

Table 5: List of acronyms and their corresponding terms