PROTEIN REPRESENTATION LEARNING BY GEOMETRIC STRUCTURE PRETRAINING

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ABSTRACT

Learning effective representations of proteins is critical in a variety of tasks in biology such as predicting protein function or structure. Existing approaches usually pretrain protein language models on a large number of unlabeled amino acid sequences and then finetune the models with some labeled data in downstream tasks. Despite the effectiveness of sequence-based approaches, the power of pretraining on known protein structures, which are available in smaller numbers only, has not been explored for protein property prediction, though protein structures are known to be determinants of protein function. In this paper, we propose to pretrain protein representations according to their 3D structures. We first present a simple yet effective encoder to learn the geometric features of a protein. We pretrain the protein structure encoder by leveraging multiview contrastive learning and compare against pretraining with various self-prediction tasks. Experimental results on both function prediction and fold classification tasks show that our proposed pretraining method outperforms or is on par with the state-of-the-art sequence-based methods, while using much less data. All codes and models will be published upon acceptance.

1 INTRODUCTION

Proteins are workhorses of the cell and are implicated in a broad range of applications ranging from therapeutics to material. They consist of a linear chain of amino acids (residues) which fold into specific conformations. Due to the advent of low cost sequencing technologies (Ma & Johnson, 2012; Ma, 2015), in recent years a massive volume of protein sequences have been newly discovered. As functional annotation of a new protein sequence remains costly and time-consuming, accurate and efficient in silico protein function annotation methods are needed to bridge the existing sequence-function gap.

Since a large number of protein functions are governed by their folded structures, several data-driven approaches rely on learning representations of the protein structures, which then can be used for a variety of tasks such as protein design (Ingraham et al., 2019; Strokach et al., 2020; Cao et al., 2021; Jing et al., 2021), structure classification (Hermosilla et al., 2021), model quality assessment (Baldassarre et al., 2021; Derevyanko et al., 2018), and function prediction (Gligorijević et al., 2021). Due to the challenge of experimental protein structure determination, the number of reported protein structures is orders of magnitude lower than the size of datasets in other machine learning application domains. For example, there are 182K experimentally-determined structures in the Protein Data Bank (PDB) (Berman et al., 2000) *vs* 47M protein sequences in Pfam (Mistry et al., 2021) and *vs* 10M annotated images in ImageNet (Russakovsky et al., 2015).

To address this gap, recent works have leveraged the large volume of unlabeled protein sequence data to learn an effective representation of known proteins (Bepler & Berger, 2019; Rives et al., 2021; Elnaggar et al., 2021). A number of studies have pretrained protein encoders on millions of sequences via self-supervised learning. However, these methods neither explicitly capture nor leverage the available protein structural information that is known to be the determinants of protein functions.

To better utilize structural information, several structure-based protein encoders (Hermosilla et al., 2021; Hermosilla & Ropinski, 2022; Wang et al., 2022a) have been proposed. However, these models have not explicitly captured the interactions between edges, which are critical in protein structure modeling (Jumper et al., 2021). Besides, very few attempts (Hermosilla & Ropinski, 2022; Chen et al.,

2022; Guo et al., 2022) have been made until recently to develop pretraining methods that exploit unlabeled 3D structures due to the scarcity of experimentally-determined protein structures. Thanks to recent advances in highly accurate deep learning-based protein structure prediction methods (Baek et al., 2021; Jumper et al., 2021), it is now possible to efficiently predict structures for a large number of protein sequences with reasonable confidence.

Motivated by this development, we develop a universal protein encoder pretrained on the largest possible number[1] of protein structures that is able to generalize to a variety of property prediction tasks. We propose a simple yet effective structure-based encoder called **GeomEtry-Aware Relational Graph Neural Network (GearNet)**, which encodes spatial information by adding different types of sequential or structural edges and then performs relational message passing on protein residue graphs. Inspired by the design of triangle attention in Evoformer (Jumper et al., 2021), we propose a *sparse edge message passing* mechanism to enhance the protein structure encoder, which is the first attempt to incorporate edge-level message passing on GNNs for protein structure encoding.

We further introduce a geometric pretraining method to learn the protein structure encoder based on the popular contrastive learning framework (Chen et al., 2020). We propose novel augmentation functions to discover biologically correlated protein substructures that co-occur in proteins (Ponting & Russell, 2002) and aim to maximize the similarity between the learned representations of substructures from the same protein, while minimizing the similarity between those from different proteins. Simultaeneously, we propose a suite of straightforward baselines based on self-prediction (Devlin et al., 2018). These pretraining tasks perform masked prediction of different geometric or physicochemical attributes, such as residue types, Euclidean distances, angles and dihedral angles. Through extensively benchmarking these pretraining techniques on diverse downstream property prediction tasks, we set up a solid starting point for pretraining protein structure representations.

Extensive experiments on several benchmarks, including Enzyme Commission number prediction (Gligorijević et al., 2021), Gene Ontology term prediction (Gligorijević et al., 2021), fold classification (Hou et al., 2018) and reaction classification (Hermosilla et al., 2021) verify our GearNet augmented with edge message passing can consistently outperform existing protein encoders on most tasks in a supervised setting. Further, by employing the proposed pretraining method, our model trained on fewer than a million samples achieves comparable or even better results than the state-of-the-art sequence-based encoders pretrained on million- or billion-scale datasets.

2 Related Work

Previous works seek to learn protein representations based on different modalities of proteins, including amino acid sequences (Rao et al., 2019; Elnaggar et al., 2021; Rives et al., 2021), multiple sequence alignments (MSAs) (Rao et al., 2021; Biswas et al., 2021; Meier et al., 2021) and protein structures (Hermosilla et al., 2021; Gligorijević et al., 2021; Somnath et al., 2021). These works share the common goal of learning informative protein representations that can benefit various downstream applications, like predicting protein function (Rives et al., 2021) and protein-protein interaction (Wang et al., 2019), as well as designing protein sequences (Biswas et al., 2021).

Compared with sequence-based methods, structure-based methods should be, in principle, a better solution to learning an informative protein representation, as the function of a protein is determined by its structure. This line of works seeks to encode spatial information in protein structures by 3D CNNs (Derevyanko et al., 2018) or graph neural networks (GNNs) (Gligorijević et al., 2021; Baldassarre et al., 2021; Jing et al., 2021; Wang et al., 2022a; Aykent & Xia, 2022). Among these methods, IEConv (Hermosilla et al., 2021) tries to fit the inductive bias of protein structure modeling, which introduced a graph convolution layer incorporating intrinsic and extrinsic distances between nodes. Another potential direction is to extract features from protein surfaces (Gainza et al., 2020; Sverrisson et al., 2021; Dai & Bailey-Kellogg, 2021). Somnath et al. (2021) combined the advantages of both worlds and proposed a parameter-efficient multi-scale model. Besides, there are also works that enhance pretrained sequence-based models by incorporating structural information in the pretraining stage (Bepler & Berger, 2021) or finetuning stage (Wang et al., 2022b).

[1]We use AlphaFoldDB v1 and v2 (Varadi et al., 2021) for pretraining, which is the largest protein structure database before March, 2022.

Despite progress in the design of structure-based encoders, there are few works focusing on structure-based pretraining for proteins. To the best of our knowledge, the only attempt is three concurrent works (Hermosilla & Ropinski, 2022; Chen et al., 2022; Guo et al., 2022), which apply contrastive learning, self-prediction and denoising score matching methods on a small set of tasks, respectively. Compared with these existing works, our proposed encoder is conceptually simpler and more effective on many different tasks, thanks to the proposed relational graph convolutional layer and edge message passing layer, which are able to efficiently capture both the sequential and structural information. Furthermore, we introduce a contrastive learning framework with novel augmentation functions to discover substructures in different proteins and four different self-prediction tasks, which can serve as a solid starting point for enabling self-supervised learning on protein structures.

3 STRUCTURE-BASED PROTEIN ENCODER

Existing protein encoders are either designed for specific tasks or cumbersome for pretraining due to the dependency on computationally expensive convolutions. In contrast, here we propose a simple yet effective protein structure encoder, named *GeomEtry-Aware Relational Graph Neural Network (GearNet)*. We utilize *sparse edge message passing* to enhance the effectiveness of GearNet, which is novel and crucial in the field of protein structure modeling, whereas previous works (Hermosilla et al., 2021; Somnath et al., 2021) only consider message passing among residues or atoms.

3.1 GEOMETRY-AWARE RELATIONAL GRAPH NEURAL NETWORK

Given protein structures, our model aims to learn representations encoding their spatial and chemical information. These representations should be invariant under translations, rotations and reflections in 3D space. To achieve this requirement, we first construct our protein graph based on spatial features invariant under these transformations.

Protein graph construction. We represent the structure of a protein as a residue-level relational graph $\mathcal{G} = (\mathcal{V}, \mathcal{E}, \mathcal{R})$, where $\mathcal{V}$ and $\mathcal{E}$ denotes the set of nodes and edges respectively, and $\mathcal{R}$ is the set of edge types. We use (i, j, r) to denote the edge from node i to node j with type r. We use n and m to denote the number of nodes and edges, respectively. In this work, each node in the protein graph represents the alpha carbon of a residue with the 3D coordinates of all nodes $\boldsymbol{x} \in \mathbb{R}^{n\times 3}$. We use $\boldsymbol{f}_i$ and $\boldsymbol{f}_{(i,j,r)}$ to denote the feature for node i and edge (i, j, r), respectively.

Then, we add three different types of directed edges into our graphs: sequential edges, radius edges and K-nearest neighbor edges. Among these, sequential edges will be further divided into 5 types of edges based on the relative sequential distance $d \in \{-2, -1, 0, 1, 2\}$ between two end nodes, where we add sequential edges only between the nodes within the sequential distance of 2. These edge types reflect different geometric properties, which all together yield a comprehensive featurization of proteins. More details of the graph and feature construction process can be found in Appendix C.1.

Relational graph convolutional layer. Upon the protein graphs defined above, we utilize a GNN to derive per-residue and whole-protein representations. One simple example of GNNs is the GCN (Kipf & Welling, 2017), where messages are computed by multiplying node features with a convolutional kernel matrix shared among all edges. To increase the capacity in protein structure modeling, IEConv (Hermosilla et al., 2021) proposed to apply a learnable kernel function on edge features. In this way, m different kernel matrices can be applied on different edges, which achieves good performance but induces high memory costs.

To balance model capacity and memory cost, we use a relational graph convolutional neural network (Schlichtkrull et al., 2018) to learn graph representations, where a convolutional kernel matrix is shared within each edge type and there are $|\mathcal{R}|$ different kernel matrices in total. Formally, the relational graph convolutional layer used in our model is defined as

$$\boldsymbol{h}_i^{(0)} = \boldsymbol{f}_i, \quad \boldsymbol{u}_i^{(l)} = \sigma\left(\mathrm{BN}\left(\sum\nolimits_{r\in\mathcal{R}} \boldsymbol{W}_r \sum\nolimits_{j\in\mathcal{N}_r(i)} \boldsymbol{h}_j^{(l-1)}\right)\right), \quad \boldsymbol{h}_i^{(l)} = \boldsymbol{h}_i^{(l-1)} + \boldsymbol{u}_i^{(l)}. \tag{1}$$

Specifically, we use node features $\boldsymbol{f}_i$ as initial representations. Then, given the node representation $\boldsymbol{h}_i^{(l)}$ for node i at the l-th layer, we compute updated node representation $\boldsymbol{u}_i^{(l)}$ by aggregating features

from neighboring nodes $\mathcal{N}_r(i)$, where $\mathcal{N}_r(i) = \{j \in \mathcal{V} | (j, i, r) \in \mathcal{E}\}$ denotes the neighborhood of node i with the edge type r, and $\boldsymbol{W}_r$ denotes the learnable convolutional kernel matrix for edge type r. Here BN denotes a batch normalization layer and we use a ReLU function as the activation $\sigma(\cdot)$. Finally, we update $\boldsymbol{h}_i^{(l)}$ with $\boldsymbol{u}_i^{(l)}$ and add a residual connection from last layer.

3.2 Edge Message Passing Layer

As in the literature of molecular representation learning, many geometric encoders show benefits from explicitly modeling interactions between edges. For example, DimeNet (Klicpera et al., 2020) uses a 2D spherical Fourier-Bessel basis function to represent angles between two edges and pass messages between edges. AlphaFold2 (Jumper et al., 2021) leverages the triangle attention designed for transformers to model pair representations. Inspired by this observation, we propose a variant of GearNet enhanced with an edge message passing layer, named as **GearNet-Edge**. The edge message passing layer can be seen as a sparse version of the pair representation update designed for graph neural networks. The main objective is to model the dependency between different interactions of a residue with other sequentially or spatially adjacent residues.

Formally, we first construct a relational graph $\mathcal{G}' = (\mathcal{V}', \mathcal{E}', \mathcal{R}')$ among edges, which is also known as *line graph* in the literature (Harary & Norman, 1960). Each node in the graph $\mathcal{G}'$ corresponds to an edge in the original graph. $\mathcal{G}'$ links edge (i, j, r_1) in the original graph to edge (w, k, r_2) if and only if $j = w$ and $i \neq k$. The type of this edge is determined by the angle between (i, j, r_1) and (w, k, r_2). We discretize the range $[0, \pi]$ into 8 bins and use the index of the bin as the edge type.

Then, we apply a similar relational graph convolutional network on the graph $\mathcal{G}'$ to obtain the message function for each edge. Formally, the edge message passing layer is defined as

$$\boldsymbol{m}_{(i,j,r_1)}^{(0)} = \boldsymbol{f}_{(i,j,r_1)}, \quad \boldsymbol{m}_{(i,j,r_1)}^{(l)} = \sigma\left(\mathrm{BN}\left(\sum\nolimits_{r \in \mathcal{R}'} \boldsymbol{W}_r' \sum\nolimits_{(w,k,r_2) \in \mathcal{N}_r'((i,j,r_1))} \boldsymbol{m}_{(w,k,r_2)}^{(l-1)}\right)\right).$$

Here we use $\boldsymbol{m}_{(i,j,r_1)}^{(l)}$ to denote the message function for edge (i, j, r_1) in the l-th layer. Similar as Eq. (1), the message function for edge (i, j, r_1) will be updated by aggregating features from its neighbors $\mathcal{N}_r'((i, j, r_1))$, where $\mathcal{N}_r'((i, j, r_1)) = \{(w, k, r_2) \in \mathcal{V}' | ((w, k, r_2), (i, j, r_1), r) \in \mathcal{E}'\}$ denotes the set of incoming edges of (i, j, r_1) with relation type r in graph $\mathcal{G}'$.

Finally, we replace the aggregation function Eq. (1) in the original graph with the following one:

$$\boldsymbol{u}_i^{(l)} = \sigma\left(\mathrm{BN}\left(\sum\nolimits_{r \in \mathcal{R}} \boldsymbol{W}_r \sum\nolimits_{j \in \mathcal{N}_r(i)} (\boldsymbol{h}_j^{(l-1)} + \mathrm{FC}(\boldsymbol{m}_{(j,i,r)}^{(l)})\right)\right), \tag{2}$$

where $\mathrm{FC}(\cdot)$ denotes a linear transformation upon the message function.

Notably, it is a novel idea to use relational message passing to model different spatial interactions among residues. In addition, to the best of our knowledge, this is the first work that explores edge message passing for macromolecular representation learning.

Invariance of GearNet and GearNet-Edge. The graph construction process and input features of GearNet and GearNet-Edge only rely on features (distances and angles) invariant to translation, rotation and reflection. Besides, the message passing layers use pre-defined edge types and other 3D information invariantly. Therefore, GearNet and GearNet-Edge can achieve E(3)-invariance.

4 Geometric Pretraining Methods

In this section, we study how to boost protein representation learning via self-supervised pretraining on a massive collection of unlabeled protein structures. Despite the efficacy of self-supervised pretraining in many domains, applying it to protein representation learning is nontrivial due to the difficulty of capturing both biochemical and spatial information in protein structures. To address the challenge, we first introduce our multiview contrastive learning method with novel augmentation functions to discover correlated co-occurrence of protein substructures and align their representations in the latent space. Furthermore, four self-prediction baselines are also proposed for pretraining structure-based encoders.

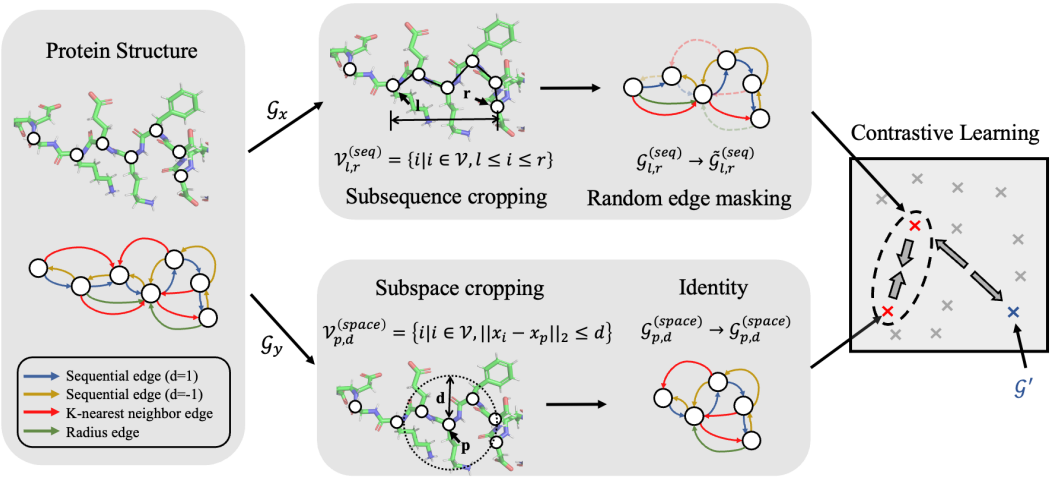

Figure 1: Demonstration of multiview contrastive learning. For each protein, we first construct the residue graph $\mathcal{G}$ based on the structural information (some edges are omitted to save space). Next, two views $\mathcal{G}_x$ and $\mathcal{G}_y$ of the protein are generated by randomly choosing the sampling scheme and noise function. For $\mathcal{G}_x$, we first extract a subsequence and then perform random edge masking with dash lines indicating masked edges. For $\mathcal{G}_y$, we perform subspace cropping and then keep the subspace graph $\mathcal{G}_{p,d}^{(\text{space})}$ unchanged. Finally, a contrastive learning loss is optimized to maximize the similarity between $\mathcal{G}_x$ and $\mathcal{G}_y$ in the latent space while minimizing its similarity with a negative sample $\mathcal{G}'$.

4.1 Multiview Contrastive Learning

It is known that structural motifs within folded protein structures are biologically related (Mackenzie & Grigoryan, 2017) and protein substructures can reflect the evolution history and functions of proteins (Ponting & Russell, 2002). Inspired by recent contrastive learning methods (Chen et al., 2020; He et al., 2020), our framework aims to preserve the similarity between these correlated substructures before and after mapping to a low-dimensional latent space. Specifically, using a similarity measurement defined in the latent space, biologically-related substructures are embedded close to each other while unrelated ones are mapped far apart. Figure 1 illustrates the high-level idea.

Constructing views that reflect protein substructures. Given a protein graph $\mathcal{G}$, we consider two different sampling schemes for constructing views. The first one is **subsequence cropping**, which randomly samples a left residue l and a right residue r and takes all residues ranging from l to r. This sampling scheme aims to capture protein domains, consecutive protein subsequences that reoccur in different proteins and indicate their functions (Ponting & Russell, 2002). However, simply sampling protein subsequences cannot fully utilize the 3D structural information in protein data. Therefore, we further introduce a **subspace cropping** scheme to discover spatially correlated structural motifs. We randomly sample a residue p as the center and select all residues within a Euclidean ball with a predefined radius d. For the two sampling schemes, we take the corresponding subgraphs from the protein residue graph $\mathcal{G} = (\mathcal{V}, \mathcal{E}, \mathcal{R})$. In specific, the subsequence graph $\mathcal{G}_{l,r}^{(\text{seq})}$ and the subspace graph $\mathcal{G}_{p,d}^{(\text{space})}$ can be written as:

$$\mathcal{V}_{l,r}^{(\text{seq})} = \{i | i \in \mathcal{V}, l \le i \le r\}, \quad \mathcal{E}_{l,r}^{(\text{seq})} = \{(i,j,r) | (i,j,r) \in \mathcal{E}, i \in \mathcal{V}_{l,r}^{(\text{seq})}, j \in \mathcal{V}_{l,r}^{(\text{seq})}\},$$
$$\mathcal{V}_{p,d}^{(\text{space})} = \{i | i \in \mathcal{V}, \|\boldsymbol{x}_i - \boldsymbol{x}_p\|_2 \le d\}, \quad \mathcal{E}_{p,d}^{(\text{space})} = \{(i,j,r) | (i,j,r) \in \mathcal{E}, i \in \mathcal{V}_{p,d}^{(\text{space})}, j \in \mathcal{V}_{p,d}^{(\text{space})}\},$$

where $\mathcal{G}_{l,r}^{(\text{seq})} = (\mathcal{V}_{l,r}^{(\text{seq})}, \mathcal{E}_{l,r}^{(\text{seq})}, \mathcal{R})$ and $\mathcal{G}_{p,d}^{(\text{space})} = (\mathcal{V}_{p,d}^{(\text{space})}, \mathcal{E}_{p,d}^{(\text{space})}, \mathcal{R})$.

After sampling the substructures, following the common practice in self-supervised learning (Chen et al., 2020), we apply a noise function to generate more diverse views and thus benefit the learned representations. Here we consider two noise functions: **identity** that applies no transformation and **random edge masking** that randomly masks each edge with a fixed probability 0.15.

Contastive learning. We follow SimCLR (Chen et al., 2020) to optimize a contrastive loss function and thus maximize the mutual information between these biologically correlated views. For each protein $\mathcal{G}$, we sample two views $\mathcal{G}_x$ and $\mathcal{G}_y$ by first randomly choosing one sampling scheme for extracting substructures and then randomly selecting one of the two noise functions with equal probability. We compute the graph representations $\boldsymbol{h}_x$ and $\boldsymbol{h}_y$ of two views using our structure-based encoder. Then, a two-layer MLP projection head is applied to map the representations to a lower-dimensional space, denoted as $\boldsymbol{z}_x$ and $\boldsymbol{z}_y$. Finally, an InfoNCE loss function is defined by distinguishing views from the same or different proteins using their similarities (Oord et al., 2018). For a positive pair x and y, we treat views from other proteins in the same mini-batch as negative pairs. Mathematically, the loss function for a positive pair of views x and y can be written as:

$$\mathcal{L}_{x,y} = -\log \frac{\exp(\text{sim}(\boldsymbol{z}_x, \boldsymbol{z}_y)/\tau)}{\sum_{k=1}^{2B} \mathbb{1}_{[k \neq x]} \exp(\text{sim}(\boldsymbol{z}_y, \boldsymbol{z}_k)/\tau)}, \tag{3}$$

where B denotes the batch size, τ denotes the temperature, $\mathbb{1}_{[k \neq x]} \in \{0, 1\}$ is an indicator function that is equal to 1 if and only if $k \neq x$. And the similarity function $\text{sim}(\boldsymbol{u}, \boldsymbol{v})$ is defined by the cosine similarity between $\boldsymbol{u}$ and $\boldsymbol{v}$.

Sizes of sampled substructures. The design of our random sampling scheme aims to extract biologically meaningful substructures for contrastive learning. To attain this goal, it is of critical importance to determine the sampling length $r - l$ for subsequence cropping and radius d for subspace cropping. On the one hand, we need sufficiently long subsequences and large subspaces to ensure that the sampled substructures can meaningfully reflect the whole protein structure and thus share high mutual information with each other. On the other hand, if the sampled substructures are too large, the generated views will be so similar that the contrastive learning problem becomes trivial. Furthermore, large substructures limit the batch size used in contrastive learning, which has been shown to be harmful for the performance (Chen et al., 2020).

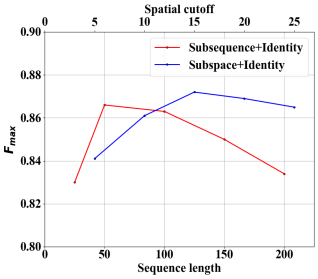

Figure 2: F_{max} on EC *v.s.* cropping sequence length (# residues) for Subsequence Cropping and spatial cutoff (Å) for Subspace Cropping.

To study the effect of sizes of sampled substructures, we show experimental results on EC using Multiview Contrast with the subsequence and substructure cropping function, respectively. To prevent the influence of noise functions, we only consider identity transformation in both settings. The results are plotted in Figure 2. For both subsequence and subspace cropping, as the sampled substructures become larger, the results first increase and then decrease from a certain threshold. This phenomenon agrees with our analysis above. In practice, we set 50 residues as the sampling length for subsequecne cropping and 15Å as the radius for subspace cropping, which are large enough to capture most structural motifs according to statistics reported in previous studies (Tateno et al., 1997).

In Appendix G, we visualize the representations of the pretrained model with this method and assign familial labels based on domain annotations. We can observe clear separation based on the familial classification of the proteins in the dataset, which proves the effectiveness of our pretraining method.

4.2 Straightforward Baselines: Self-Prediction Methods

Another line of model pre-training research is based on the recent progress of self-prediction methods in natural language processing (Devlin et al., 2018; Brown et al., 2020). Along that line, given a protein, our objective can be formulated as predicting one part of the protein given the remainder of the structure. Here, we propose four self-supervised tasks based on physicochemical or geometric properties: residue types, distances, angles and dihedrals.

The four methods perform masked prediction on single residues, residue pairs, triplets and quadruples, respectively. Masked residue type prediction is widely used for pretraining protein language models (Bepler & Berger, 2021) and also known as masked inverse folding in the protein community (Yang et al., 2022). Distances, angles and dihedrals have been shown to be important features that reflect the relative position between residues (Klicpera et al., 2020). For angle and dihedral prediction, we sample adjacent edges to better capture local structual information. The four objectives are summarized in Table 1 and the details will be discussed in Appendix. D.

Method	Loss function	Sampled items
Residue Type Prediction	$\mathcal{L}_i = \text{CE}(f_{\text{residue}}(\boldsymbol{h}'_i), \boldsymbol{f}_i)$	Single residue
Distance Prediction	$\mathcal{L}_{(i,j,r)} = (f_{\text{dist}}(\boldsymbol{h}'_i, \boldsymbol{h}'_j) - \lVert \boldsymbol{x}_i - \boldsymbol{x}_j \rVert_2)^2$	Single edge
Angle Prediction	$\mathcal{L}_{(i,j,r_1),(j,k,r_2)} = \text{CE}(f_{\text{angle}}(\boldsymbol{h}'_i, \boldsymbol{h}'_j, \boldsymbol{h}'_k), \text{bin}(\angle ijk))$	Adjacent edge pairs
Dihedral Prediction	$\mathcal{L}_{(i,j,r_1),(j,k,r_2),(k,t,r_3)} = \text{CE}(f_{\text{dih}}(\boldsymbol{h}'_i, \boldsymbol{h}'_j, \boldsymbol{h}'_k, \boldsymbol{h}'_t), \text{bin}(\angle ijkt))$	Adjacent edge triplets

Table 1: Self-prediction methods. We use $f_{\text{residue}}(\cdot)$, $f_{\text{dist}}(\cdot)$, $f_{\text{angle}}(\cdot)$, $f_{\text{dih}}(\cdot)$ to denote the MLP head in each task. $\text{CE}(\cdot)$ denotes the cross entropy loss and $\text{bin}(\cdot)$ is used to discretize the angle. $\boldsymbol{f}_i$ and $\boldsymbol{x}_i$ denote the residue type feature and coordinate of node i, respectively. $\boldsymbol{h}'_i$ denotes the representation of node i after masking the corresponding sampled items in each task.

5 EXPERIMENTS

In this section, we first introduce our experimental setup for pretraining and then evaluate our models on four standard downstream tasks including Enzyme Commission number prediction, Gene Ontology term prediction, fold classification and reaction classification.

5.1 EXPERIMENTAL SETUP

Pretraining datasets. We use the AlphaFold protein structure database (CC-BY 4.0 License) (Jumper et al., 2021; Varadi et al., 2021) for pretraining. This database contains the protein structures predicted by the AlphaFold2 model, and we employ both 365K proteome-wide predictions and 440K Swiss-Prot (Consortium, 2021) predictions in our experiments. In Appendix F, we further report the results of pretraining on different datasets.

Downstream tasks. We adopt two tasks proposed in Gligorijević et al. (2021) and two tasks used in Hermosilla et al. (2021) for downstream evaluation. ***Enzyme Commission (EC) number prediction*** seeks to predict the EC numbers of different proteins, which describe their catalysis of biochemical reactions. The EC numbers are selected from the third and fourth levels of the EC tree (Webb et al., 1992), forming 538 binary classification tasks. ***Gene Ontology (GO) term prediction*** aims to predict whether a protein belongs to some GO terms. These terms classify proteins into hierarchically related functional classes organized into three ontologies: molecular function (MF), biological process (BP) and cellular component (CC). ***Fold classification*** is first proposed in Hou et al. (2018), with the goal to predict the fold class label given a protein. ***Reaction classification*** aims to predict the enzyme-catalyzed reaction class of a protein, in which all four levels of the EC number are employed to depict the reaction class. Although this task is essentially the same with EC prediction, we include it to make a fair comparison with the baselines in Hermosilla et al. (2021).

Dataset splits. For EC and GO prediction, we follow the multi-cutoff split methods in Gligorijević et al. (2021) to ensure that the test set only contains PDB chains with sequence identity no more than 95% to the training set as used in Wang et al. (2022b) (See Appendix. F for results at lower identity cutoffs). For fold classification, Hou et al. (2018) provides three different test sets: *Fold*, in which proteins from the same superfamily are unseen during training; *Superfamily*, in which proteins from the same family are not present during training; and *Family*, in which proteins from the same family are present during training. For reaction classification, we adopt dataset splits proposed in Hermosilla et al. (2021), where proteins have less than 50% sequence similarity in-between splits.

Baselines. Following Wang et al. (2022b) and Hermosilla et al. (2021), we compare our encoders with many existing protein representation learning methods, including four sequence-based encoders (CNN (Shanehsazzadeh et al., 2020), ResNet (Rao et al., 2019), LSTM (Rao et al., 2019) and Transformer (Rao et al., 2019)), six structure-based encoders (GCN (Kipf & Welling, 2017), GAT (Veličković et al., 2018), GVP (Jing et al., 2021), 3DCNN_MQA (Derevyanko et al., 2018), GraphQA (Baldassarre et al., 2021) and New IEConv (Hermosilla & Ropinski, 2022)). We also include two models pretrained on large-scale sequence datasets (ProtBERT-BFD (Elnaggar et al., 2021), ESM-1b (Rives et al., 2021)) and two models combining pretrained sequence-based encoders with structural information (DeepFRI (Gligorijević et al., 2021) and LM-GVP (Wang et al., 2022b)). For LM-GVP and New IEConv, we only include results reported in the original paper due to the computational burden and the lack of codes. We do not include MSA-based baselines, since these

Table 2: F_{max} on EC and GO prediction and Accuracy (%) on fold and reaction classification. [†] denotes results taken from Wang et al. (2022b) and [*] denotes results taken from Hermosilla et al. (2021) and Hermosilla & Ropinski (2022). Bold numbers indicate the best results under w/o pretraining and w/ pretraining settings. For pretraining, we select the model with the best performance when training from scratch, *i.e.*, GearNet-Edge for EC, GO, Reaction and GearNet-Edge-IEConv for Fold Classification. We omit the model name and use the pretraining methods to name our pretrained models.

	Method	Pretraining Dataset (Size)	EC	GO			Fold Classification				Reaction
				BP	MF	CC	Fold	Super.	Fam.	Avg.	
w/o pretraining	CNN (Shanehsazzadeh et al., 2020)	-	0.545	0.244	0.354	0.287	11.3	13.4	53.4	26.0	51.7
	ResNet (Rao et al., 2019)	-	0.605	0.280	0.405	0.304	10.1	7.21	23.5	13.6	24.1
	LSTM (Rao et al., 2019)	-	0.425	0.225	0.321	0.283	6.41	4.33	18.1	9.61	11.0
	Transformer (Rao et al., 2019)	-	0.238	0.264	0.211	0.405	9.22	8.81	40.4	19.4	26.6
	GCN (Kipf & Welling, 2017)	-	0.320	0.252	0.195	0.329	16.8*	21.3*	82.8*	40.3*	67.3*
	GAT (Veličković et al., 2018)	-	0.368	0.284[†]	0.317[†]	0.385[†]	12.4	16.5	72.7	33.8	55.6
	GVP (Jing et al., 2021)	-	0.489	0.326[†]	0.426[†]	0.420[†]	16.0	22.5	83.8	40.7	65.5
	3DCNN_MQA (Derevyanko et al., 2018)	-	0.077	0.240	0.147	0.305	31.6*	45.4*	92.5*	56.5*	72.2*
	GraphQA (Baldassarre et al., 2021)	-	0.509	0.308	0.329	0.413	23.7*	32.5*	84.4*	46.9*	60.8*
	New IEConv (Hermosilla & Ropinski, 2022)	-	0.735	0.374	0.544	0.444	47.6*	70.2*	99.2*	72.3*	**87.2***
	GearNet	-	0.730	0.356	0.503	0.414	28.4	42.6	95.3	55.4	79.4
	GearNet-IEConv	-	0.800	0.381	0.563	0.422	42.3	64.1	99.1	68.5	83.7
	GearNet-Edge	-	**0.810**	**0.403**	**0.580**	**0.450**	44.0	66.7	99.1	69.9	86.6
	GearNet-Edge-IEConv	-	**0.810**	**0.400**	**0.581**	0.430	**48.3**	**70.3**	**99.5**	**72.7**	85.3
w/ pretraining	DeepFRI (Gligorijević et al., 2021)	Pfam (10M)	0.631	0.399	0.465	0.460	15.3*	20.6*	73.2*	36.4*	63.3*
	ESM-1b (Rives et al., 2021)	UniRef50 (24M)	0.864	0.452	**0.657**	0.477	26.8	60.1	97.8	61.5	83.1
	ProtBERT-BFD (Elnaggar et al., 2021)	BFD (2.1B)	0.838	0.279[†]	0.456[†]	0.408[†]	26.6*	55.8*	97.6*	60.0*	72.2*
	LM-GVP (Wang et al., 2022b)	UniRef100 (216M)	0.664	0.417[†]	0.545[†]	**0.527**[†]	-	-	-	-	-
	New IEConv (Hermosilla & Ropinski, 2022)	PDB (476K)	-	-	-	-	50.3*	**80.6***	99.7*	76.9*	**87.6***
	Residue Type Prediction	AlphaFoldDB (805K)	0.843	0.430	0.604	0.465	48.8	71.0	99.4	73.0	86.6
	Distance Prediction	AlphaFoldDB (805K)	0.839	0.448	0.616	0.464	50.9	73.5	99.4	74.6	**87.5**
	Angle Prediction	AlphaFoldDB (805K)	0.853	0.458	0.625	0.473	**56.5**	76.3	99.6	77.4	86.8
	Dihedral Prediction	AlphaFoldDB (805K)	0.859	0.458	0.626	0.465	51.8	77.8	99.6	75.9	87.0
	Multiview Contrast	AlphaFoldDB (805K)	**0.874**	**0.490**	**0.654**	0.488	54.1	**80.5**	**99.9**	**78.1**	**87.5**

evolution-based methods require a lot of resources for the computation and storage of MSAs but have been shown to be inferior to ESM-1b on function prediction tasks in Hu et al. (2022).

Training. On the four downstream tasks, we train GearNet and GearNet-Edge from scratch. As we find that the IEConv layer is important for predicting fold labels, we also enhance our model by incorporating this as an additional layer (details in Appendix C.2). These models are referred as GearNet-IEConv and GearNet-Edge-IEConv, respectively. Following Wang et al. (2022b) and Hermosilla & Ropinski (2022), the models are trained for 200 epochs on EC and GO prediction and for 300 epochs on fold and reaction classification. For pretraining, the models with the best performance when trained from scratch are selected, *i.e.*, GearNet-Edge for EC, GO, Reaction and GearNet-Edge-IEConv for Fold Classification. The models are pretrained on the AlphaFold protein database with our proposed five methods for 50 epochs. All these models are trained on 4 Tesla A100 GPUs. More details can be found in Appendix E.3.

Evaluation. For EC and GO prediction, we evaluate the performance with the protein-centric maximum F-score F_{max}, which is commonly used in the CAFA challenges (Radivojac et al., 2013) (See Appendix E.2 for details). For fold and reaction classification, the performance is measured with the mean accuracy. Models with the best performance on validation sets are selected for evaluation.

5.2 Results

We report results for four downstream tasks in Table 2, including all models with and without pretraining. The following conclusions can be drawn from the results:

Our structure-based encoders outperform all baselines without pretraining on 7 of 8 datasets. By comparing the first three blocks, we find that the vanilla GearNet can already obtain competitive results against other baselines on three function prediction tasks (EC, GO, Reaction). After adding the edge message passing mechanism, GearNet-Edge significantly outperforms other baselines on EC, GO-BP and GO-MF and achieves competitive results on GO-CC. Although no clear improvements are observed on function prediction tasks (EC, GO, Reaction) by adding IEConv layers, GearNet-Edge-IEConv can achieve the best results on the fold classification task. This can be understood since fold classification requires the encoder to capture sufficient structural information for determining the

fold labels of proteins. Compared with GearNet-Edge, which only includes the distance and angle information as features, the IEConv layer is better at capturing structural details by applying different kernel matrices dependent on relative positional features. These strong performance demonstrates the advantages of our proposed structure-based encoders.

Structure-based encoders benefit a lot from pretraining with unlabeled structures. Comparing the results in the third and last two blocks, it can be observed that models with all proposed pretraining methods show large improvements over models trained from scratch. Among these methods, Multiview Contrast is the best on 7 of 8 datasets and achieve the state-of-the-art results on EC, GO-BP, GO-MF, Fold and Reaction tasks. This proves the effectiveness of our pretraining strategies.

Pretrained structure-based encoders perform on par with or even better than sequence-based encoders pretrained with much more data. The last three blocks show the comparision between pretrained sequence-based and structure-based models. It should be noted that our models are pretrained on a dataset with fewer than one million structures, whereas all sequence-based pretraining baselines are pretrained on million- or billion-scale sequence databases. Though pretrained with an order of magnitude less data, our model can achieve comparable or even better results against these sequence-based models. Besides, our model is the only one that can achieve good performance on all four tasks, given that sequence-based models do not perform well on fold classification. This again shows the potential of structure-based pretraining for learning protein representations.

5.3 Ablation Studies

To analyze the contribution of different components in our proposed methods, we perform ablation studies on the EC prediction task. The results are shown in Table 3.

Relational graph convolutional layers. To show the effects of relational convolutional layers, we replace it with graph convolutional layers that share a single kernel matrix among all edges. As reported in the table, results can be significantly improved by using relational convolution, which suggests the importance of treating edges as different types.

Method	F_{max}
GearNet-Edge	**0.810**
- w/o relational convolution	0.752
- w/o edge message passing	0.730
GearNet-Edge (Multiview Contrast)	**0.874**
- subsequence + identity	0.866
- subspace+ identity	**0.872**
- subsequence + random edge masking	0.869
- subspace + random edge masking	**0.876**

Table 3: Ablation studies on EC.

Edge message passing layers. We also compare the results of GearNet with and without edge message passing layers, the results of which are shown in Tables 2, 3. It can be observed that the performance consistently increases after performing edge message passing. This demonstrates the effectiveness of our proposed mechanism.

Different augmentations in Multiview Contrast. We investigate the contribution of each augmentation operation proposed in the Multiview Contrast method. Instead of randomly sampling cropping and noise functions, we pretrain our model with four deterministic combinations of augmentations, respectively. As shown in Table 3, all the four combinations can yield good results, which suggests that arbitrary combinations of the proposed cropping and noise schemes can yield informative partial views of proteins. Besides, it can be observed that the results of Subspace Cropping are consistently better than those of Subsequence Cropping with different noise functions, which implies that it is a better way to utilize 3D information to extract meaningful protein substructures.

6 Conclusions and Future Work

In this work, we propose a simple yet effective structure-based encoder for protein representation learning, which performs relational message passing on protein residue graphs. A novel edge message passing mechanism is introduced to explicitly model interactions between edges, which shows consistent improvements. Moreover, a multiview contrastive learning method is proposed for pretraining accompanied with four self-prediction baselines. Comprehensive experiments over multiple benchmark tasks verify that our model pretrained with Multiview Contrast outperforms previous encoders when trained from scratch and achieve comparable or even better results compared to the state-of-the-art baselines while pretraining with much less data. We believe that our work is an important step towards adopting self-supervised learning methods on protein structure understanding.

REPRODUCIBILITY STATEMENT

To guarantee the reproducibility of our method, we provide all the training details needed for reproducing the results in the paper. The datasets and splits are discussed in Section 5.1 and publicly available (Varadi et al., 2021; Gligorijević et al., 2021; Hermosilla et al., 2021). We also report all the hyperparameters and comprehensive implementation details in Appendix E.3. All codes and pretrained models will be released once the paper is published.

REFERENCES

Mehmet Akdel, Douglas EV Pires, Eduard Porta Pardo, Jürgen Jänes, Arthur O Zalevsky, Bálint Mészáros, Patrick Bryant, Lydia L Good, Roman A Laskowski, Gabriele Pozzati, et al. A structural biology community assessment of alphafold 2 applications. *bioRxiv*, 2021.

Ethan C Alley, Grigory Khimulya, Surojit Biswas, Mohammed AlQuraishi, and George M Church. Unified rational protein engineering with sequence-based deep representation learning. *Nature methods*, 16(12):1315–1322, 2019.

Sarp Aykent and Tian Xia. Gbpnet: Universal geometric representation learning on protein structures. *Proceedings of the 28th ACM SIGKDD Conference on Knowledge Discovery and Data Mining*, 2022.

Minkyung Baek, Frank DiMaio, Ivan Anishchenko, Justas Dauparas, Sergey Ovchinnikov, Gyu Rie Lee, Jue Wang, Qian Cong, Lisa N Kinch, R Dustin Schaeffer, et al. Accurate prediction of protein structures and interactions using a three-track neural network. *Science*, 373(6557):871–876, 2021.

Federico Baldassarre, David Menéndez Hurtado, Arne Elofsson, and Hossein Azizpour. Graphqa: protein model quality assessment using graph convolutional networks. *Bioinformatics*, 37(3): 360–366, 2021.

Albert P Bartók, Mike C Payne, Risi Kondor, and Gábor Csányi. Gaussian approximation potentials: The accuracy of quantum mechanics, without the electrons. *Physical review letters*, 104(13): 136403, 2010.

Albert P Bartók, Risi Kondor, and Gábor Csányi. On representing chemical environments. *Physical Review B*, 87(18):184115, 2013.

Jörg Behler and Michele Parrinello. Generalized neural-network representation of high-dimensional potential-energy surfaces. *Physical review letters*, 98(14):146401, 2007.

Tristan Bepler and Bonnie Berger. Learning protein sequence embeddings using information from structure. *arXiv preprint arXiv:1902.08661*, 2019.

Tristan Bepler and Bonnie Berger. Learning the protein language: Evolution, structure, and function. *Cell Systems*, 12(6):654–669, 2021.

Helen M Berman, John Westbrook, Zukang Feng, Gary Gilliland, Talapady N Bhat, Helge Weissig, Ilya N Shindyalov, and Philip E Bourne. The protein data bank. *Nucleic acids research*, 28(1): 235–242, 2000.

Surojit Biswas, Grigory Khimulya, Ethan C Alley, Kevin M Esvelt, and George M Church. Low-n protein engineering with data-efficient deep learning. *Nature Methods*, 18(4):389–396, 2021.

Tom B Brown, Benjamin Mann, Nick Ryder, Melanie Subbiah, Jared Kaplan, Prafulla Dhariwal, Arvind Neelakantan, Pranav Shyam, Girish Sastry, Amanda Askell, et al. Language models are few-shot learners. *arXiv preprint arXiv:2005.14165*, 2020.

Yue Cao, Payel Das, Vijil Chenthamarakshan, Pin-Yu Chen, Igor Melnyk, and Yang Shen. Fold2seq: A joint sequence(1d)-fold(3d) embedding-based generative model for protein design. In Marina Meila and Tong Zhang (eds.), *Proceedings of the 38th International Conference on Machine Learning*, volume 139 of *Proceedings of Machine Learning Research*, pp. 1261–1271. PMLR, 18–24 Jul 2021. URL `https://proceedings.mlr.press/v139/cao21a.html`.

Can Chen, Jingbo Zhou, Fan Wang, Xue Liu, and Dejing Dou. Structure-aware protein self-supervised learning. *arXiv preprint arXiv:2204.04213*, 2022.

Chi Chen, Weike Ye, Yunxing Zuo, Chen Zheng, and Shyue Ping Ong. Graph networks as a universal machine learning framework for molecules and crystals. *Chemistry of Materials*, 31(9):3564–3572, 2019.

Ting Chen, Simon Kornblith, Mohammad Norouzi, and Geoffrey Hinton. A simple framework for contrastive learning of visual representations. In *International conference on machine learning*, pp. 1597–1607. PMLR, 2020.

Stefan Chmiela, Alexandre Tkatchenko, Huziel E Sauceda, Igor Poltavsky, Kristof T Schütt, and Klaus-Robert Müller. Machine learning of accurate energy-conserving molecular force fields. *Science advances*, 3(5):e1603015, 2017.

The UniProt Consortium. Uniprot: the universal protein knowledgebase in 2021. *Nucleic Acids Research*, 49(D1):D480–D489, 2021.

Bowen Dai and Chris Bailey-Kellogg. Protein interaction interface region prediction by geometric deep learning. *Bioinformatics*, 2021.

Georgy Derevyanko, Sergei Grudinin, Yoshua Bengio, and Guillaume Lamoureux. Deep convolutional networks for quality assessment of protein folds. *Bioinformatics*, 34(23):4046–4053, 2018.

Jacob Devlin, Ming-Wei Chang, Kenton Lee, and Kristina Toutanova. Bert: Pre-training of deep bidirectional transformers for language understanding. *arXiv preprint arXiv:1810.04805*, 2018.

Ahmed Elnaggar, Michael Heinzinger, Christian Dallago, Ghalia Rehawi, Wang Yu, Llion Jones, Tom Gibbs, Tamas Feher, Christoph Angerer, Martin Steinegger, Debsindhu Bhowmik, and Burkhard Rost. Prottrans: Towards cracking the language of lifes code through self-supervised deep learning and high performance computing. *IEEE Transactions on Pattern Analysis and Machine Intelligence*, pp. 1–1, 2021. doi: 10.1109/TPAMI.2021.3095381.

Pablo Gainza, Freyr Sverrisson, Frederico Monti, Emanuele Rodola, D Boscaini, MM Bronstein, and BE Correia. Deciphering interaction fingerprints from protein molecular surfaces using geometric deep learning. *Nature Methods*, 17(2):184–192, 2020.

Justin Gilmer, Samuel S Schoenholz, Patrick F Riley, Oriol Vinyals, and George E Dahl. Neural message passing for quantum chemistry. In *International conference on machine learning*, pp. 1263–1272. PMLR, 2017.

Vladimir Gligorijević, P Douglas Renfrew, Tomasz Kosciolek, Julia Koehler Leman, Daniel Berenberg, Tommi Vatanen, Chris Chandler, Bryn C Taylor, Ian M Fisk, Hera Vlamakis, et al. Structure-based protein function prediction using graph convolutional networks. *Nature communications*, 12 (1):1–14, 2021.

Yuzhi Guo, Jiaxiang Wu, Hehuan Ma, and Junzhou Huang. Self-supervised pre-training for protein embeddings using tertiary structures. In *AAAI*, 2022.

William L Hamilton, Rex Ying, and Jure Leskovec. Inductive representation learning on large graphs. In *Proceedings of the 31st International Conference on Neural Information Processing Systems*, pp. 1025–1035, 2017.

Frank Harary and Robert Z Norman. Some properties of line digraphs. *Rendiconti del circolo matematico di palermo*, 9(2):161–168, 1960.

Kaveh Hassani and Amir Hosein Khasahmadi. Contrastive multi-view representation learning on graphs. In *International Conference on Machine Learning*, pp. 4116–4126. PMLR, 2020.

Kaiming He, Haoqi Fan, Yuxin Wu, Saining Xie, and Ross Girshick. Momentum contrast for unsupervised visual representation learning. In *Proceedings of the IEEE/CVF Conference on Computer Vision and Pattern Recognition*, pp. 9729–9738, 2020.

Liang He, Shizhuo Zhang, Lijun Wu, Huanhuan Xia, Fusong Ju, He Zhang, Siyuan Liu, Yingce Xia, Jianwei Zhu, Pan Deng, et al. Pre-training co-evolutionary protein representation via a pairwise masked language model. *arXiv preprint arXiv:2110.15527*, 2021.

Dan Hendrycks and Kevin Gimpel. Gaussian error linear units (gelus). *arXiv preprint arXiv:1606.08415*, 2016.

Pedro Hermosilla and Timo Ropinski. Contrastive representation learning for 3d protein structures. In *Submitted to The Tenth International Conference on Learning Representations*, 2022. URL `https://openreview.net/forum?id=VINWzIM6_6`.

Pedro Hermosilla, Marco Schäfer, Matěj Lang, Gloria Fackelmann, Pere Pau Vázquez, Barbora Kozlíková, Michael Krone, Tobias Ritschel, and Timo Ropinski. Intrinsic-extrinsic convolution and pooling for learning on 3d protein structures. *International Conference on Learning Representations*, 2021.

Jie Hou, Badri Adhikari, and Jianlin Cheng. Deepsf: deep convolutional neural network for mapping protein sequences to folds. *Bioinformatics*, 34(8):1295–1303, 2018.

Min Hu, Fajie Yuan, Kevin Kaichuang Yang, Fusong Ju, Jingyu Su, Hongya Wang, Fei Yang, and Qiuyang Ding. Exploring evolution-based & -free protein language models as protein function predictors. *ArXiv*, abs/2206.06583, 2022.

Weihua Hu, Bowen Liu, Joseph Gomes, Marinka Zitnik, Percy Liang, Vijay Pande, and Jure Leskovec. Strategies for pre-training graph neural networks. *arXiv preprint arXiv:1905.12265*, 2019.

John Ingraham, Vikas Garg, Regina Barzilay, and Tommi Jaakkola. Generative models for graph-based protein design. *Advances in Neural Information Processing Systems*, 32:15820–15831, 2019.

Bowen Jing, Stephan Eismann, Pratham N. Soni, and Ron O. Dror. Learning from protein structure with geometric vector perceptrons. In *International Conference on Learning Representations*, 2021. URL `https://openreview.net/forum?id=1YLJDvSx6J4`.

Peter Bjørn Jørgensen, Karsten Wedel Jacobsen, and Mikkel N Schmidt. Neural message passing with edge updates for predicting properties of molecules and materials. *arXiv preprint arXiv:1806.03146*, 2018.

John Jumper, Richard Evans, Alexander Pritzel, Tim Green, Michael Figurnov, Olaf Ronneberger, Kathryn Tunyasuvunakool, Russ Bates, Augustin Žídek, Anna Potapenko, et al. Highly accurate protein structure prediction with alphafold. *Nature*, 596(7873):583–589, 2021.

Thomas N Kipf and Max Welling. Variational graph auto-encoders. *arXiv preprint arXiv:1611.07308*, 2016.

Thomas N. Kipf and Max Welling. Semi-supervised classification with graph convolutional networks. In *International Conference on Learning Representations*, 2017.

Johannes Klicpera, Janek Groß, and Stephan Günnemann. Directional message passing for molecular graphs. In *International Conference on Learning Representations (ICLR)*, 2020.

Johannes Klicpera, Florian Becker, and Stephan Günnemann. Gemnet: Universal directional graph neural networks for molecules. *arXiv preprint arXiv:2106.08903*, 2021.

Yi Liu, Limei Wang, Meng Liu, Xuan Zhang, Bora Oztekin, and Shuiwang Ji. Spherical message passing for 3d graph networks. *arXiv preprint arXiv:2102.05013*, 2021.

Amy X Lu, Haoran Zhang, Marzyeh Ghassemi, and Alan M Moses. Self-supervised contrastive learning of protein representations by mutual information maximization. *BioRxiv*, 2020.

Bin Ma. Novor: real-time peptide de novo sequencing software. *Journal of the American Society for Mass Spectrometry*, 26(11):1885–1894, 2015.

Bin Ma and Richard Johnson. De novo sequencing and homology searching. *Molecular & cellular proteomics*, 11(2), 2012.

Craig O. Mackenzie and Gevorg Grigoryan. Protein structural motifs in prediction and design. *Current opinion in structural biology*, 44:161–167, 2017.

Leland McInnes, John Healy, and James Melville. Umap: Uniform manifold approximation and projection for dimension reduction. *arXiv preprint arXiv:1802.03426*, 2018.

Joshua Meier, Roshan Rao, Robert Verkuil, Jason Liu, Tom Sercu, and Alexander Rives. Language models enable zero-shot prediction of the effects of mutations on protein function. *bioRxiv*, 2021.

Jaina Mistry, Sara Chuguransky, Lowri Williams, Matloob Qureshi, Gustavo A Salazar, Erik LL Sonnhammer, Silvio CE Tosatto, Lisanna Paladin, Shriya Raj, Lorna J Richardson, et al. Pfam: The protein families database in 2021. *Nucleic Acids Research*, 49(D1):D412–D419, 2021.

Alexey G Murzin, Steven E Brenner, Tim Hubbard, and Cyrus Chothia. Scop: a structural classification of proteins database for the investigation of sequences and structures. *Journal of molecular biology*, 247(4):536–540, 1995.

Aaron van den Oord, Yazhe Li, and Oriol Vinyals. Representation learning with contrastive predictive coding. *arXiv preprint arXiv:1807.03748*, 2018.

Chris Paul Ponting and Robert R Russell. The natural history of protein domains. *Annual review of biophysics and biomolecular structure*, 31:45–71, 2002.

Jiezhong Qiu, Qibin Chen, Yuxiao Dong, Jing Zhang, Hongxia Yang, Ming Ding, Kuansan Wang, and Jie Tang. Gcc: Graph contrastive coding for graph neural network pre-training. In *Proceedings of the 26th ACM SIGKDD International Conference on Knowledge Discovery & Data Mining*, pp. 1150–1160, 2020.

Predrag Radivojac, Wyatt T Clark, Tal Ronnen Oron, Alexandra M Schnoes, Tobias Wittkop, Artem Sokolov, Kiley Graim, Christopher Funk, Karin Verspoor, Asa Ben-Hur, et al. A large-scale evaluation of computational protein function prediction. *Nature methods*, 10(3):221–227, 2013.

Roshan Rao, Nicholas Bhattacharya, Neil Thomas, Yan Duan, Xi Chen, John Canny, Pieter Abbeel, and Yun S Song. Evaluating protein transfer learning with tape. In *Advances in Neural Information Processing Systems*, 2019.

Roshan Rao, Jason Liu, Robert Verkuil, Joshua Meier, John F Canny, Pieter Abbeel, Tom Sercu, and Alexander Rives. Msa transformer. *bioRxiv*, 2021.

Alexander Rives, Joshua Meier, Tom Sercu, Siddharth Goyal, Zeming Lin, Jason Liu, Demi Guo, Myle Ott, C Lawrence Zitnick, Jerry Ma, et al. Biological structure and function emerge from scaling unsupervised learning to 250 million protein sequences. *Proceedings of the National Academy of Sciences*, 118(15), 2021.

Yu Rong, Yatao Bian, Tingyang Xu, Weiyang Xie, Ying Wei, Wenbing Huang, and Junzhou Huang. Self-supervised graph transformer on large-scale molecular data. *arXiv preprint arXiv:2007.02835*, 2020.

Olga Russakovsky, Jia Deng, Hao Su, Jonathan Krause, Sanjeev Satheesh, Sean Ma, Zhiheng Huang, Andrej Karpathy, Aditya Khosla, Michael Bernstein, et al. Imagenet large scale visual recognition challenge. *International journal of computer vision*, 115(3):211–252, 2015.

Michael Schlichtkrull, Thomas N Kipf, Peter Bloem, Rianne Van Den Berg, Ivan Titov, and Max Welling. Modeling relational data with graph convolutional networks. In *European semantic web conference*, pp. 593–607. Springer, 2018.

Kristof T Schütt, Farhad Arbabzadah, Stefan Chmiela, Klaus R Müller, and Alexandre Tkatchenko. Quantum-chemical insights from deep tensor neural networks. *Nature communications*, 8(1):1–8, 2017a.

Kristof T Schütt, Pieter-Jan Kindermans, Huziel E Sauceda, Stefan Chmiela, Alexandre Tkatchenko, and Klaus-Robert Müller. Schnet: A continuous-filter convolutional neural network for modeling quantum interactions. *arXiv preprint arXiv:1706.08566*, 2017b.

Amir Shanehsazzadeh, David Belanger, and David Dohan. Is transfer learning necessary for protein landscape prediction? *arXiv preprint arXiv:2011.03443*, 2020.

Vignesh Ram Somnath, Charlotte Bunne, and Andreas Krause. Multi-scale representation learning on proteins. *Advances in Neural Information Processing Systems*, 34, 2021.

Martin Steinegger and Johannes Söding. Clustering huge protein sequence sets in linear time. *Nature communications*, 9(1):1–8, 2018.

Alexey Strokach, David Becerra, Carles Corbi-Verge, Albert Perez-Riba, and Philip M Kim. Fast and flexible protein design using deep graph neural networks. *Cell Systems*, 11(4):402–411, 2020.

Mukund Sundararajan, Ankur Taly, and Qiqi Yan. Axiomatic attribution for deep networks. In *International Conference on Machine Learning*, pp. 3319–3328. PMLR, 2017.

Baris E Suzek, Hongzhan Huang, Peter McGarvey, Raja Mazumder, and Cathy H Wu. Uniref: comprehensive and non-redundant uniprot reference clusters. *Bioinformatics*, 23(10):1282–1288, 2007.

Freyr Sverrisson, Jean Feydy, Bruno E Correia, and Michael M Bronstein. Fast end-to-end learning on protein surfaces. In *Proceedings of the IEEE/CVF Conference on Computer Vision and Pattern Recognition*, pp. 15272–15281, 2021.

Y Tateno, K Ikeo, T Imanishi, H Watanabe, T Endo, Y Yamaguchi, Y Suzuki, K Takahashi, K Tsunoyama, M Kawai, et al. Evolutionary motif and its biological and structural significance. *Journal of molecular evolution*, 44(1):S38–S43, 1997.

Mihaly Varadi, Stephen Anyango, Mandar Deshpande, Sreenath Nair, Cindy Natassia, Galabina Yordanova, David Yuan, Oana Stroe, Gemma Wood, Agata Laydon, et al. Alphafold protein structure database: massively expanding the structural coverage of protein-sequence space with high-accuracy models. *Nucleic acids research*, 2021.

Ashish Vaswani, Noam Shazeer, Niki Parmar, Jakob Uszkoreit, Llion Jones, Aidan N Gomez, Łukasz Kaiser, and Illia Polosukhin. Attention is all you need. In *Advances in neural information processing systems*, pp. 5998–6008, 2017.

Petar Veličković, Guillem Cucurull, Arantxa Casanova, Adriana Romero, Pietro Liò, and Yoshua Bengio. Graph Attention Networks. *International Conference on Learning Representations*, 2018. URL `https://openreview.net/forum?id=rJXMpikCZ`. accepted as poster.

Limei Wang, Haoran Liu, Yi Liu, Jerry Kurtin, and Shuiwang Ji. Learning protein representations via complete 3d graph networks. *ArXiv*, abs/2207.12600, 2022a.

Yanbin Wang, Zhu-Hong You, Shan Yang, Xiao Li, Tong-Hai Jiang, and Xi Zhou. A high efficient biological language model for predicting protein–protein interactions. *Cells*, 8(2):122, 2019.

Zichen Wang, Steven A Combs, Ryan Brand, Miguel Romero Calvo, Panpan Xu, George Price, Nataliya Golovach, Emmanuel O Salawu, Colby J Wise, Sri Priya Ponnapalli, et al. Lm-gvp: an extensible sequence and structure informed deep learning framework for protein property prediction. *Scientific reports*, 12(1):1–12, 2022b.

Oren F Webb, Tommy J Phelps, Paul R Bienkowski, Philip M Digrazia, David C White, and Gary S Sayler. Enzyme nomenclature. 1992.

Minghao Xu, Hang Wang, Bingbing Ni, Hongyu Guo, and Jian Tang. Self-supervised graph-level representation learning with local and global structure. *arXiv preprint arXiv:2106.04113*, 2021.

Kevin Kaichuang Yang, Niccol'o Zanichelli, and Hugh Yeh. Masked inverse folding with sequence transfer for protein representation learning. *bioRxiv*, 2022.

Yuning You, Tianlong Chen, Yongduo Sui, Ting Chen, Zhangyang Wang, and Yang Shen. Graph contrastive learning with augmentations. *Advances in Neural Information Processing Systems*, 33: 5812–5823, 2020.

Zhaocheng Zhu, Chence Shi, Zuobai Zhang, Shengchao Liu, Minghao Xu, Xinyu Yuan, Yangtian Zhang, Junkun Chen, Huiyu Cai, Jiarui Lu, et al. Torchdrug: A powerful and flexible machine learning platform for drug discovery. *arXiv preprint arXiv:2202.08320*, 2022.

A MORE RELATED WORK

A.1 SEQUENCE-BASED METHODS FOR PROTEIN REPRESENTATION LEARNING

Sequence-based protein representation learning is mainly inspired by the methods of modeling natural language sequences. Recent methods aim to capture the biochemical and co-evolutionary knowledge underlying a large-scale protein sequence corpus by self-supervised pretraining, and such knowledge is then transferred to specific downstream tasks by finetuning. Typical pretraining objectives explored in existing methods include next amino acid prediction (Alley et al., 2019; Elnaggar et al., 2021), masked language modeling (MLM) (Rao et al., 2019; Elnaggar et al., 2021; Rives et al., 2021), pairwise MLM (He et al., 2021) and contrastive predictive coding (CPC) (Lu et al., 2020). Compared to sequence-based approaches that learn in the whole sequence space, MSA-based methods (Rao et al., 2021; Biswas et al., 2021; Meier et al., 2021) leverage the sequences within a protein family to capture the conserved and variable regions of homologous sequences, which imply specific structures and functions of the protein family.

A.2 STRUCTURE-BASED METHODS FOR BIOLOGICAL MOLECULES

Following the early efforts (Behler & Parrinello, 2007; Bartók et al., 2010; 2013; Chmiela et al., 2017) of building machine learning systems for molecules by hand-crafted atomic features, recent works exploited end-to-end message passing neural networks (MPNNs) (Gilmer et al., 2017) to encode the structures of small molecules and macromolecules like proteins. Specifically, existing methods employed node/atom message passing (Gilmer et al., 2017; Schütt et al., 2017a;b), edge/bond message passing (Jørgensen et al., 2018; Chen et al., 2019) and directional information (Klicpera et al., 2020; Liu et al., 2021; Klicpera et al., 2021) to encode 2D or 3D molecular graphs.

Compared to small molecules, structural representations of proteins are more diverse, including residue-level, atom-level graphs and protein surfaces. There are recent models designed for residue-level graphs (Hermosilla et al., 2021; Hermosilla & Ropinski, 2022) and protein surfaces (Gainza et al., 2020; Sverrisson et al., 2021), which achieved impressive results on various tasks.

A.3 PRETRAINING GRAPH NEURAL NETWORKS

Our work is also related to the recent efforts of pretraining graph neural networks (GNNs), which sought to learn graph representations in a self-supervised fashion. In this domain, various self-supervised pretext tasks, like edge prediction (Kipf & Welling, 2016; Hamilton et al., 2017), context prediction (Hu et al., 2019; Rong et al., 2020), node/edge attribute reconstruction (Hu et al., 2019) and contrastive learning (Hassani & Khasahmadi, 2020; Qiu et al., 2020; You et al., 2020; Xu et al., 2021), are designed to acquire knowledge from unlabeled graphs. In this work, we focus on learning representations of residue-level graphs of proteins in a self-supervised way. To attain this goal, we design novel protein-specific pretraining methods to learn the proposed structure-based encoder.

B POTENTIAL NEGATIVE IMPACT

This research project focuses on learning effective protein representations via pretraining with a large number of unlabeled protein structures. Compared to the conventional sequence-based pretraining methods, our approach is able to leverage structural information and thus provide better representations. This merit enables more in-depth analysis of protein research and can potentially benefit many real-world applications, like protein function prediction and sequence design.

Limitations. In this paper, we only use 805K protein structures for pretraining. As the AlphaFold Protein Structure Database now covers over 100 million proteins, it would be possible to train huge and more advanced structure-based models on larger datasets in the future. Scalability, however, should be kept in mind. Thanks to their simplicity, our models could readily accommodate larger datasets, while more cumbersome procedures might not. Besides, we only consider function and fold prediction tasks. Another promising direction is to apply our proposed methods on more tasks, *e.g.*, protein-protein interaction modeling and protein-guided ligand molecule design, which underpins many important biological processes and applications.

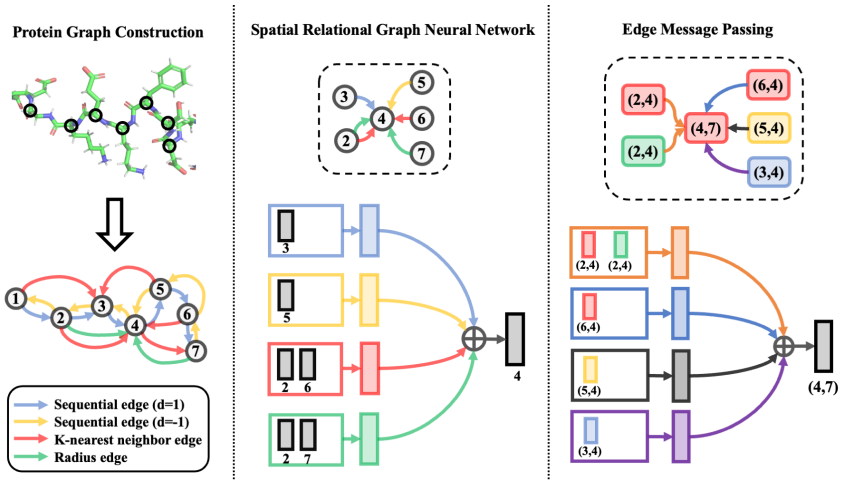

Figure 3: The pipeline for GearNet and GearNet-edge. First, we construct a relational protein residue graph with sequential, radius and knn edges (some edges are omitted in the figure to save space). Then, a relational graph convolutional layer is applied. Similar message passing layers can be applied on the edge graph to improve the model capacity. This figure shows the update iteration for node 4 and edge $(4, 7, \text{red})$, respectively.

It cannot be denied that some harmful activities could be augmented by powerful pretrained models, *e.g.*, designing harmful drugs. We expect future studies will mitigate these issues.

C More Details of GearNet

In this section, we describe more details about the implementation of our GearNet. The whole pipeline of our structure-based encoder is depicted in Figure 3.

C.1 Protein Graph Construction

For graph construction, we use three different ways to add edges:

1. **Sequential edges.** The i-th residue and the j-th residue will be linked by an edge if the sequential distance between them is below a predefined threshold d_{seq}, i.e., $|j - i| < d_{\text{seq}}$. The type of each sequential edge is determined by their relative position $d = j - i$ in the sequence. Hence, there are $2d_{\text{seq}} - 1$ types of sequential edges.
2. **Radius edges.** Following previous works, we also add edges between two nodes i and j when the Euclidean distance between them is smaller than a threshold d_{radius}.
3. **K-nearest neighbor edges.** Since the scales of spatial coordinates may vary among different proteins, a node will be also connected to its k-nearest neighbors based on the Euclidean distance. In this way, the density of spatial edges are guaranteed to be comparable among different protein graphs.

Since we are not interested in spatial edges between residues close with each other in the sequence, we further add a filter to the latter two kinds of edges. Specifically, for a radius or KNN edge connecting the i-th residue and j-th residue, it will be removed if the sequential distance between them is lower than a long range interaction cutoff d_{long}, *i.e.*, $|i - j| < d_{\text{long}}$.

In this paper, we set the sequential distance threshold $d_{\text{seq}} = 3$, the radius $d_{\text{radius}} = 10.0$Å, the number of neighbors $k = 10$ and the long range interaction cutoff $d_{\text{long}} = 5$. By regarding radius edges and KNN edges as two separate edge types, there will be totally $2d_{\text{seq}} + 1 = 7$ different types of edges.

Necessity of spatial edges. Here we explain the necessity of radius and KNN edges by statistics and intuitions. These two kinds of edges result in very different degree distributions. In Figure 4, we plot the average degree distribution over all proteins in AlphaFold Database v1. If we only consider

KNN edges, the node degrees in protein graphs are close to a constant, which makes it difficult to capture those areas with dense interactions between residues. If we only consider radius edges, then there will be about 45,000 proteins with average degrees lower than two. In these sparse graphs, pretraining cannot capture structural information effectively, *e.g.*, Angle Prediction with limited edge pairs and Dihedral Prediction with limited edge triplets. Such sparsity can be hard to overcome by tuning radius cutoff, for the various scales of average distance on different proteins. By simply combining two kinds of edges, we can overcome these issues.

Node and edge features. Most previous structure-based encoders designed for biological molecules (Baldassarre et al., 2021; Hermosilla et al., 2021) used many chemical and spatial features, some of which are difficult to obtain or time-consuming to calculate. In contrast, we only use the one-hot encoding of residue types with one additional dimension for unknown types as node features, denoted as $\boldsymbol{f} \in \{0,1\}^{n\times 21}$, which is enough to learn good representation as shown in our experiments.

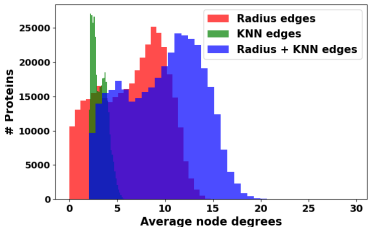

Figure 4: The average degree distribution on AlphaFold Database.

The feature $\boldsymbol{f}_{(i,j,r)}$ for an edge (i,j,r) is the concatenation of the node features of two end nodes, the one-hot encoding of the edge type, and the sequential and spatial distances between them:

$$\boldsymbol{f}_{(i,j,r)} = \text{Cat}\left(\boldsymbol{f}_i, \boldsymbol{f}_j, \text{onehot}(r), |i-j|, \|\boldsymbol{x}_i - \boldsymbol{x}_j\|_2\right), \tag{4}$$

where $\text{Cat}(\cdot)$ denotes the concatenation operation.

C.2 Enhance GearNet with IEConv Layers

In our experiments, we find that IEConv layers are useful for predicting fold labels in spite of their relatively poor performance on function prediction tasks. Therefore, we enhance our models by adding a simplified IEConv layer as an additional layer, which achieve better results than the original IEConv. Next, we describe how to simplify the IEConv layer and how to combine it with our model.

Simplify the IEConv layer. The original IEConv layer relies on intrinsic and extrinsic distances between two nodes, which are computationally expensive. Hence, we follow the modifications proposed in Hermosilla & Ropinski (2022), which show improvements as reported in their experiments. We briefly describe these modifications for completeness.

In the IEConv layer, we keep the edges in our graph $\mathcal{G}$ and use $\tilde{\boldsymbol{h}}_i^{(l)}$ to denote the hidden representation for node i in the l-th layer. The update equation for node i is defined as:

$$\tilde{\boldsymbol{h}}_i^{(l)} = \sum\nolimits_{j\in\mathcal{N}(i)} k_o(f(\mathcal{G},i,j)) \cdot \boldsymbol{h}_j^{(l-1)}, \tag{5}$$

where $\mathcal{N}(i)$ is the set of neighbors of i, $f(\mathcal{G},i,j)$ is the edge feature between i and j and $k_o(\cdot)$ is an MLP mapping the feature to a kernel matrix. Instead of intrinsic and extrinsic distances in the original IEConv layer, we follow New IEConv, which adopts three relative positional features proposed in Ingraham et al. (2019) and further augments them with additional input functions.

We aim to apply this layer on our constructed protein residue graph instead of the radius graph in the original paper. Therefore, we simply remove the dynamically changed receptive fields, pooling layer and smoothing tricks in our setting.

Combine IEConv with GearNet. Our model is very flexible to incorporate other message passing layers. To incorporate IEConv layers, we use our graph and hidden representations as input and replace the update equation Eq. (1) with

$$\boldsymbol{h}_i^{(l)} = \boldsymbol{h}_i^{(l-1)} + \boldsymbol{u}_i^{(l)} + \tilde{\boldsymbol{h}}_i^{(l)}. \tag{6}$$

D SELF-PREDICTION METHODS

The high-level ideas of the four self-prediction methods are demonstrated in Figure 5.

Residue Type Prediction is based on the masked language modeling objective, which has been widely used in pretraining large protein language models (Bepler & Berger, 2021). For each protein, we randomly mask node features of some residues and then predict these masked residue types via structure-based encoders. This method is also known as Attribute Masking in the literature of molecules (Hu et al., 2019) and Masked Inverse Folding in the protein community (Yang et al., 2022).

Distance Prediction aims to learn local spatial structures by predicting the Euclidean distance between two nodes connected in the protein graph. A fixed number of edges are randomly selected and masked from the original graph. Then, the representations of two end nodes will be used to predict the distances between them.

Besides, angles and dihedrals between adjacent edges are also important features that reflect the relative position between residues Klicpera et al. (2021). Similarly, we can define the masked geometric losses **Angle Prediction** and **Dihedral Prediction** by randomly selecting and masking adjacent edge pairs and triplets. Here we discretize the angles and dihedrals by cutting the range $[0, \pi]$ into 8 bins. Then, the representations of end nodes in the masked graph will be used to predict which bin the angles between them will belong to.

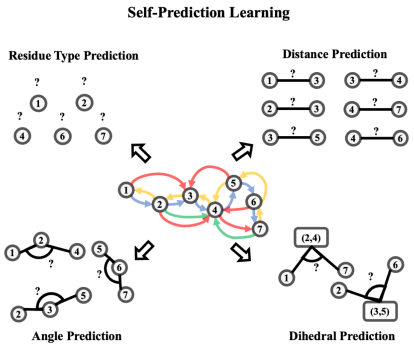

Figure 5: Illustration of self-prediction methods.

E EXPERIMENTAL DETAILS

E.1 DATASET STATISTICS

Table 4: Dataset statistics for downstream tasks.

Dataset	# Proteins		
	# Train	# Validation	# Test
Enzyme Commission	15,550	1,729	1,919
Gene Ontology	29,898	3,322	3,415
Fold Classification - *Fold*	12,312	736	718
Fold Classification - *Superfamily*	12,312	736	1,254
Fold Classification - *Family*	12,312	736	1,272
Reaction Classification	29,215	2,562	5,651

Dataset statistics of downstream tasks are summarized in Table 4. Details are introduced as follows.

Enzyme Commission and Gene Ontology. Following DeepFRI (Gligorijević et al., 2021), the EC numbers are selected from the third and fourth levels of the EC tree, forming 538 binary classification tasks, while the GO terms with at least 50 and no more than 5000 training samples are selected. The non-redundant sets are partitioned into training, validation and test sets according to the sequence identity. We retrieve all protein chains from PDB with the code in their codebase and remove those with obsolete pdb ids, so the statistics will be slightly different from that in the original paper.

Fold Classification. We directly use the dataset in Hermosilla et al. (2021), which consolidated 16,712 proteins with 1,195 different folds from the SCOPe 1.75 database (Murzin et al., 1995).

Reaction Classification. The dataset comprises 37,428 proteins categorized into 384 reaction classes. The split methods are described in Hermosilla et al. (2021), where they cluster protein chains via sequence similarities and ensure that protein chains from the same cluster are in the same split.

E.2 EVALUATION METRICS

Now we introduce the details of evaluation metrics for EC and GO prediction. These two tasks aim to answer the question: whether a protein has some particular functions, which can be seen as multiple binary classification tasks.

The first metric, protein-centric maximum F-score F_{max}, is defined by first calculating the precision and recall for each protein and then taking the average score over all proteins. More specifically, for a given target protein i and a decision threshold $t \in [0, 1]$, the precision and recall are computed as:

$$\text{precision}_i(t) = \frac{\sum_f \mathbb{1}[f \in P_i(t) \cap T_i]}{\sum_f \mathbb{1}[f \in P_i(t)]}, \tag{7}$$

and

$$\text{recall}_i(t) = \frac{\sum_f \mathbb{1}[f \in P_i(t) \cap T_i]}{\sum_f \mathbb{1}[f \in T_i]}, \tag{8}$$

where f is a function term in the ontology, T_i is a set of experimentally determined function terms for protein i, $P_i(t)$ denotes the set of predicted terms for protein i with scores greater than or equal to t and $\mathbb{1}[\cdot] \in \{0, 1\}$ is an indicator function that is equal to 1 *iff* the condition is true.

Then, the average precision and recall over all proteins at threshold t is defined as:

$$\text{precision}(t) = \frac{1}{M(t)} \sum_i \text{precision}_i(t), \tag{9}$$

and

$$\text{recall}(t) = \frac{1}{N} \sum_i \text{recall}_i(t), \tag{10}$$

where we use N to denote the number of proteins and $M(t)$ to denote the number of proteins on which at least one prediction was made above threshold t, *i.e.*, $|P_i(t)| > 0$.

Combining these two measures, the maximum F-score is defined as the maximum value of F-measure over all thresholds. That is,

$$\text{F}_{\text{max}} = \max_t \left\{ \frac{2 \cdot \text{precision}(t) \cdot \text{recall}(t)}{\text{precision}(t) + \text{recall}(t)} \right\}. \tag{11}$$

The second metric, pair-centric area under precision-recall curve $\text{AUPR}_{\text{pair}}$, is defined as the average precision scores for all protein-function pairs, which is exactly the micro average precision score for multiple binary classification.

E.3 IMPLEMENTATION DETAILS

In this subsection, we describe implementation details of all baselines and our methods. For all models, the outputs will be fed into a three-layer MLP to make final prediction. The dimension of hidden layers in the MLP is equal to the dimension of model outputs.

CNN (Shanehsazzadeh et al., 2020). Following the finding in Shanehsazzadeh et al. (2020), we employ a convolutional neural network (CNN) to encode protein sequences. Specifically, 2 convolutional layers with 1024 hidden dimensions and kernel size 5 constitute this baseline model.

ResNet (Rao et al., 2019). We also adopt a deep CNN model, *i.e.*, the ResNet for protein sequences proposed by Rao et al. (2019), in our benchmark. This model is with 12 residual blocks and 512 hidden dimensions, and it uses the GELU (Hendrycks & Gimpel, 2016) activation function.

LSTM (Rao et al., 2019). The bidirectional LSTM model proposed by Rao et al. (2019) is another baseline for protein sequence encoding. It is composed of three bidirectional LSTM layers with 640 hidden dimensions.

Transformer (Rao et al., 2019). The self-attention-based Transformer encoder (Vaswani et al., 2017) is a strong model in natural language processing (NLP), Rao et al. (2019) adapts this model into the field of protein sequence modeling. We also adopt it as one of our baselines. This model has a comparable size with BERT-Small (Devlin et al., 2018), which contains 4 Transformer blocks with 512 hidden dimensions and 8 attention heads, with activation GELU (Hendrycks & Gimpel, 2016).

GCN (Kipf & Welling, 2017). We take GCN as a baseline to encode the residue graph derived by our graph construction scheme. We adopt the implementation in TorchDrug (Zhu et al., 2022), where 6 GCN layers with the hidden dimension of 512 are used. We run the results of GCN on EC and GO by ourselves and take its results on Fold and Reaction classification from Hermosilla et al. (2021).

GAT (Veličković et al., 2018). We adopt another popular graph neural network, GAT, as a structure-based baseline. We follow the implementation in TorchDrug and use 6 GAT layers with the hidden dimension of 512 and 1 attention head per layer for encoding. The results on EC, Fold and Reaction classification are based on our runs, and the results on GO are taken from Wang et al. (2022b).

GVP (Jing et al., 2021). The GVP model (Jing et al., 2021) is a decent protein structure encoder. It iteratively updates the scalar and vector representations of a protein, and these representations possess the merit of invariance and equivariance. In our benchmark, we evaluate this baseline method following the official source code. In specific, 3 GVP layers with 32 feature dimensions (20 scalar and 4 vector channels) constitute the GVP model.

3DCNN_MQA (Derevyanko et al., 2018). We implement the 3DCNN model from the paper (Derevyanko et al., 2018) with a box width of 40.0 and input resolution of $120 \times 120 \times 120$. The model has 6 residual blocks and 128 hidden dimensions with ELU activation function.

GraphQA (Baldassarre et al., 2021). Following hyperparameters in the original paper, we construct residue graphs based on bond and spatial information and re-implement the graph neural network. The best model has 4 layers with 128 node features, 32 edge features and 512 global features.

New IEConv (Hermosilla & Ropinski, 2022). Since the code for New IEConv has not been made public when the paper is written, we reproduce the method according to the description in the paper and achieve similar results on Fold and Reaction classification tasks. Then, we evaluate the method on EC and GO prediction tasks with the default hyperparameters reported in the original paper and follow the standard training procedure on these two tasks.

DeepFRI (Gligorijević et al., 2021). We also evaluate DeepFRI (Gligorijević et al., 2021) in our benchmark, which is a popular structure-based encoder for protein function prediction. DeepFRI employs an LSTM model to extract residue features and further constructs a residue graph to propagate messages among residues, in which a 3-layer graph convolutional network (GCN) (Kipf & Welling, 2017) is used. We directly utilize the official model checkpoint for baseline evaluation.

ESM-1b (Rives et al., 2021). Besides the from-scratch sequence encoders above, we also compare with two state-of-the-art pretrained protein language models. ESM-1b (Rives et al., 2021) is a huge Transformer encoder model whose size is larger than BERT-Large (Devlin et al., 2018), and it is pretrained on 24 million protein sequences from UniRef50 (Suzek et al., 2007) by masked language modeling (MLM) (Devlin et al., 2018). In our evaluation, we finetune the ESM-1b model with the learning rate that is one-tenth of that of the MLP prediction head.

ProtBERT-BFD (Elnaggar et al., 2021). The other protein language model evaluated in our benchmark is ProtBERT-BFD (Elnaggar et al., 2021) whose size also excesses BERT-Large (Devlin et al., 2018). This model is pretrained on 2.1 billion protein sequences from BFD (Steinegger & Söding, 2018) by MLM (Devlin et al., 2018). The evaluation of ProtBERT-BFD uses the same learning rate configuration as ESM-1b.

LM-GVP (Wang et al., 2022b). To further enhance the effectiveness of GVP (Jing et al., 2021), Wang et al. (2022b) proposed to prepend a protein language model, *i.e.* ProtBERT (Elnaggar et al.,

Table 5: Hyperparameter configurations of our model on different datasets. The batch size reported in the table refers to the batch size on each GPU. All the hyperparameters are chosen by the performance on the validation set.

Hyperparameter		EC	GO	Fold	Reaction
GNN	#layer	6	6	6	6
	hidden dim.	512	512	512	512
	dropout	0.1	0.1	0.2	0.2
Learning	optimizer	Adam	Adam	SGD	SGD
	learning rate	1e-4	1e-4	1e-3	1e-3
	weight decay	0	0	5e-4	5e-4
	batch size	2	2	2	2
	# epoch	200	200	300	300

Table 6: F_{max} on EC and GO tasks under different sequence cutoffs (30% / 40% / 50% / 70% / 95%).

Method	EC	GO-BP	GO-MF	GO-CC
CNN	0.366 / 0.361 / 0.372 / 0.429 / 0.545	0.197 / 0.195 / 0.197 / 0.211 / 0.244	0.238 / 0.243 / 0.256 / 0.292 / 0.354	0.258 / 0.257 / 0.260 / 0.263 / 0.387
ResNet	0.409 / 0.412 / 0.450 / 0.526 / 0.605	0.230 / 0.230 / 0.234 / 0.249 / 0.280	0.282 / 0.288 / 0.308 / 0.347 / 0.405	0.277 / 0.273 / 0.280 / 0.278 / 0.304
LSTM	0.247 / 0.249 / 0.270 / 0.333 / 0.425	0.194 / 0.192 / 0.195 / 0.205 / 0.225	0.223 / 0.229 / 0.245 / 0.276 / 0.321	0.263 / 0.264 / 0.269 / 0.270 / 0.283
Transformer	0.167 / 0.173 / 0.175 / 0.197 / 0.238	0.267 / 0.265 / 0.262 / 0.262 / 0.264	0.184 / 0.187 / 0.195 / 0.204 / 0.211	0.378 / 0.382 / 0.388 / 0.395 / 0.405
GCN	0.245 / 0.246 / 0.246 / 0.280 / 0.320	0.251 / 0.250 / 0.248 / 0.248 / 0.252	0.180 / 0.183 / 0.187 / 0.194 / 0.195	0.318 / 0.318 / 0.320 / 0.323 / 0.329
GearNet	0.557 / 0.570 / 0.615 / 0.693 / 0.730	0.309 / 0.309 / 0.315 / 0.336 / 0.356	0.382 / 0.397 / 0.425 / 0.474 / 0.503	0.381 / 0.385 / 0.393 / 0.398 / 0.414
GearNet-edge	**0.625 / 0.646 / 0.694 / 0.757 / 0.810**	**0.345 / 0.347 / 0.354 / 0.378 / 0.403**	**0.444 / 0.461 / 0.490 / 0.537 / 0.580**	**0.394 / 0.394 / 0.401 / 0.408 / 0.450**
DeepFRI	0.470 / 0.505 / 0.545 / 0.600 / 0.631	0.361 / 0.362 / 0.371 / 0.391 / 0.399	0.374 / 0.383 / 0.409 / 0.446 / 0.465	0.440 / 0.441 / 0.444 / 0.451 / 0.460
ESM-1b	0.737 / 0.764 / 0.797 / 0.839 / 0.864	0.394 / 0.399 / 0.407 / 0.429 / 0.452	**0.546 / 0.562 / 0.588 / 0.625 / 0.657**	**0.462 / 0.465 / 0.468 / 0.465 / 0.477**
Multiview Contrast	**0.744 / 0.769 / 0.808 / 0.848 / 0.874**	**0.436 / 0.442 / 0.449 / 0.471 / 0.490**	0.533 / 0.548 / 0.573 / 0.612 / **0.654**	0.459 / 0.460 / **0.467 / 0.469 / 0.488**

2021), before GVP to additionally utilize protein sequence representations. We also adopt this hybrid model as one of our baselines, and its implementation follows the official source code.

Our methods. For pretraining, we use Adam optimizer with learning rate 0.001 and train a model for 50 epochs. Then, the pretrained model will be finetuned on downstream datasets.

For **Multiview Contrast**, we set the cropping length of subsequence operation as 50, the radius of subspace operation as 15Å, the mask rate of random edge masking operation as 0.15. The temperature τ in the InfoNCE loss function is set as 0.07. When pretraining GearNet-Edge and GearNet-Edge-IEConv, we use 96 and 24 as batch sizes, respectively.

For **Distance Prediction**, we set the number of sampled residue pairs as 256. The batch size will be set as 128 and 32 for GearNet-Edge and GearNet-Edge-IEConv, respectively. For **Residue Type**, **Angle** and **Dihedral Prediction**, we set the number of sampled residues, residue triplets and residue quadrants as 512. The batch size will be set as 96 and 32 for GearNet-Edge and GearNet-Edge-IEConv, respectively.

For downstream evaluation, the hidden representations in each layer of GearNet will be concatenated for the final prediction. Table 5 lists the hyperparameter configurations for different downstream tasks. For the four tasks, we use the same optimizer and number of epochs as in the original papers to make fair comparison. For EC and GO prediction, we use ReduceLROnPlateau scheduler with factor 0.6 and patience 5, while we use StepLR scheduler with step size 50 and gamma 0.5 for fold and reaction classification.

F Additional Experimental Results on EC and GO Prediction

Results under different sequence identity cutoffs. Besides the experiments in Section 5, where 95% is used as the sequence identity cutoff for EC and GO dataset splitting, we also test our models and several important baselines under four lower sequence identity cutoffs and show the experimental results in Table 6. The aim of this experiment is to test the robustness of different models under different hold-out test sets, with lowering cutoff indicating lower similarity between training and test sets. It can be observed that, at lower cutoffs, our model can still achieve the best performance among models without pretraining and get comparable or better results against ESM-1b after pretraining.

Table 7: AUPR on EC and GO prediction. [†] denotes results taken from Wang et al. (2022b). For pretraining, we select the model with the best performance when training from scratch, *i.e.*, GearNet-Edge. We omit the model name and use pretraining methods to name our pretrained models.

	Method	Pretraining Dataset (Size)	EC	GO		
				BP	MF	CC
w/o pretraining	CNN (Shanehsazzadeh et al., 2020)	-	0.526	0.159	0.351	0.204
	ResNet (Rao et al., 2019)	-	0.590	0.205	0.434	0.214
	LSTM (Rao et al., 2019)	-	0.414	0.156	0.334	0.192
	Transformer (Rao et al., 2019)	-	0.218	0.156	0.177	0.210
	GCN (Kipf & Welling, 2017)	-	0.319	0.136	0.147	0.175
	GAT (Veličković et al., 2018)	-	0.320	0.171[†]	0.329[†]	0.249[†]
	GVP (Jing et al., 2021)	-	0.482	0.224[†]	0.458[†]	0.279[†]
	3DCNN_MQA (Derevyanko et al., 2018)	-	0.029	0.132	0.075	0.144
	GraphQA (Baldassarre et al., 2021)	-	0.543	0.199	0.347	0.265
	New IEConv (Hermosilla & Ropinski, 2022)	-	0.775	**0.273**	**0.572**	**0.316**
	GearNet	-	0.751	0.211	0.490	0.276
	GearNet-IEConv	-	0.835	0.231	0.547	0.259
	GearNet-Edge	-	0.835	0.251	**0.570**	0.303
	GearNet-Edge-IEConv	-	**0.843**	0.244	0.561	0.284
w/ pretraining	DeepFRI (Gligorijević et al., 2021)	Pfam (10M)	0.547	0.282	0.462	0.363
	ESM-1b (Rives et al., 2021)	UniRef50 (24M)	0.889	**0.332**	**0.639**	0.324
	ProtBERT-BFD (Elnaggar et al., 2021)	BFD (2.1B)	0.859	0.188[†]	0.464[†]	0.234[†]
	LM-GVP (Wang et al., 2022b)	UniRef100 (216M)	0.710	0.302[†]	0.580[†]	**0.423**[†]
	Residue Type Prediction	AlphaFoldDB (805K)	0.870	0.267	0.583	0.311
	Distance Prediction	AlphaFoldDB (805K)	0.863	0.274	0.586	0.327
	Angle Prediction	AlphaFoldDB (805K)	0.880	0.291	0.603	0.331
	Dihedral Prediction	AlphaFoldDB (805K)	0.881	0.304	0.603	0.338
	Multiview Contrast	AlphaFoldDB (805K)	**0.892**	0.292	0.596	0.336

AUPR on EC and GO prediction. We have reported experimental results on EC and GO prediction with F_{max} as the metric in Section 5. Here we report another popular metric AUPR in Table 7. Note that we still use the best model selected by F_{max} on validation sets. It can be observed that our model can still achieve the best performance on EC prediction in both from scratch and pretrained settings. However, there are still non-trivial gaps between our models with the state-of-the-art results. This probably is because of the inconsistency between the two evaluation metrics. It would be interesting to study the relationship between these two metrics and develop a model good at both in future works.

Sampling schemes in self-prediction methods. Different sampling schemes may lead to different results for self-prediction methods. We study the effects of sampling schemes using Dihedral Prediction as an example. Instead of sampling dihedral angles formed by three consecutive edges, we try to predict the dihedrals formed by four randomly sampled nodes. We observe that the F_{max} decreases from 0.859 to 0.821. This suggests that it is better of learning residue representations to capture local spatial information instead of global information. The change of sampling schemes will make self-prediction tasks more difficult to solve, which even brings negative effects after pretraining.

Pretraining on different datasets. We use the AlphaFold protein structure database as our pretraining database, because it contains the largest number of protein structures and is planned to cover over 100 million proteins. However, the structures in this database are not experimentally determined but predicted by AlphaFold2. Therefore, it is interesting to see the performance of our methods when pretraining on different datasets.

To study the effects of the choice of pretraining dataset, we build another dataset using structures extracted from Protein Data Bank (PDB) (Berman et al., 2000). Specifically, we extract 123,505 experimentally-determined protein structures from PDB whose resolutions are between 0.0 and 2.5 angstroms, and we further extract 305,265 chains from these proteins to construct the final dataset.

Next, we pretrain our five methods on AlphaFold Database v1 (proteome-wide structure predictions), AlphaFold Database v2 (Swiss-Prot structure predictions) and Protein Data Bank and then evaluate the pretrained models on the EC prediction task. The results are reported in Table 8. As can be seen in the table, our methods can achieve comparable performance on different pretraining datasets. Consequently, our methods are robust to the choice of pretraining datasets.

Table 8: Results of GearNet-Edge pretrained on different pretraining datasets with different methods. Models are evaluated on the EC prediction task.

Dataset	# Proteins	Multiview Contrast		Residue Type Prediction		Distance Prediction		Angle Prediction		Dihedral Prediction	
		$AUPR_{pair}$	F_{max}	$AUPR_{pair}$	F_{max}	$AUPR_{pair}$	F_{max}	$AUPR_{pair}$	F_{max}	$AUPR_{pair}$	F_{max}
AlphaFold Database (v1 + v2)	804,872	0.892	0.874	0.870	0.834	0.863	0.839	0.880	0.853	0.881	0.859
AlphaFold Database (v1)	365,198	0.890	0.874	0.869	0.842	0.871	0.843	0.879	0.854	0.877	0.852
AlphaFold Database (v2)	439,674	0.890	0.874	0.868	0.838	0.868	0.846	0.881	0.853	0.883	0.861
Protein Data Bank	305,265	0.881	0.859	0.870	0.841	0.865	0.847	0.880	0.857	0.886	0.858

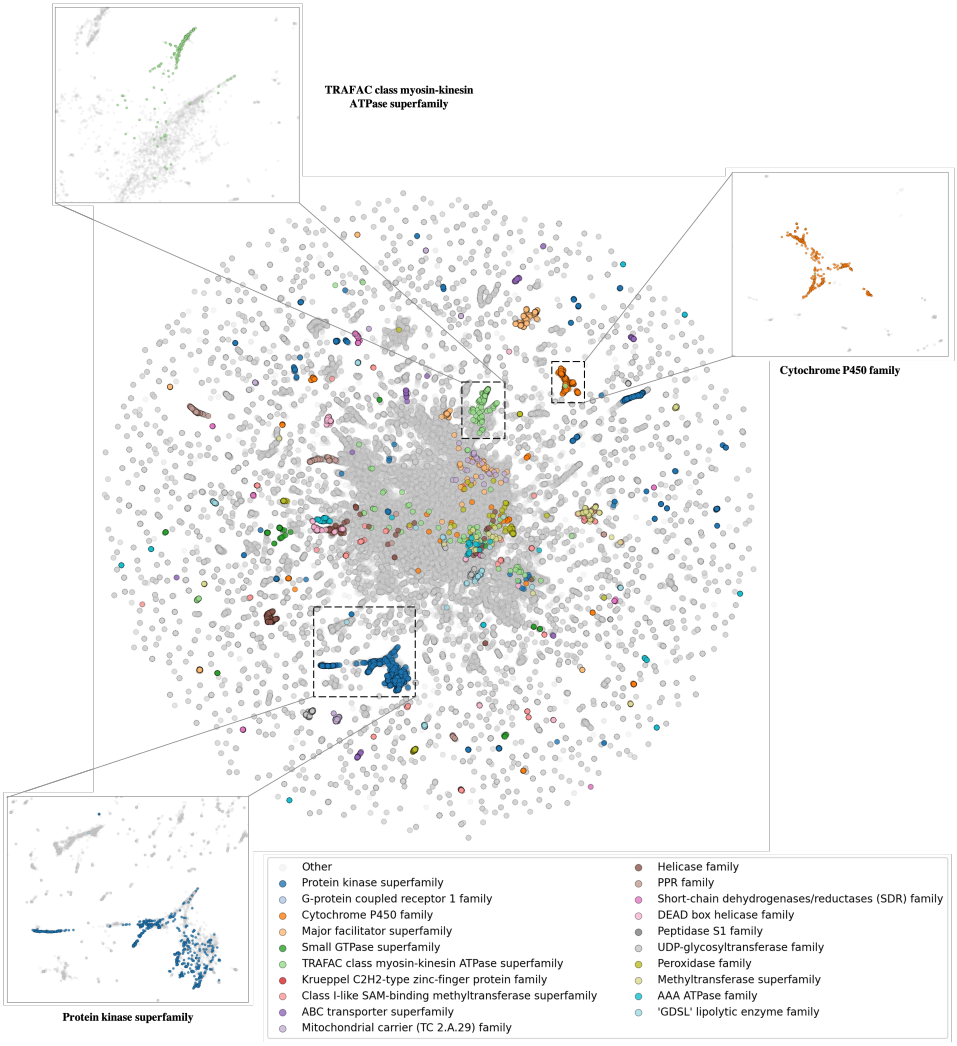

Figure 6: Latent space visualization of GearNet-Edge (Multiview Contrast) on AlphaFold Database v1.

G Latent Space Visualization

For qualitatively evaluating the quality of the protein embeddings learned by our pretraining method, we visualize the latent space of the GearNet-Edge model pretrained by Multiview Contrast. Specifically, we utilize the pretrained model to extract the embeddings of all the proteins in AlphaFold Database v1, and these embeddings are mapped to the two-dimensional space by UMAP (McInnes et al., 2018) for visualization. Following Akdel et al. (2021), we highlight the 20 most common superfamilies within the database by different colors. The visualization results are shown in Fig. 6. It can be observed that our pretrained model tends to group the proteins from the same superfamily together and divide the ones from different superfamilies apart. In particular, it succeeds in clearly separating

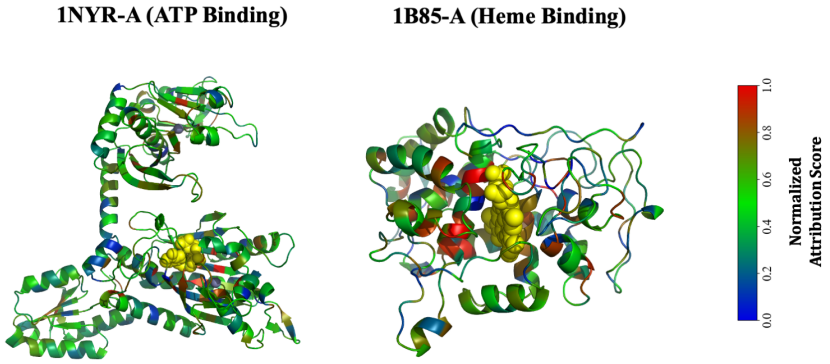

Figure 7: Identification of active sites on proteins responsible for binding based on attribution scores. Two proteins binding to specific targets are selected for illustration (1NYR-A for ATP binding and 1B85-A for Heme binding). For these two complexes, ligands are shown in yellow spheres while the residues of the receptors are colored based on attribution scores. Residues with higher attribution scores are colored in red while those with lower scores are colored in blue.

three superfamilies, *i.e.*, Protein kinase superfamily, Cytochrome P450 family and TRAFAC class myosin-kinesin ATPase superfamily. Such a decent capability of discriminating protein superfamilies, to some degree, interprets our model's superior performance on Fold Classification.

H Residue-Level Explanation

Protein functions are often reflected by specific regions on the 3D protein structures. For example, the binding ability of a protein to a ligand is highly related to the binding interface between them. Hence, to better interpret our prediction, we apply Integrated Gradients (IG) (Sundararajan et al., 2017), a model-agnostic attribution method, on our model to obtain residue-level interpretation. Specifically, we first select two molecular functions, ATP binding (GO:0005524) and Heme binding (GO:0020037), from GO terms that are related to ligand binding. For each functional term, we pick one protein and feed it into the best model trained on the GO-MF dataset. Then, we use IG to generate the feature attribution scores for each protein. The method will integrate the gradient along a straight-line path between a baseline input and the original input. Here the original input and baseline input are the node feature $\boldsymbol{f}$ and a zero vector, respectively. The final attribution score for each protein will be obtained by summing over the feature dimension. The normalized score distribution over all residues are visualized in Figure 7. As can be seen, our model is able to identify the active sites around the ligand, which are likely to be responsible for binding. Note that these attributions are directly generated from our model without any supervision, which suggests the decent interpretability of our model.